

# Package ‘ASURAT’

July 13, 2025

**Type** Package

**Title** Functional annotation-driven unsupervised clustering for  
single-cell data

**Version** 1.12.0

**Description** ASURAT is a software for single-cell data analysis.  
Using ASURAT, one can simultaneously perform unsupervised clustering and  
biological interpretation in terms of cell type, disease, biological process,  
and signaling pathway activity. Inputting a single-cell RNA-seq data and  
knowledge-based databases, such as Cell Ontology, Gene Ontology, KEGG, etc.,  
ASURAT transforms gene expression tables into original multivariate tables,  
termed sign-by-sample matrices (SSMs).

**License** GPL-3 + file LICENSE

**biocViews** GeneExpression, SingleCell, Sequencing, Clustering,  
GeneSignaling

**VignetteBuilder** knitr

**Encoding** UTF-8

**LazyData** TRUE

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grid, grDevices, graphics

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---

|              |                                               |
|--------------|-----------------------------------------------|
| add_metadata | <i>Add metadata of variables and samples.</i> |
|--------------|-----------------------------------------------|

---

Description

This function adds metadata of variables and samples.

Usage

```
add_metadata(sce = NULL, mitochondria_symbol = NULL)
```

**Arguments**

`sce` A SingleCellExperiment object.

`mitochondria_symbol` A string representing for mitochondrial genes. This function computes percents of reads that map to the mitochondrial genes. Examples are ‘^MT-’, ‘^mt-’, etc.

**Value**

A SingleCellExperiment object.

**Examples**

```
data(pbmc_eg)
pbmc <- add_metadata(sce = pbmc_eg, mitochondria_symbol = "^MT-")
```

---

|        |                                                                                 |
|--------|---------------------------------------------------------------------------------|
| ASURAT | <i>Functional annotation-driven unsupervised clustering of SingleCell data.</i> |
|--------|---------------------------------------------------------------------------------|

---

**Description**

ASURAT is a software for single-cell data analysis. Using ASURAT, one can simultaneously perform unsupervised clustering and biological interpretation in terms of cell type, disease, biological process, and signaling pathway activity. Inputting a single-cell RNA-seq data and knowledge-based databases, such as Cell Ontology, Gene Ontology, KEGG, etc., ASURAT transforms gene expression tables into original multivariate tables, termed sign-by-sample matrices (SSMs).

---

|             |                                                              |
|-------------|--------------------------------------------------------------|
| bubble_sort | <i>Perform bubble sorting, counting the number of steps.</i> |
|-------------|--------------------------------------------------------------|

---

**Description**

Perform bubble sorting, counting the number of steps.

**Usage**

```
bubble_sort(listdata)
```

**Arguments**

`listdata` A list of vector and integer. For example, in R code, `listdata = list(vec = c(1, 0, 1, ...), cnt = 0)`. The integer (`cnt = 0`) is the initial number of steps for bubble sorting.

**Value**

A List.

**Examples**

```
bubble_sort(list(vec = c(1, 1, 0), cnt = 0))
```

---

|                  |                                                            |
|------------------|------------------------------------------------------------|
| cluster_genesets | <i>Cluster each functional gene set into three groups.</i> |
|------------------|------------------------------------------------------------|

---

**Description**

This function clusters each functional gene set into strongly, variably, and weakly correlated gene sets.

**Usage**

```
cluster_genesets(sce = NULL, cormat = NULL, th_posi = NULL, th_neg = NULL)
```

**Arguments**

|         |                                                  |
|---------|--------------------------------------------------|
| sce     | A SingleCellExperiment object.                   |
| cormat  | A correlation matrix of gene expressions.        |
| th_posi | A threshold of positive correlation coefficient. |
| th_neg  | A threshold of negative correlation coefficient. |

**Value**

A SingleCellExperiment object.

