

# Package ‘CARDspa’

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**Title** Spatially Informed Cell Type Deconvolution for Spatial Transcriptomics

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**Description** CARD is a reference-based deconvolution method that estimates cell type composition in spatial transcriptomics based on cell type specific expression information obtained from a reference scRNA-seq data. A key feature of CARD is its ability to accommodate spatial correlation in the cell type composition across tissue locations, enabling accurate and spatially informed cell type deconvolution as well as refined spatial map construction. CARD relies on an efficient optimization algorithm for constrained maximum likelihood estimation and is scalable to spatial transcriptomics with tens of thousands of spatial locations and tens of thousands of genes.

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assign_sc_cords	<i>The function to assign the spatial location information for each single cell</i>
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## Description

The function to assign the spatial location information for each single cell

## Usage

```
assign_sc_cords(mappint_spot_cell_cor, cords_new, numcell, sc_eset, ct_varname)
```

## Arguments

mappint_spot_cell_cor	a mapped correlation matrix indicating the relationship between each measured spatial location and the single cell in the scRNAseq reference
cords_new	output from the function get_high_res_cords
numcell	a numeric value indicating the number of single cells in each measured location, we suggest 20 for ST technology, 7 for 10x Viisum and 2 for Slide-seq
sc_eset	a single cell experiment object stored in CARD object
ct_varname	character, the name of the column in metaData that specifies the cell type annotation information, stroed in CARD object

## Value

Return the assigned spatial location information for the mapped single cell

---

CARD-class	<i>Each CARD object has a number of slots which store information. Key slots to access are listed below.</i>
------------	--

---

## Description

Each CARD object has a number of slots which store information. Key slots to access are listed below.

## Value

Return an object of CARD class

**Slots**

sc_eset	The filtered scRNA-seq data along with meta data stored in the format of SingleCellExperiment.
spatial_countMat	The filtered spatial count data.
spatial_location	The weights for combining p-values from multiple kernels.
Proportion_CARD	The estimated cell type proportion by CARD with each row is a spatial location and each column is a cell type.
project	The name of the project, default is deconvolution.
info_parameters	The parameters that are used in model fitting.
algorithm_matrix	The intermediate matrices that are used in the model fitting step.
refined_prop	The refined cell type proportion matrix estimated by CARD for the newly grided spatial locations. The number of initial grids are defined by the user.
refined_expression	The refined predicted expression matrix (normalized) estimated by CARD for the newly grided spatial locations. The number of initial grids are defined by the user.

CARDfree

*SpatialDeconv function based on Conditional Autoregressive model***Description**

SpatialDeconv function based on Conditional Autoregressive model

**Usage**

```
CARDfree(
  XinputIn,
  UIn,
  WIn,
  phiIn,
  max_iterIn,
  epsilonIn,
  initV,
  initb,
  initSigma_e2,
  initLambda
)
```

**Arguments**

XinputIn	The input of normalized spatial data
UIn	The input of cell type specific basis matrix B
WIn	The constructed W weight matrix from Gaussian kernel
phiIn	The phi value
max_iterIn	Maximum iterations
epsilonIn	epsilon for convergence
initV	Initial matrix of cell type compositions V
initb	Initial vector of cell type specific intercept
initSigma_e2	Initial value of residual variance
initLambda	Initial vector of cell type specific scalar.

**Value**

A list

---

CARDfree-class	<i>Each CARDfree object has a number of slots which store information. Key slots to access are listed below.</i>
----------------	--

---

**Description**

Each CARDfree object has a number of slots which store information. Key slots to access are listed below.

**Value**

Return an object of CARDfree class

**Slots**

spatial\_countMat The filtered spatial count data.

spatial\_location The weights for combining p-values from multiple kernels.

