

# Package ‘TrajectoryGeometry’

July 7, 2025

**Title** This Package Discovers Directionality in Time and Pseudo-times  
Series of Gene Expression Patterns

**Version** 1.17.0

**Description** Given a time series or pseudo-times series of gene expression data, we might wish to know: Do the changes in gene expression in these data exhibit directionality? Are there turning points in this directionality. Do different subsets of the data move in different directions? This package uses spherical geometry to probe these sorts of questions. In particular, if we are looking at (say) the first  $n$  dimensions of the PCA of gene expression, directionality can be detected as the clustering of points on the  $(n-1)$ -dimensional sphere.

**License** MIT + file LICENSE

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**Depends** R ( $\geq 4.1$ )

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**Author** Michael Shapiro [aut, cre] (ORCID:  
<<https://orcid.org/0000-0002-2769-9320>>)

**Maintainer** Michael Shapiro <michael.shapiro@crick.ac.uk>

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analyseBranchPoint	<i>Analyse branch point.</i>
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---

## Description

This function takes a single cell trajectory and analyses it starting from successively later points in pseudotime, with the rationale that a more consistent directionality will be followed after the branch point.

**Usage**

```
analyseBranchPoint(
  attributes,
  pseudotime,
  randomizationParams,
  statistic,
  start = (max(pseudotime) - min(pseudotime)) * 0.25,
  stop = (max(pseudotime) - min(pseudotime)) * 0.75,
  step = (max(pseudotime) - min(pseudotime)) * 0.05,
  nSamples = 1000,
  nWindows = 10,
  d = ncol(attributes),
  N = 1
)
```

**Arguments**

<code>attributes</code>	- An $n \times d$ (cell $\times$ attribute) matrix of numeric attributes for single cell data. Rownames should be cell names.
<code>pseudotime</code>	- A named numeric vector of pseudotime values for cells.
<code>randomizationParams</code>	- A character vector which is used to control the production of randomized paths for comparison.
<code>statistic</code>	- Allowable values are 'median', 'mean' or 'max'.
<code>start</code>	- The first pseudotime value (percentage of the trajectory) from which to analyse the trajectory from. Defaults to 25% of the way through the trajectory.
<code>stop</code>	- The last pseudotime value (as a percentage of the trajectory) from which to analyse the trajectory from. Defaults to 75% of the way through the trajectory.
<code>step</code>	- The size of the step to take between successively later starting points in pseudotime. Defaults to 5% of the trajectory length.
<code>nSamples</code>	- The number of sampled paths to generate (defaults to 1000).
<code>nWindows</code>	- The number of windows pseudotime should be split into to sample cells from (defaults to 10).
<code>d</code>	- The dimension under consideration. This defaults to <code>ncol(attributes)</code> .
<code>N</code>	- The number of random paths to generated for statistical comparison to the given path (defaults to 1000).

**Value**

This returns a list of results for `analyseSingleCellTrajectory`, named by trajectory starting point. Each result from `analyseSingleCellTrajectory` is a list which contains an entry for each sampled path. Each of these entries is a list containing information comparing the sampled path in question to random paths. The entries consist of: `pValue` - the p-value for the path and statistic in question; `sphericalData` - a list containing the projections of the path to the sphere, the center of that sphere and the statistic for distance to that center; `randomDistances` - the corresponding distances for randomly chosen; `paths`; `randomizationParams` - the choice of randomization parameters

## Examples

```
chol_branch_point_results = analyseBranchPoint(chol_attributes[,seq_len(3)],
  chol_pseudo_time[!is.na(chol_pseudo_time)],
  randomizationParams = c('byPermutation',
    'permuteWithinColumns'),
  statistic = "mean",
  start = 0,
  stop = 50,
  step = 5,
  nSamples = 10,
  N = 1)
```

---

analyseSingleCellTrajectory

*Analyse a single cell trajectory.*

---

## Description

This function analyses a single cell trajectory by sampling multiple paths and comparing each path to random paths. It takes vector of pseudotime values, and a matrix of attribute values (cell x attribute). It also optionally takes the number of pseudotime windows to sample a single cell from. This defaults to 10. The function returns a list of Answers for each comparison of a sampled path to a random path.