**Examples**

```
data(pbmc_eg)
data(human_GO_eg)
mat <- t(as.matrix(SummarizedExperiment::assay(pbmc_eg, "centered")))
pbmc_cormat <- cor(mat, method = "spearman")
pbmcs <- list(GO = pbmc_eg)
S4Vectors::metadata(pbmcs$GO) <- list(sign = human_GO_eg[["BP"]])
pbmcs$GO <- remove_signs(sce = pbmcs$GO, min_ngenes = 2, max_ngenes = 1000)
pbmcs$GO <- cluster_genesets(sce = pbmcs$GO, cormat = pbmc_cormat,
                             th_posi = 0.24, th_neg = -0.20)
# The results are stored in `metadata(pbmcs$GO)$sign`.
```

---

|                  |                                                                        |
|------------------|------------------------------------------------------------------------|
| compute_sepI_all | <i>Compute separation indices for each cluster against the others.</i> |
|------------------|------------------------------------------------------------------------|

---

**Description**

This function computes separation indices for each cluster versus the others.

**Usage**

```
compute_sepI_all(sce = NULL, labels = NULL, nrand_samples = NULL)
```

**Arguments**

|               |                                                                                                               |
|---------------|---------------------------------------------------------------------------------------------------------------|
| sce           | A SingleCellExperiment object.                                                                                |
| labels        | A vector of labels of all the samples (cells).                                                                |
| nrand_samples | An integer for the number of samples used for random sampling, which samples at least one sample per cluster. |

**Value**

A SingleCellExperiment object.

**Examples**

```
data(pbmcs_eg)
labels <- SummarizedExperiment::colData(pbmcs_eg$G0)$seurat_clusters
pbmcs_eg$G0 <- compute_sepI_all(sce = pbmcs_eg$G0, labels = labels,
                               nrand_samples = 10)
# The results are stored in `metadata(pbmcs_eg$G0)$marker_signs`.
```

---

|                       |                                                                          |
|-----------------------|--------------------------------------------------------------------------|
| compute_sepI_clusters | <i>Compute separation indices of sign scores for given two clusters.</i> |
|-----------------------|--------------------------------------------------------------------------|

---

**Description**

This function computes separation indices of sign scores for given two clusters.

**Usage**

```
compute_sepI_clusters(
  sce = NULL,
  labels = NULL,
  nrand_samples = NULL,
  ident_1 = NULL,
  ident_2 = NULL
)
```

**Arguments**

|               |                                                                                                               |
|---------------|---------------------------------------------------------------------------------------------------------------|
| sce           | A SingleCellExperiment object.                                                                                |
| labels        | A vector of labels of all the samples.                                                                        |
| nrand_samples | An integer for the number of samples used for random sampling, which samples at least one sample per cluster. |
| ident_1       | Label names identifying cluster numbers, e.g., ident_1 = 1, ident_1 = c(1, 3).                                |
| ident_2       | Label names identifying cluster numbers, e.g., ident_2 = 2, ident_2 = c(2, 4).                                |

**Value**

A SingleCellExperiment object.

**Examples**

```
data(pbmcs_eg)
labels <- SummarizedExperiment::colData(pbmcs_eg$G0)$seurat_clusters
pbmcs_eg$G0 <- compute_sepI_clusters(sce = pbmcs_eg$G0, labels = labels,
                                   nrand_samples = 10, ident_1 = 1,
                                   ident_2 = c(0, 2))
# The results are stored in `metadata(pbmcs_eg$G0)$marker_signs`.
```

---

|              |                                                                     |
|--------------|---------------------------------------------------------------------|
| create_signs | <i>Define signs for strongly and variably correlated gene sets.</i> |
|--------------|---------------------------------------------------------------------|

---

**Description**

This function define signs for strongly and variably correlated gene sets.

**Usage**

```
create_signs(sce = NULL, min_cnt_strg = 2, min_cnt_vari = 2)
```

**Arguments**

|              |                                                                    |
|--------------|--------------------------------------------------------------------|
| sce          | A SingleCellExperiment object.                                     |
| min_cnt_strg | An integer for the cutoff value for strongly correlated gene sets. |
| min_cnt_vari | An integer for the cutoff value for variably correlated gene sets. |

**Value**

A SingleCellExperiment object.