Proportion\_CARD The estimated cell type proportion by CARD with each row is a spatial location and each column is a cell type.

estimated\_refMatrix The estimated reference matrix by CARDfree with each row represents a gene and each column represents a cell type cluster.

project The name of the project, default is deconvolution.

markerList The nlist of cell type specific markers, with each element represents the vector of cell type specific markers

info\_parameters The paramters that are used in model fitting.

algorithm\_matrix The intermediate matrices that are used in the model fitting step.

refined\_prop The refined cell type proportion matrix estimated by CARD for the newly grided spatial locations. The number of initial grids are defined by the user.

refined\_expression The refined predicted expression matrix (normalized) estimated by CARD for the newly grided spatial locations. The number of initial grids are defined by the user.

---

CARDref	<i>SpatialDeconv function based on Conditional Autoregressive model</i>
---------	---

---

**Description**

SpatialDeconv function based on Conditional Autoregressive model

**Usage**

```
CARDref(
  XinputIn,
  UIn,
  WIn,
  phiIn,
  max_iterIn,
  epsilonIn,
  initV,
  initb,
  initSigma_e2,
  initLambda
)
```

**Arguments**

XinputIn	The input of normalized spatial data
UIn	The input of cell type specific basis matrix B
WIn	The constructed W weight matrix from Gaussian kernel
phiIn	The phi value
max_iterIn	Maximum iterations
epsilonIn	epsilon for convergence
initV	Initial matrix of cell type compositions V
initb	Initial vector of cell type specific intercept
initSigma_e2	Initial value of residual variance
initLambda	Initial vector of cell type sepcific scalar.

**Value**

A list

---

CARD_deconvolution	<i>Spatially Informed Cell Type Deconvolution for Spatial Transcriptomics by CARD</i>
--------------------	---

---

**Description**

Spatially Informed Cell Type Deconvolution for Spatial Transcriptomics by CARD

**Usage**

```
CARD_deconvolution(
  sc_count,
  sc_meta,
  spatial_count,
  spatial_location,
  ct_varname,
  ct_select,
```

```

    sample_varname,
    mincountgene = 100,
    mincountspot = 5,
    sce = NULL,
    spe = NULL
  )

```

### Arguments

<code>sc_count</code>	Raw scRNA-seq count data, each column is a cell and each row is a gene.
<code>sc_meta</code>	data frame, with each row representing the cell type and/or sample information of a specific cell. The row names of this data frame should match exactly with the column names of the <code>sc_count</code> data
<code>spatial_count</code>	Raw spatial resolved transcriptomics data, each column is a spatial location, and each row is a gene.
<code>spatial_location</code>	data frame, with two columns representing the x and y coordinates of the spatial location. The rownames of this data frame should match exactly with the columns of the <code>spatial_count</code> .
<code>ct_varname</code>	character, the name of the column in <code>metaData</code> that specifies the cell type annotation information
<code>ct_select</code>	vector of cell type names that you are interested in to deconvolute, default as <code>NULL</code> . If <code>NULL</code> , then use all cell types provided by single cell dataset;
<code>sample_varname</code>	character, the name of the column in <code>metaData</code> that specifies the sample information. If <code>NULL</code> , we just use the whole as one sample.
<code>mincountgene</code>	Minimum counts for each gene
<code>mincountspot</code>	Minimum counts for each spatial location
<code>sce</code>	a <code>SingleCellExperiment</code> object containing scRNA-seq count data in the <code>counts</code> assay, and cell types and sample information in the <code>colData</code> .
<code>spe</code>	a <code>SpatialExperiment</code> object containing spatial data in the <code>counts</code> assay, and spatial coordinates in the <code>spatialCoords</code> .

### Value

Returns a `SpatialExperiment` object with estimated cell type proportion stored in `object$Proportion_CARD`.