## Usage

```
analyseSingleCellTrajectory(
  attributes,
  pseudotime,
  randomizationParams,
  statistic,
  nSamples = 1000,
  nWindows = 10,
  d = ncol(attributes),
  N = 1000
)
```

## Arguments

attributes	- An n x d (cell x attribute) matrix of numeric attributes for single cell data. Rownames should be cell names.
pseudotime	- A named numeric vector of pseudotime values for cells.
randomizationParams	- A character vector which is used to control the production of randomized paths for comparison.
statistic	- Allowable values are 'median', 'mean' or 'max'.
nSamples	- The number of sampled paths to generate (default 1000).
nWindows	- The number of windows pseudotime should be split into to sample cells from (defaults to 10).
d	- The dimension under consideration. This defaults to ncol(attributes).
N	- The number of random paths to generated for statistical comparison to the given path (defaults to 1000).

**Value**

This returns a list, where each entry is itself a list containing information comparing a sampled path to random paths. These entries consist of: pValue - the p-value for the path and statistic in question; sphericalData - a list containing the projections of the path to the sphere, the center of that sphere and the statistic for distance to that center; randomDistances - the corresponding distances for randomly chosen; paths; randomizationParams - the choice of randomization parameters

**Examples**

```
chol_answers = analyseSingleCellTrajectory(chol_attributes[,seq_len(3)],
                                           chol_pseudo_time_normalised,
                                           nSamples = 10,
                                           randomizationParams =
                                             c('byPermutation',
                                               'permuteWithinColumns'),
                                           statistic = "mean",
                                           N = 1)
hep_answers = analyseSingleCellTrajectory(hep_attributes[,seq_len(3)],
                                           hep_pseudo_time_normalised,
                                           nSamples = 10,
                                           randomizationParams =
                                             c('byPermutation',
                                               'permuteWithinColumns'),
                                           statistic = "mean",
                                           N = 1)
```

---

chol\_answers

*chol\_answers*


---

**Description**

Results of running analyseSingleCellTrajectory() on a trajectory describing the development of cholangiocytes from hepatoblasts.

**Usage**

```
chol_answers
```

**Format**

A list

Results of running analyseSingleCellTrajectory() on a trajectory describing the development of cholangiocytes from hepatoblasts.

**Source**

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

chol_attributes	<i>chol_attributes</i>
-----------------	------------------------

---

### Description

PCA projections derived from normalised gene expression values for single cells, and filtered for cells which feature in a trajectory from hepatoblast to cholangiocyte. The columns are the PCs and the rows are the cells.

### Usage

```
chol_attributes
```

### Format

A matrix

PCA projections derived from normalised gene expression values for single cells, and filtered for cells which feature in a trajectory from hepatoblast to cholangiocyte. The columns are the PCs and the rows are the cells.

### Source

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

chol_branch_point_results	<i>chol_branch_point_results</i>
---------------------------	----------------------------------

---

### Description

Results of running `analyseBranchPoint()` on a trajectory describing the development of cholangiocytes from hepatoblasts.

### Usage

```
chol_branch_point_results
```

### Format

A list

Results of running `analyseBranchPoint()` on a trajectory describing the development of cholangiocytes from hepatoblasts.

### Source

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

chol_pseudo_time	<i>chol_pseudo_time</i>
------------------	-------------------------

---

**Description**

A vector of pseudotime values for a trajectory describing the development of cholangiocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.

**Usage**

```
chol_pseudo_time
```

**Format**

A vector

A vector of pseudotime values for a trajectory describing the development of cholangiocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.

**Source**

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

chol_pseudo_time_normalised	<i>chol_pseudo_time_normalised</i>
-----------------------------	------------------------------------

---

**Description**

A vector of pseudotime values, normalised to range from 0 to 100, for a trajectory describing the development of cholangiocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.

**Usage**

```
chol_pseudo_time_normalised
```

**Format**

A vector

A vector of pseudotime values, normalised to range from 0 to 100, for a trajectory describing the development of cholangiocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.