Examples

```
data(pbmc_eg)
data(human_GO_eg)
mat <- t(as.matrix(SummarizedExperiment::assay(pbmc_eg, "centered")))
pbmc_cormat <- cor(mat, method = "spearman")
pbmcs <- list(GO = pbmc_eg)
S4Vectors::metadata(pbmcs$GO) <- list(sign = human_GO_eg[["BP"]])
pbmcs$GO <- remove_signs(sce = pbmcs$GO, min_ngenes = 2, max_ngenes = 1000)
pbmcs$GO <- cluster_genesets(sce = pbmcs$GO, cormat = pbmc_cormat,
                             th_posi = 0.24, th_negs = -0.20)
pbmcs$GO <- create_signs(sce = pbmcs$GO, min_cnt_strg = 2, min_cnt_vari = 2)
# The results are stored in `metadata(pbmcs$GO)$sign_all`.
```

---

|                 |                                                                      |
|-----------------|----------------------------------------------------------------------|
| human_COMSig_eg | <i>A list of small Cell Ontology and MSigDB databases for human.</i> |
|-----------------|----------------------------------------------------------------------|

---

Description

A list of small Cell Ontology and MSigDB databases for human.

Usage

```
human_COMSig_eg
```

Format

A list of dataframe.

---

|             |                                                          |
|-------------|----------------------------------------------------------|
| human_GO_eg | <i>A list of small Gene Ontology database for human.</i> |
|-------------|----------------------------------------------------------|

---

Description

A list of small Gene Ontology database for human.

Usage

```
human_GO_eg
```

Format

A list of dataframe.

---

|               |                                                 |
|---------------|-------------------------------------------------|
| human_KEGG_eg | <i>A list of small KEGG database for human.</i> |
|---------------|-------------------------------------------------|

---

### Description

A list of small KEGG database for human.

### Usage

```
human_KEGG_eg
```

### Format

A list of dataframe.

---

|                |                                                                              |
|----------------|------------------------------------------------------------------------------|
| makeSignMatrix | <i>Create a new SingleCellExperiment object for sign-by-sample matrices.</i> |
|----------------|------------------------------------------------------------------------------|

---

### Description

This function creates a new SingleCellExperiment object for sign-by-sample matrices (SSM) by concatenating SSMs for strongly and variably correlated gene sets.

### Usage

```
makeSignMatrix(sce = NULL, weight_strg = 0.5, weight_vari = 0.5)
```

### Arguments

|             |                                                       |
|-------------|-------------------------------------------------------|
| sce         | A SingleCellExperiment object.                        |
| weight_strg | A weight parameter for strongly correlated gene sets. |
| weight_vari | A weight parameter for variably correlated gene sets. |

### Value

A SingleCellExperiment object.

**Examples**

```

data(pbmcs_eg)
data(human_GO_eg)
mat <- t(as.matrix(SummarizedExperiment::assay(pbmcs_eg, "centered")))
pbmc_cormat <- cor(mat, method = "spearman")
pbmcs <- list(GO = pbmc_eg)
S4Vectors::metadata(pbmcs$GO) <- list(sign = human_GO_eg[["BP"]])
pbmcs$GO <- remove_signs(sce = pbmcs$GO, min_ngenes = 2, max_ngenes = 1000)
pbmcs$GO <- cluster_genesets(sce = pbmcs$GO, cormat = pbmc_cormat,
                           th_posi = 0.24, th_negs = -0.20)
pbmcs$GO <- create_signs(sce = pbmcs$GO, min_cnt_strg = 2, min_cnt_vari = 2)
pbmcs$GO <- makeSignMatrix(sce = pbmcs$GO, weight_strg = 0.5,
                          weight_vari = 0.5)
# The results can be checked by, e.g., assay(pbmcs$GO, "counts").