### Examples

```

library(RcppML)
library(NMF)
library(RcppArmadillo)
data(spatial_count)
data(spatial_location)
data(sc_count)
data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",

```

```

ct_select = unique(sc_meta$cellType),
sample_varname = "sampleInfo",
mincountgene = 100,
mincountspot = 5
)

```

---

CARD_imputation	<i>Construct an enhanced spatial expression map on the unmeasured tissue locations</i>
-----------------	--

---

## Description

Construct an enhanced spatial expression map on the unmeasured tissue locations

## Usage

```
CARD_imputation(CARD_object, num_grids, ineibor = 10, exclude = NULL)
```

## Arguments

CARD_object	SpatialExperiment Object created by CARD_deconvolution with estimated cell type compositions on the original spatial resolved transcriptomics data.
num_grids	Initial number of newly grided spatial locations. The final number of newly grided spatial locations will be lower than this value since the newly grided locations outside the shape of the tissue will be filtered
ineibor	Numeric, number of neighbors used in the imputation on newly grided spatial locations, default is 10.
exclude	Vector, the rownames of spatial location data on the original resolution that you want to exclude. This is to avoid the weird detection of the shape.

## Value

Return a SpatialExperiment object with the refined cell type compositions estimated for newly grided spots and the refined predicted gene expression (normalized).

## Examples

```

data(spatial_count)
data(spatial_location)
data(sc_count)
data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",
  ct_select = unique(sc_meta$cellType),
  sample_varname = "sampleInfo",
  mincountgene = 100,
  mincountspot = 5
)

```



```
CARD_obj <- CARD_imputation(
  CARD_obj,
  num_grids = 200,
  ineibor = 10,
  exclude = NULL
)
```

---

CARD_refFree	<i>Extension of CARD into a reference-free version of deconvolution: CARDfree.</i>
--------------	--

---

## Description

Extension of CARD into a reference-free version of deconvolution: CARDfree.

## Usage

```
CARD_refFree(
  markerlist,
  spatial_count,
  spatial_location,
  mincountgene = 100,
  mincountspot = 5,
  spe = NULL
)
```

## Arguments

markerlist	a list of marker genes, with each element of the list being the vector of cell type specific marker genes
spatial_count	Raw spatial resolved transcriptomics data, each column is a spatial location, and each row is a gene.
spatial_location	data frame, with two columns representing the x and y coordinates of the spatial location. The rownames of this data frame should match exactly with the columns of the spatial_count.
mincountgene	Minimum counts for each gene
mincountspot	Minimum counts for each spatial location
spe	a SpatialExperiment object containing spatial data in the counts assay, and spatial coordinates in the spatialCoords.

## Value

Returns a SpatialExperiment object with estimated cell type proportion stored in object\$Proportion\_CARD. Because this is a reference-free version, the columns of estimated proportion is not cell type but cell type cluster

**Examples**

```

library(RcppML)
library(NMF)
library(RcppArmadillo)
data(markerList)
data(spatial_count)
data(spatial_location)
CARDfree_obj <- CARD_refFree(
  markerlist = markerList[8:16],
  spatial_count = spatial_count[1:2500, ],
  spatial_location = spatial_location,
  mincountgene = 100,
  mincountspot = 5
)

```

---

CARD_scmapping	<i>Extension of CARD into performing single cell Mapping from non-single cell spatial transcriptomics dataset.</i>
----------------	--

---

**Description**

Extension of CARD into performing single cell Mapping from non-single cell spatial transcriptomics dataset.

**Usage**

```
CARD_scmapping(CARD_object, shapeSpot = "Square", numcell, ncore = 10)
```

**Arguments**

CARD_object	CARD object create by the CARD_deconvolution function.
shapeSpot	a character indicating whether the sampled spatial coordinates for single cells locating in a Square-like region or a Circle-like region. The center of this region is the measured spatial location in the non-single cell resolution spatial transcriptomics data. The default is 'Square', the other shape is 'Circle'
numcell	a numeric value indicating the number of single cells in each measured location, we suggest 20 for ST technology, 7 for 10x Viisum and 2 for Slide-seq
ncore	a numeric value indicating the number of cores used to accelerating the procedure