**Source**

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

circleOnTheUnitSphere    *Circle on the unit sphere*

---

### Description

Find a circle on the unit 2-sphere

### Usage

```
circleOnTheUnitSphere(center, radius, N = 36)
```

### Arguments

center	- The center of the circle.
radius	- The radius of the circle.
N	- The number of segments to approximate the circle. It defaults to 36.

### Details

Given a point on the unit 2-sphere and a radius given as a spherical distance, this finds the circle.

It's not clear to me this should be exported, but it's handy to do this for testing and debugging.

### Value

This returns an approximation to the the circle as a  $N+1 \times 3$  matrix

### Examples

```
pole = c(1,0,0)
radius = pi / 4
circle = circleOnTheUnitSphere(pole,radius)
```

---

crooked\_path    *Crooked path*

---

### Description

A path of  $n$  points in dimension  $d$  is an  $n \times d$  matrix. This particular path is relatively crooked.

### Usage

```
crooked_path
```

### Format

A  $14 \times 3$  matrix

This path changes direction after the 5th point.



**Source**

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometry>  
The second author wishes to categorically deny that this variable is named after the course of his own life.

---

crooked_path_center	<i>Crooked path center</i>
---------------------	----------------------------

---

**Description**

The point on the unit sphere minimizing mean spherical distance to the projection of crooked\_path

**Usage**

```
crooked_path_center
```

**Format**

A vector of length 3

A unit vector of length 3 minimizing mean spherical distance to the points of the projection of crooked\_path.

**Source**

Synthetic data.

---

crooked_path_projection	<i>Crooked path projection</i>
-------------------------	--------------------------------

---

**Description**

The projection of the last 8 points on crooked\_path onto the unit sphere as seen from the 6th.  
This is a collection of 8 unit points in dimension 3.

**Usage**

```
crooked_path_projection
```

**Format**

An 8 x 3 matrix

The projection of crooked\_path[7:14,] onto the unit sphere as seen from crooked\_path[6,] .

**Source**

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometry>

---

crooked_path_radius	<i>Crooked path radius</i>
---------------------	----------------------------

---

### Description

The mean spherical distance from the points of the projection of crooked\_path to the point minimizing this mean distance.

### Usage

```
crooked_path_radius
```

### Format

Numeric

The mean spherical distance from the points of the projection of crooked\_path to the point minimizing this mean distance.

### Source

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometryI>

---

distanceBetweenTrajectories	<i>Get distances between trajectories.</i>
-----------------------------	--

---

### Description

This function compares two single cell trajectories (representative of different lineages within the same dataset), and finds the minimum euclidean distance between the first and the second trajectory at each point in pseudotime. Please note, attributes can either be values for single cells, or attributes which have been smoothed over pseudotime. Likewise the pseudotime values should be for single cells, or for smoothed attributes over pseudotime

### Usage

```
distanceBetweenTrajectories(attributes1, pseudotime1, attributes2)
```

### Arguments

attributes1	- An n x d (cell x attribute) matrix of numeric attributes for the first single cell trajectory.
pseudotime1	- A named numeric vector of pseudotime values for the first single cell trajectory, names should match rownames of attributes1.
attributes2	- An n x d (cell x attribute) matrix of numeric attributes for the sencond single cell trajectory.

**Value**

results - a dataframe containing pseudotime values (for the first trajectory), and distances (the minimum euclidian distance between the two trajectories at that point in pseudotime).

**Examples**

```
distances = distanceBetweenTrajectories(chol_attributes,
                                         chol_pseudo_time[!is.na(chol_pseudo_time)],
                                         hep_attributes)
```

---

findSphereClusterCenter

*Find a center for points on the unit sphere*

---

**Description**

This function takes a set of points on the d-1 sphere in d-space and finds a center for these. Depending on choice of statistic, this center is a point on the sphere which minimizes either the median distance, the mean distance or the maximum distance of the center to the given points. "Distance" here is taken to mean angle between the points, i.e., arccos of their dot product.

**Usage**

```
findSphereClusterCenter(points, statistic, normalize = FALSE)
```

**Arguments**

points	- A set of n points on the (d-1) sphere given as an n x d matrix.
statistic	- The statistic to be minimized. Allowable values are 'median', 'mean' or 'max'.
normalize	- If this is set to TRUE, the function will start by normalizing the input points.