```

---

|          |                                                                               |
|----------|-------------------------------------------------------------------------------|
| pbmcs_eg | <i>A list of SingleCellExperiment objects made from sign-sample matrices.</i> |
|----------|-------------------------------------------------------------------------------|

---

**Description**

A list of SingleCellExperiment objects, consisting of small sign-by-sample matrices, pbmcs\_eg\$CM (using Cell Ontology and MSigDB databases), pbmcs\_eg\$GO (using Gene Ontology database), and pbmcs\_eg\$KG (KEGG). Here, pbmcs\_eg\$CM, pbmcs\_eg\$GO, and pbmcs\_eg\$KG include 87, 72, and 64 signs, respectively, and 50 cells.

**Usage**

```
pbmcs_eg
```

**Format**

A list of SingleCellExperiment objects.

---

|         |                                                                         |
|---------|-------------------------------------------------------------------------|
| pbmc_eg | <i>A SingleCellExperiment object made from a gene expression table.</i> |
|---------|-------------------------------------------------------------------------|

---

**Description**

A SingleCellExperiment object, including 50 genes and 50 cells. The original data "4k PBMCs from a Healthy Donor" was downloaded from 10x Genomics database.

**Usage**

```
pbmc_eg
```

**Format**

SingleCellExperiment object.

**Source**

<https://support.10xgenomics.com/single-cell-gene-expression>

---

|                  |                                                                   |
|------------------|-------------------------------------------------------------------|
| plot_dataframe3D | <i>Visualize a three-dimensional data with labels and colors.</i> |
|------------------|-------------------------------------------------------------------|

---

**Description**

This function visualizes a three-dimensional data with labels and colors.

**Usage**

```
plot_dataframe3D(  
  dataframe3D = NULL,  
  labels = NULL,  
  colors = NULL,  
  theta = 30,  
  phi = 30,  
  title = "",  
  xlabel = "",  
  ylabel = "",  
  zlabel = ""  
)
```

**Arguments**

|             |                                                                         |
|-------------|-------------------------------------------------------------------------|
| dataframe3D | A dataframe with three columns.                                         |
| labels      | NULL or a vector of labels of all the samples, corresponding to colors. |
| colors      | NULL or a vector of colors of all the samples, corresponding to labels. |
| theta       | Angle of the plot.                                                      |
| phi         | Angle of the plot.                                                      |
| title       | Title.                                                                  |
| xlabel      | x-axis label.                                                           |
| ylabel      | y-axis label.                                                           |
| zlabel      | z-axis label.                                                           |

**Value**

A scatter3D object in plot3D package.

**Examples**

```

data(pbmcs_eg)
mat <- SingleCellExperiment::reducedDim(pbmcs_eg$CM, "UMAP")[, 1:3]
dataframe3D <- as.data.frame(mat)
labels <- SummarizedExperiment::colData(pbmcs_eg$CM)$seurat_clusters
plot_dataframe3D(dataframe3D = dataframe3D, labels = labels, colors = NULL,
                 theta = 45, phi = 20, title = "PBMC (CO & MSigDB)",
                 xlabel = "UMAP_1", ylabel = "UMAP_2", zlabel = "UMAP_3")

```

---

|                    |                                                 |
|--------------------|-------------------------------------------------|
| plot_multiheatmaps | <i>Visualize multivariate data by heatmaps.</i> |
|--------------------|-------------------------------------------------|

---

**Description**

This function visualizes multivariate data by heatmaps.

**Usage**

```

plot_multiheatmaps(
  ssm_list = NULL,
  gem_list = NULL,
  ssmlabel_list = NULL,
  gemlabel_list = NULL,
  nrand_samples = NULL,
  show_row_names = FALSE,
  title = NULL
)

```

**Arguments**

|                |                                                                                                                                                                                                              |
|----------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| ssm_list       | A list of sign-by-sample matrices.                                                                                                                                                                           |
| gem_list       | A list of gene-by-sample matrices.                                                                                                                                                                           |
| ssmlabel_list  | NULL or a list of dataframes of sample (cell) labels and colors. The length of the list must be as same as that of ssm_list, and the order of labels in each list must be as same as those in ssm_list.      |
| gemlabel_list  | NULL or a list of dataframes of sample (cell) annotations and colors. The length of the list must be as same as that of gem_list, and the order of labels in each list must be as same as those in gem_list. |
| nrand_samples  | Number of samples (cells) used for random sampling.                                                                                                                                                          |
| show_row_names | TRUE or FALSE: if TRUE, row names are shown.                                                                                                                                                                 |
| title          | Title.                                                                                                                                                                                                       |

**Value**

A ComplexHeatmap object.