**Value**

Returns a SingleCellExperiment SCE object with the mapped expression at single cell resolution and the spatial location information of each single cell

**Examples**

```
library(SingleCellExperiment)
data(spatial_count)
data(spatial_location)
data(sc_count)
data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",
  ct_select = unique(sc_meta$cellType),
  sample_varname = "sampleInfo",
  mincountgene = 100,
  mincountspot = 5
)
scMapping <- CARD_scmapping(
  CARD_obj,
  shapeSpot = "Square",
  numcell = 20,
  ncore = 2)
print(scMapping)
```

---

CARD_visualize_Cor	<i>Visualize the cell type proportion correlation</i>
--------------------	---

---

**Description**

Visualize the cell type proportion correlation

**Usage**

```
CARD_visualize_Cor(proportion, colors = colors)
```

**Arguments**

proportion	Data frame, cell type proportion estimated by CARD in either original resolution or enhanced resolution.
colors	Vector of color names that you want to use, if NULL, we will use the default color scale c("#91a28c","white","#8f2c37")

**Value**

Returns a ggcorrplot figure.

**Examples**

```
library(ggplot2)
data(spatial_count)
data(spatial_location)
data(sc_count)
```

```

data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",
  ct_select = unique(sc_meta$cellType),
  sample_varname = "sampleInfo",
  mincountgene = 100,
  mincountspot = 5
)
CARD_visualize_Cor(CARD_obj$Proportion_CARD, colors = NULL)

```

---

CARD_visualize_gene	<i>Visualize the spatial distribution of cell type proportion</i>
---------------------	---

---

## Description

Visualize the spatial distribution of cell type proportion

## Usage

```

CARD_visualize_gene(
  spatial_expression,
  spatial_location,
  gene_visualize,
  colors = colors,
  NumCols
)

```

## Arguments

<code>spatial_expression</code>	Data frame, spatial gene expression in either original resolution or enhanced resolution.
<code>spatial_location</code>	Data frame, spatial location information.
<code>gene_visualize</code>	Vector of selected gene names that are interested to visualize
<code>colors</code>	Vector of color names that you want to use, if NULL, we will use the default color scale in viridis palette
<code>NumCols</code>	Numeric, number of columns in the figure panel, it depends on the number of cell types you want to visualize.

## Value

Returns a ggplot2 figure.

**Examples**

```

library(ggplot2)
library(SummarizedExperiment)
library(SpatialExperiment)
data(spatial_count)
data(spatial_location)
data(sc_count)
data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",
  ct_select = unique(sc_meta$cellType),
  sample_varname = "sampleInfo",
  mincountgene = 100,
  mincountspot = 5
)
CARD_visualize_gene(
  spatial_expression = assays(CARD_obj)$spatial_countMat,
  spatial_location = spatialCoords(CARD_obj),
  gene_visualize = c("A4GNT", "AAMDC", "CD248"),
  colors = NULL,
  NumCols = 3
)

```

---

CARD_visualize_pie	<i>Visualize the spatial distribution of cell type proportion in a geom scatterpie plot</i>
--------------------	---

---

**Description**

Visualize the spatial distribution of cell type proportion in a geom scatterpie plot

**Usage**

```
CARD_visualize_pie(proportion, spatial_location, colors = NULL, radius = NULL)
```

**Arguments**

proportion	Data frame, cell type proportion estimated by CARD in either original resolution or enhanced resolution.
spatial_location	Data frame, spatial location information.
colors	Vector of color names that you want to use, if NULL, we will use the color palette "Spectral" from RColorBrewer package.
radius	Numeric value about the radius of each pie chart, if NULL, we will calculate it inside the function.

**Value**

Returns a ggplot2 figure.