**Value**

This returns a point in dimension d given as a vector.

**Examples**

```
projection = projectPathToSphere(straight_path)
center = findSphereClusterCenter(projection, 'mean')
```

---

`findSphericalDistance`    *Find the spherical distance from a given point to a set of points.*

---

### Description

This function takes a point (typically a center) and a set of points and finds the spherical distance between the given point and each of the others. If requested, it will first normalize all of them.

### Usage

```
findSphericalDistance(center, points, normalize = FALSE)
```

### Arguments

<code>center</code>	- The proposed point from which distance to the others should be measured. This is a numerical vector of length d.
<code>points</code>	- The set of target points for which spherical distance to the center should be calculated. This is in the form of a n x d matrix.
<code>normalize</code>	- If this is set to TRUE, the function will start by normalizing the input points.

### Value

This returns a vector of n spherical distances in radians.

### Examples

```
distances = findSphericalDistance(straight_path_center,
                                   straight_path_projection)
```

---

`generateRandomPaths`    *Produce random paths modeled on a given path*

---

### Description

This function takes a path and produces N random paths of the same dimension and length based on it. This can be done either by permuting the entries in path or by taking steps from the initial point of path. Exact behaviour is controlled by `randomizationParams`.

### Usage

```
generateRandomPaths(
  path,
  from = 1,
  to = nrow(path),
  d = ncol(path),
  randomizationParams,
  N
)
```

**Arguments**

- path                   - This is an mxn dimensional matrix. Each row is considered a point.
- from                   - The starting place along the path which will be treated as the center of the sphere. This defaults to 1.
- to                     - The end point of the path. This defaults to nrow(path).
- d                      - The dimension under consideration. This defaults to ncol(path)
- randomizationParams   - A character vector controlling the randomization method used. It's first entry must be either 'byPermutation' or 'bySteps' See the vignette for further details.
- N                      - The number of random paths required.

**Value**

This function returns a list of random paths. Each path is a matrix.

**Examples**

```
randomizationParams = c('byPermutation','permuteWithinColumns')
randomPaths = generateRandomPaths(crooked_path,from=6,to=nrow(crooked_path),
                                   d=ncol(crooked_path),randomizationParams=randomizationParams,
                                   N=10)
```

---

generateRandomUnitVector

*Generate random unit vector.*

---

**Description**

This function generates a random unit vector in in dimension d.

**Usage**

```
generateRandomUnitVector(d)
```

**Arguments**

- d                    - The dimension.

**Value**

A unit vector in dimension d.

**Examples**

```
randomUnitVector = generateRandomUnitVector(5)
```

---

getDistanceDataForPaths

*Produce distance statistics for random paths*


---

### Description

This function takes a list of paths and a choice of statistic (median, mean or max) and returns that statistic for the appropriate center for each path. Each path is an  $n \times d$  matrix. In use, it is assumed that these will be the randomized paths. It is therefore assumed that they are already of the correct dimensions.

### Usage

```
getDistanceDataForPaths(paths, statistic)
```

### Arguments

paths	- A list of paths. Each of these is an $n \times d$ matrix.
statistic	- Allowable values are 'median', 'mean' or 'max'.

### Value

This returns a vector of  $n$  distances.

### Examples

```
paths =
  generateRandomPaths(path=straight_path, randomizationParam='bySteps', N=5)
distance = getDistanceDataForPaths(paths=paths, statistic='max')
```

---

getSphericalData

*This is a simplified wrapper for pathToSphericalData*


---

### Description

It handles the case in which from, to and  $d$  are all given by the dimensions of the path

### Usage

```
getSphericalData(path, statistic)
```

### Arguments

path	- an $m \times n$ matrix. Each row is considered a point
statistic	- one of 'mean', 'median' or 'max'

### Value

This function returns a list whose elements are the projections of the path to the sphere, the center for those projections, the median, mean or max distance from the center to those projections and the name of the statistic used.