## Examples

```
data(pbmcs_eg)
mat_CM <- SummarizedExperiment::assay(pbmcs_eg$CM, "counts")
mat_GO <- SummarizedExperiment::assay(pbmcs_eg$GO, "counts")
mat_KG <- SummarizedExperiment::assay(pbmcs_eg$KG, "counts")
ssm_list <- list(SSM_COMSig = mat_CM, SSM_GO = mat_GO, SSM_KEGG = mat_KG)
se <- SingleCellExperiment::altExp(pbmcs_eg$CM, "logcounts")
mat <- SummarizedExperiment::assay(se, "counts")
se <- SingleCellExperiment::altExp(pbmcs_eg$CM, "logcounts")
gem_list <- list(GeneExpr = SummarizedExperiment::assay(se, "counts"))
labels <- list() ; ssmlabel_list <- list()
for(i in seq_along(pbmcs_eg)){
  fa <- SummarizedExperiment::colData(pbmcs_eg[[i]])$seurat_clusters
  labels[[i]] <- data.frame(label = fa)
  colors <- rainbow(length(unique(labels[[i]]$label)))[labels[[i]]$label]
  labels[[i]]$color <- colors
  ssmlabel_list[[i]] <- labels[[i]]
}
names(ssmlabel_list) <- c("Label_COMSig", "Label_GO", "Label_KEGG")
phases <- SummarizedExperiment::colData(pbmcs_eg$CM)$Phase
label_CC <- data.frame(label = phases, color = NA)
gemlabel_list <- list(CellCycle = label_CC)
plot_multiheatmaps(ssm_list = ssm_list, gem_list = gem_list,
  ssmlabel_list = ssmlabel_list,
  gemlabel_list = gemlabel_list, nrand_samples = 50,
  show_row_names = FALSE, title = "PBMC")
```

---

remove\_samples

*Remove samples based on expression profiles across variables.*

---

## Description

This function removes sample data by setting minimum and maximum threshold values for the metadata.

## Usage

```
remove_samples(
  sce = NULL,
  min_nReads = NULL,
  max_nReads = NULL,
  min_nGenes = NULL,
  max_nGenes = NULL,
  min_percMT = NULL,
  max_percMT = NULL
)
```

**Arguments**

|            |                                                                                    |
|------------|------------------------------------------------------------------------------------|
| sce        | A SingleCellExperiment object.                                                     |
| min_nReads | A minimum threshold value of the number of reads.                                  |
| max_nReads | A maximum threshold value of the number of reads.                                  |
| min_nGenes | A minimum threshold value of the number of non-zero expressed genes.               |
| max_nGenes | A maximum threshold value of the number of non-zero expressed genes.               |
| min_percMT | A minimum threshold value of the percent of reads that map to mitochondrial genes. |
| max_percMT | A maximum threshold value of the percent of reads that map to mitochondrial genes. |

**Value**

A SingleCellExperiment object.

**Examples**

```
data(pbmc_eg)
pbmc <- add_metadata(sce = pbmc_eg, mitochondria_symbol = "^MT-")
pbmc <- remove_samples(sce = pbmc, min_nReads = 0, max_nReads = 1e+10,
                      min_nGenes = 0, max_nGenes = 1e+10,
                      min_percMT = NULL, max_percMT = NULL)
```

---

|              |                                                          |
|--------------|----------------------------------------------------------|
| remove_signs | <i>Remove signs including too few or too many genes.</i> |
|--------------|----------------------------------------------------------|

---

**Description**

This function removes signs including too few or too many genes.