**Examples**

```
library(ggplot2)
library(SpatialExperiment)
data(spatial_count)
data(spatial_location)
data(sc_count)
data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",
  ct_select = unique(sc_meta$cellType),
  sample_varname = "sampleInfo",
  mincountgene = 100,
  mincountspot = 5
)
colors <- c(
  "#FFD92F", "#4DAF4A", "#FCCDE5", "#D9D9D9", "#377EB8", "#7FC97F",
  "#BEAED4", "#FDC086", "#FFFF99", "#386CB0", "#F0027F", "#BF5B17",
  "#666666", "#1B9E77", "#D95F02", "#7570B3", "#E7298A", "#66A61E",
  "#E6AB02", "#A6761D"
)
CARD_visualize_pie(
  proportion = CARD_obj$Proportion_CARD,
  spatial_location = spatialCoords(CARD_obj),
  colors = colors,
  radius = 0.52
)
```

---

CARD_visualize_prop	Visualize the spatial distribution of cell type proportion
---------------------	--

---

**Description**

Visualize the spatial distribution of cell type proportion

**Usage**

```
CARD_visualize_prop(
  proportion,
  spatial_location,
  ct_visualize = ct_visualize,
  colors = c("lightblue", "lightyellow", "red"),
  NumCols,
  pointSize = 3
)
```

**Arguments**

proportion	Data frame, cell type proportion estimated by CARD in either original resolution or enhanced resolution.
spatial_location	Data frame, spatial location information.
ct_visualize	Vector of selected cell type names that are interested to visualize
colors	Vector of color names that you want to use, if NULL, we will use the default color scale c("lightblue","lightyellow","red")
NumCols	Numeric, number of columns in the figure panel, it depends on the number of cell types you want to visualize.
pointSize	Size of each point used for plotting

**Value**

Returns a ggplot2 figure.

**Examples**

```
library(ggplot2)
library(SpatialExperiment)
data(spatial_count)
data(spatial_location)
data(sc_count)
data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",
  ct_select = unique(sc_meta$cellType),
  sample_varname = "sampleInfo",
  mincountgene = 100,
  mincountspot = 5
)
ct_visualize <- c(
  "Acinar_cells", "Cancer_clone_A", "Cancer_clone_B",
  "Ductal_terminal_ductal_like", "Ductal_CRISP3_high-centroacinar_like",
  "Ductal_MHC_Class_II", "Ductal_APOL1_high-hypoxic", "Fibroblasts"
)
CARD_visualize_prop(
  proportion = CARD_obj$Proportion_CARD,
  spatial_location = spatialCoords(CARD_obj),
  ct_visualize = ct_visualize,
  colors = c("lightblue", "lightyellow", "red"),
  NumCols = 4,
  pointSize = 3.0
)
```

---

CARD\_visualize\_prop\_2CT

*Visualize the spatial distribution of two cell type proportions on the same plot*

---

## Description

Visualize the spatial distribution of two cell type proportions on the same plot

## Usage

```
CARD_visualize_prop_2CT(
  proportion,
  spatial_location,
  ct2_visualize = ct2_visualize,
  colors = NULL
)
```

## Arguments

proportion	Data frame, cell type proportion estimated by CARD in either original resolution or enhanced resolution.
spatial_location	Data frame, spatial location information.
ct2_visualize	Vector of selected two cell type names that are interested to visualize, here we only focus on two cell types
colors	list of color names that you want to use for each cell type, if NULL, we will use the default color scale list <code>list(c("lightblue", "lightyellow", "red"), c("lightblue", "lightyellow", "black"))</code>

## Value

Returns a ggplot2 figure.