**Examples**

```
sphericalData = getSphericalData(straight_path, 'max')
```

---

getStepLengths	<i>Find the step lengths:</i>
----------------	-------------------------------

---

**Description**

This finds the lengths of the steps along a path

**Usage**

```
getStepLengths(path, from = 1, to = nrow(path), d = ncol(path))
```

**Arguments**

- |      |  |
|------|--|
| path | - This is an mxn dimensional matrix. Each row is considered a point.                                       |
| from | - The starting place along the path which will be treated as the center of the sphere. This defaults to 1. |
| to   | - The end point of the path. This defaults to nrow(path).  |
| d    | - The dimension under consideration. This defaults to ncol(path)   |

**Value**

This function returns the length of each step in a path.

**Examples**

```
stepLengths = getStepLengths(path=crooked_path)
stepLengths = getStepLengths(path=crooked_path, from=4)
```

---

hep_answers	<i>hep_answers</i>
-------------	--------------------

---

**Description**

Results of running analyseSingleCellTrajectory() on a trajectory describing the development of hepatocytes from hepatoblasts.

**Usage**

```
hep_answers
```

**Format**

A list

Results of running analyseSingleCellTrajectory() on a trajectory describing the development of hepatocytes from hepatoblasts.

**Source**

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

hep_attributes	<i>hep_attributes</i>
----------------	-----------------------

---

**Description**

PCA projections derived from normalised gene expression values for single cells, and filtered for cells which feature in a trajectory from hepatoblast to hepatocyte. The columns are the PCs and the rows are the cells.

**Usage**

hep\_attributes

**Format**

A matrix

PCA projections derived from normalised gene expression values for single cells, and filtered for cells which feature in a trajectory from hepatoblast to hepatocyte. The columns are the PCs and the rows are the cells.

**Source**

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

hep_pseudo_time	<i>hep_pseudo_time</i>
-----------------	------------------------

---

**Description**

A vector of pseudotime values for a trajectory describing the development of hepatocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.

**Usage**

hep\_pseudo\_time

**Format**

A vector

A vector of pseudotime values for a trajectory describing the development of hepatocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.



## Source

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

```
hep_pseudo_time_normalised
      hep_pseudo_time_normalised
```

---

## Description

A vector of pseudotime values, normalised to range from 0 to 100, for a trajectory describing the development of hepatocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.

## Usage

```
hep_pseudo_time_normalised
```

## Format

A vector

A vector of pseudotime values for a trajectory describing the development of hepatocytes from hepatoblasts. Pseudotime values have been inferred using the SlingShot package. The vector is named according to cell ID.

## Source

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

```
orthonormalBasis      Find an orthonormal basis in dimension 3
```

---

## Description

Given a vector in R3, this normalizes it and then uses it as the first basis vector in an orthonormal basis. We'll use this to find circles around points on the sphere.

## Usage

```
orthonormalBasis(x)
```

## Arguments

x                      - A vector of length 3

## Value

This function returns an orthonormal basis in the the form of a 3 x 3 matrix in which the first vector is parallel to v

**Examples**

```
anOrthonormalBasis = orthonormalBasis(c(1,1,1))
```

---

oscillation

*Oscillation*


---

**Description**

This a path which prepends small oscillations to straight path. Its purpose is to illustrate instability of spherical projection near the beginning of a path.

**Usage**

```
oscillation
```

**Format**

A matrix

This a path which prepends small oscillations to straight path. Its purpose is to illustrate instability of spherical projection near the beginning of a path.

**Source**

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometryL>

---

pathProgression

*Measure a path's progression*


---

**Description**

This function measures the progress of a path in a specified direction. This direction will typically be the center of its projection onto the sphere as revealed using your favorite statistic.

**Usage**

```
pathProgression(path, from = 1, to = nrow(path), d = ncol(path), direction)
```

**Arguments**

path	- An n x d matrix
from	- The point along the path to be taken as the starting point. This defaults to 1.
to	- The point along the path to be used as the end point. This defaults to nrow(path).
d	- The dimension to be used. This defaults to ncol(path).
direction	- A non-zero numeric whose length is the the dimension.

**Value**

This returns a numeric given the signed distance projection of the path along the line through its starting point in the given direction.