**Usage**

```
remove_signs(sce = NULL, min_ngenes = 2, max_ngenes = 1000)
```

**Arguments**

|            |                                                          |
|------------|----------------------------------------------------------|
| sce        | A SingleCellExperiment object.                           |
| min_ngenes | Minimum number of genes, which must be greater than one. |
| max_ngenes | Maximum number of genes, which must be greater than one. |

**Value**

A SingleCellExperiment object.

## Examples

```
data(pbmc_eg)
data(human_GO_eg)
pbmcs <- list(GO = pbmc_eg)
S4Vectors::metadata(pbmcs$GO) <- list(sign = human_GO_eg[["BP"]])
pbmcs$GO <- remove_signs(sce = pbmcs$GO, min_ngenes = 2, max_ngenes = 1000)
# The results are stored in `metadata(pbmcs$GO)$sign`.
```

---

remove\_signs\_manually *Remove signs by specifying keywords.*

---

## Description

This function removes signs by specifying keywords.

## Usage

```
remove_signs_manually(sce = NULL, keywords = NULL)
```

## Arguments

|          |                                  |
|----------|----------------------------------|
| sce      | A SingleCellExperiment object.   |
| keywords | keywords separated by pipes 'l'. |

## Value

A SingleCellExperiment object.

## Examples

```
data(pbmc_eg)
data(human_GO_eg)
mat <- t(as.matrix(SummarizedExperiment::assay(pbmc_eg, "centered")))
pbmc_cormat <- cor(mat, method = "spearman")
pbmcs <- list(GO = pbmc_eg)
S4Vectors::metadata(pbmcs$GO) <- list(sign = human_GO_eg[["BP"]])
pbmcs$GO <- remove_signs(sce = pbmcs$GO, min_ngenes = 2, max_ngenes = 1000)
pbmcs$GO <- cluster_genesets(sce = pbmcs$GO, cormat = pbmc_cormat,
                             th_posi = 0.24, th_neg = -0.20)
pbmcs$GO <- create_signs(sce = pbmcs$GO, min_cnt_strg = 2, min_cnt_vari = 2)
keywords <- "Covid19|foofoo|hogehoge"
pbmcs$GO <- remove_signs_manually(sce = pbmcs$GO, keywords = keywords)
# The results are stored in `metadata(pbmcs$GO)$sign_SCG`,
# `metadata(pbmcs$GO)$sign_VCG`, and `metadata(pbmcs$GO)$sign_all`.
```

---

remove\_signs\_redundant

*Remove redundant signs using semantic similarity matrices.*


---

## Description

This function removes redundant signs using semantic similarity matrices.

## Usage

```
remove_signs_redundant(
  sce = NULL,
  similarity_matrix = NULL,
  threshold = NULL,
  keep_rareID = NULL
)
```

## Arguments

|                   |                                                                                               |
|-------------------|-----------------------------------------------------------------------------------------------|
| sce               | A SingleCellExperiment object.                                                                |
| similarity_matrix | A semantic similarity matrix.                                                                 |
| threshold         | A threshold value of semantic similarity, used for regarding biological terms as similar ones |
| keep_rareID       | If TRUE, biological terms with the larger ICs are kept.                                       |

## Value

A SingleCellExperiment object.

## Examples

```
data(pbmc_eg)
data(human_GO_eg)
mat <- t(as.matrix(SummarizedExperiment::assay(pbmc_eg, "centered")))
pbmc_cormat <- cor(mat, method = "spearman")
pbmcs <- list(GO = pbmc_eg)
S4Vectors::metadata(pbmcs$GO) <- list(sign = human_GO_eg[["BP"]])
pbmcs$GO <- remove_signs(sce = pbmcs$GO, min_ngenes = 2, max_ngenes = 1000)
pbmcs$GO <- cluster_genesets(sce = pbmcs$GO, cormat = pbmc_cormat,
  th_posi = 0.24, th_neg = -0.20)
pbmcs$GO <- create_signs(sce = pbmcs$GO, min_cnt_strg = 2, min_cnt_vari = 2)
pbmcs$GO <- remove_signs_redundant(
  sce = pbmcs$GO, similarity_matrix = human_GO_eg$similarity_matrix$BP,
  threshold = 0.80, keep_rareID = TRUE)
# The results are stored in `metadata(pbmcs$GO)$sign_SCG`,
# `metadata(pbmcs$GO)$sign_VCG`, `metadata(pbmcs$GO)$sign_all`,
# and if there exist, `metadata(pbmcs$GO)$sign_SCG_redundant` and
```

```
# `metadata(pbmcs$GO)$sign_VCG_redundant`.
```