---

plotPathProjectionCenterAndCircle

*Plot a path, its projection, its center and its circle*


---

### Description

This function assumes you have a path in dimension 3 and you have found the projection for the portion under consideration, the center for its projection and the circle (i.e., radius) for the appropriate statistic. Scales the path to keep it comparable to the sphere and plots all this in your favorite color. It can be called repeatedly to add additional paths in different colors.

### Usage

```
plotPathProjectionCenterAndCircle(
  path,
  from = 1,
  to = nrow(path),
  projection,
  center,
  radius,
  color,
  circleColor = "white",
  pathPointSize = 8,
  projectionPointSize = 8,
  scale = 1.5,
  newFigure = TRUE
)
```

### Arguments

path	- A path of dimension 3 in the form of an N x 3 matrix.
from	- The starting place of the section under consideration. This is used for marking the relevant portion. It defaults to 1.
to	- Likewise. It defaults to nrow(path).
projection	- The projection of the relevant portion of the path.
center	- The center of the projection points.
radius	- The radius of the circle.
color	- The color to use for this path and its associated data.
circleColor	- Sets the colour of the circle. Defaults to white.
pathPointSize	- Sets the size of points which represent the path. Defaults to 8.
projectionPointSize	- Sets the size of points which represent the projected path. Defaults to 8.
scale	- The path will be start (its actual start) at 0 and will be scaled so that its most distant point will be at this distance from the origin. This is to keep it comparable in size to the sphere. It defaults to 1.5. Caution should be used here when plotting multiple paths.
newFigure	- When plotting a single figure or the first of multiple figures, this should be set to TRUE which is its default. Otherwise, set this to FALSE in order to add additional paths to the same figure.

**Value**

This returns 0.

**Examples**

```
plotPathProjectionCenterAndCircle(path=straight_path,
                                   projection=straight_path_projection,
                                   center=straight_path_center,
                                   radius=straight_path_radius,
                                   color='red',
                                   newFigure=TRUE)
```

---

projectPathToSphere	<i>Project a path onto the unit sphere</i>
---------------------	--

---

**Description**

This function takes a path in d dimensional space and projects it onto the d-1 sphere. It takes as additional arguments the starting and ending points under consideration and the dimension to be considered.

**Usage**

```
projectPathToSphere(path, from = 1, to = nrow(path), d = ncol(path))
```

**Arguments**

path	- This is an mxn dimensional matrix. Each row is considered a point.
from	- The starting place along the path which will be treated as the center of the sphere. This defaults to 1.
to	- The end point of the path. This defaults to nrow(path).
d	- The dimension under consideration. This defaults to ncol(path)

**Value**

This returns a projection of the path onto the d-1 sphere in the form of a (to - from) x d matrix.

**Examples**

```
projection1 = projectPathToSphere(straight_path)
projection2 = projectPathToSphere(crooked_path, from=6)
```

---

samplePath

*Sample a path from single cell data*


---

### Description

This function takes vector of pseudotime values, and a matrix of attribute values (cell x attribute). It also optionally takes the number of pseudotime windows to sample a single cell from. This defaults to 10. The function returns a matrix of sampled attribute values which form the coordinates of the sampled path. The matrix of attribute values should consist of numeric values relevant to a pseudotime trajectory i.e. gene expression values or PCA projections. The vector of pseudotime values should be named according to cell names. Similarly the row names of the matrix of attribute values should be cell names. Row names for the returned matrix of the sampled path give the window number a cell was sampled from.

### Usage

```
samplePath(attributes, pseudotime, nWindows = 10)
```

### Arguments

attributes	- An n x d (cell x attribute) matrix of numeric attributes for single cell data. Rownames should be cell names.
pseudotime	- A named numeric vector of pseudotime values for cells.
nWindows	- The number of windows pseudotime should be split into to sample cells from. Defaults to 10.

### Value

sampledPath - A path consisting of a matrix of attributes of sampled cells. The rownames refer to the pseudotime windows cell was sampled from.

### Examples

```
samplePath(chol_attributes, chol_pseudo_time_normalised)
samplePath(hep_attributes, hep_pseudo_time_normalised)
```

---

single\_cell\_matrix

*single\_cell\_matrix*


---

### Description

PCA projections derived from normalised gene expression values for single cells. The columns are the PCs and the rows are the cells.

### Usage

```
single_cell_matrix
```

**Format**

A matrix

PCA projections derived from normalised gene expression values for single cells. The columns are the PCs and the rows are the cells.

**Source**

Single-cell data has been obtained from GEO (GSE90047) and the script used for upstream processing is available at <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

straight_path	<i>Straight path</i>
---------------	----------------------

---

**Description**

A path of  $n$  points in dimension  $d$  is an  $n \times d$  matrix. This particular path is relatively straight.

**Usage**

```
straight_path
```

**Format**

A  $14 \times 3$  matrix

The path is roughly in the (1,0,0) direction.

**Source**

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

straight_path_center	<i>Straight path center</i>
----------------------	-----------------------------

---

**Description**

The point on the unit sphere minimizing mean spherical distance to the projection so straight\_path

**Usage**

```
straight_path_center
```

**Format**

A vector of length 3

A unit vector of length 3 minimizing distance to the points of the projection of straight\_path.

**Source**

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometryData>

---

straight\_path\_projection

*Straight path projection*


---

### Description

The projection of straight\_path onto the unit sphere. This is a collection of 13 unit points in dimension 3

### Usage

```
straight_path_projection
```

### Format

A 13 x 3 matrix

The projection of straight\_path[2:14,] onto the unit sphere as seen from straight\_path[1,] .

### Source

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometry>

---

straight\_path\_radius

*Straight path radius*


---

### Description

The mean spherical distance from the points of the projection of straight\_path to the point minimizing this mean distance.

### Usage

```
straight_path_radius
```

### Format

Numeric

The mean spherical distance from the points of the projection of straight\_path to the point minimizing this mean distance.

### Source

This was created by code in createSyntheticData.R available from <https://github.com/AnnaLaddach/TrajectoryGeometry>





---

```
visualiseBranchPointStats
```

*Visualise Branch Point Stats*

---

### Description

This function creates plots and extracts statistics for analysing branch points. It returns plots and underlying data for visualising distance metrics and  $-\log_{10}$  transformed pvalues (comparison to random trajectories) for trajectories with different starting points.

### Usage

```
visualiseBranchPointStats(branchPointData, average = "mean")
```

### Arguments

`branchPointData` - the result of `analyseBranchPoint`

`average` - if there are multiple distances available for each sampled trajectory, calculate the average using "mean" or "median" (defaults to "mean").

### Value

a list containing: `branchPointValues` - dataframe containing data underlying distance plot in long format `pValues` - dataframe containing data underlying p-value plot in long format `distancePlot` - ggplot object, violin plots of distance metric for sampled paths for different trajectory different starting points `pValue` - ggplot object, line plot of  $-\log_{10}$  transformed p-values for comparing sampled paths to random paths for different trajectory starting points

### Examples

```
cholBranchPointStats = visualiseBranchPointStats(chol_branch_point_results)
```

---

```
visualiseTrajectoryStats
```

*Visualise Trajectory Stats*

---

### Description

This function creates plots and extracts statistics for comparisons of metrics for sampled paths to random paths. It can also create plots for comparing two sets of sampled paths by providing the `traj2Data` argument.

### Usage

```
visualiseTrajectoryStats(
  traj1Data,
  metric,
  average = "mean",
  traj2Data = list()
)
```

**Arguments**

- traj1Data        - the result of analyseSingleCellTrajectory
- metric           - either "pValue" or "distance"
- average          - if there are multiple distances available for each sampled trajectory, calculate the average using "mean" or "median" (defaults to "mean").
- traj2Data        - traj2Data either an empty list or the result of analyseSingleCellTrajectory

**Value**

a list containing: stats - output of wilcox test (paired if comparing sampled to random paths, unpaired if comparing sampled paths for two different trajectories) values - dataframe containing plotted data in long format plot - ggplot object

**Examples**

```
cholResultDistance = visualiseTrajectoryStats(chol_answers, "distance")
hepResultDistance = visualiseTrajectoryStats(hep_answers, "distance")
distanceComparison = visualiseTrajectoryStats(chol_answers, "distance",
  traj2Data = hep_answers)
```

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