---

|                  |                                                                      |
|------------------|----------------------------------------------------------------------|
| remove_variables | <i>Remove variables based on expression profiles across samples.</i> |
|------------------|----------------------------------------------------------------------|

---

### Description

This function removes low expressed variable data.

### Usage

```
remove_variables(sce = NULL, min_nsamples = 0)
```

### Arguments

|              |                                                                                                                            |
|--------------|----------------------------------------------------------------------------------------------------------------------------|
| sce          | A SingleCellExperiment object.                                                                                             |
| min_nsamples | An integer. This function removes variables for which the numbers of non-zero expressing samples are less than this value. |

### Value

A SingleCellExperiment object.

### Examples

```
data(pbmcs_eg)
pbmc <- add_metadata(sce = pbmc_eg, mitochondria_symbol = "^MT-")
pbmc <- remove_variables(sce = pbmc, min_nsamples = 10)
```

---

|                         |                                                                             |
|-------------------------|-----------------------------------------------------------------------------|
| remove_variables_second | <i>Remove variables based on the mean expression levels across samples.</i> |
|-------------------------|-----------------------------------------------------------------------------|

---

### Description

This function removes variable data such that the mean expression levels across samples are less than 'min\_meannReads'.

### Usage

```
remove_variables_second(sce = NULL, min_meannReads = 0)
```

**Arguments**

sce                    A SingleCellExperiment object.

min\_meannReads    An integer. This function removes variables for which the mean read counts are less than this value.

**Value**

A SingleCellExperiment object.

**Examples**

```
data(pbmc_eg)
pbmc <- remove_variables_second(sce = pbmc_eg, min_meannReads = 0.01)
```

---

select\_signs\_manually    *Select signs by specifying keywords.*

---

**Description**

This function selects signs by specifying keywords.

**Usage**

```
select_signs_manually(sce = NULL, keywords = NULL)
```

**Arguments**

sce                    An ASURAT object.

keywords              Keywords separated by a pipe.

**Value**

An ASURAT object.

**Examples**

```
data(pbmc_eg)
data(human_GO_eg)
mat <- t(as.matrix(SummarizedExperiment::assay(pbmc_eg, "centered")))
pbmc_cormat <- cor(mat, method = "spearman")
pbmcs <- list(GO = pbmc_eg)
S4Vectors::metadata(pbmcs$GO) <- list(sign = human_GO_eg[["BP"]])
pbmcs$GO <- remove_signs(sce = pbmcs$GO, min_ngenes = 2, max_ngenes = 1000)
pbmcs$GO <- cluster_genesets(sce = pbmcs$GO, cormat = pbmc_cormat,
                             th_posi = 0.24, th_neg = -0.20)
pbmcs$GO <- create_signs(sce = pbmcs$GO, min_cnt_strg = 2, min_cnt_vari = 2)
keywords <- "cell|process"
```

```
pbmcs$G0 <- select_signs_manually(sce = pbmcs$G0, keywords = keywords)
# The results are stored in `metadata(pbmcs$G0)$sign_SCG`,
# `metadata(pbmcs$G0)$sign_VCG`, and `metadata(pbmcs$G0)$sign_all`.
```

---

**swap\_pass***Perform one-shot adjacent swapping for each element.*

---

**Description**

Perform one-shot adjacent swapping for each element.

**Usage**

```
swap_pass(listdata)
```

**Arguments**

**listdata**            A list of vector and integer.

**Value**

A List.

**Examples**

```
swap_pass(list(vec = c(1, 1, 0), cnt = 0))
```

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