## Examples

```
library(ggplot2)
library(SpatialExperiment)
data(spatial_count)
data(spatial_location)
data(sc_count)
data(sc_meta)
CARD_obj <- CARD_deconvolution(
  sc_count = sc_count,
  sc_meta = sc_meta,
  spatial_count = spatial_count,
  spatial_location = spatial_location,
  ct_varname = "cellType",
  ct_select = unique(sc_meta$cellType),
  sample_varname = "sampleInfo",
  mincountgene = 100,
  mincountspot = 5
)
```



```

CARD_visualize_prop_2CT(
  proportion = CARD_obj$Proportion_CARD,
  spatial_location = spatialCoords(CARD_obj),
  ct2_visualize = c("Cancer_clone_A", "Cancer_clone_B"),
  colors = list(c("lightblue", "lightyellow", "red"), c(
    "lightblue", "lightyellow",
    "black"
  ))
)

```

---

createCARDfreeObject    *Create the CARD object*

---

## Description

Create the CARD object

## Usage

```

createCARDfreeObject(
  markerlist,
  spatial_count,
  spatial_location,
  mincountgene = 100,
  mincountspot = 5,
  spe = NULL
)

```

## Arguments

markerlist	a list of marker genes, with each element of the list being the vector of cell type specific marker genes
spatial_count	Raw spatial resolved transcriptomics data, each column is a spatial location, and each row is a gene.
spatial_location	data frame, with two columns representing the x and y coordinates of the spatial location. The rownames of this data frame should match exactly with the columns of the spatial_count.
mincountgene	Minimum counts for each gene
mincountspot	Minimum counts for each spatial location
spe	a SpatialExperiment object containing spatial data in the counts assay, and spatial coordinates in the spatialCoords.

## Value

Returns CARDfree object with filtered spatial count and marker gene list.

---

createCARDObject	Create the CARD object
------------------	------------------------

---

## Description

Create the CARD object

## Usage

```
createCARDObject(
  sc_count,
  sc_meta,
  spatial_count,
  spatial_location,
  ct_varname,
  ct_select,
  sample_varname,
  mincountgene = 100,
  mincountspot = 5,
  sce = NULL,
  spe = NULL
)
```

## Arguments

sc_count	Raw scRNA-seq count data, each column is a cell and each row is a gene.
sc_meta	data frame, with each row representing the cell type and/or sample information of a specific cell. The row names of this data frame should match exactly with the column names of the sc_count data
spatial_count	Raw spatial resolved transcriptomics data, each column is a spatial location, and each row is a gene.
spatial_location	data frame, with two columns representing the x and y coordinates of the spatial location. The rownames of this data frame should match exactly with the columns of the spatial_count.
ct_varname	character, the name of the column in metadata that specifies the cell type annotation information
ct_select	vector of cell type names that you are interested in to deconvolute, default as NULL. If NULL, then use all cell types provided by single cell dataset;
sample_varname	character, the name of the column in metadata that specifies the sample information. If NULL, we just use the whole as one sample.
mincountgene	Minimum counts for each gene
mincountspot	Minimum counts for each spatial location
sce	a SingleCellExperiment object containing scRNA-seq count data in the counts assay, and cell types and sample information in the colData.
spe	a SpatialExperiment object containing spatial data in the counts assay, and spatial coordinates in the spatialCoords.

**Value**

Returns CARD object with filtered spatial count and single cell RNA-seq dataset.

---

create_ref	<i>Construct the mean gene expression basis matrix (B), this is the faster version</i>
------------	--

---

**Description**

Construct the mean gene expression basis matrix (B), this is the faster version

**Usage**

```
create_ref(sc_eset, ct_select = NULL, ct_varname, sample_varname = NULL)
```

**Arguments**

sc_eset	S4 class for storing data from single-cell experiments. This format is usually created by the package SingleCellExperiment with stored counts, along with the usual metadata for genes and cells.
ct_select	vector of cell type names that you are interested in to deconvolute, default as NULL. If NULL, then use all cell types provided by single cell dataset;
ct_varname	character, the name of the column in metaData that specifies the cell type annotation information
sample_varname	character, the name of the column in metaData that specifies the sample information. If NULL, we just use the whole as one sample.

**Value**

Return a list of basis (B) matrix

---

get_high_res_cords	<i>The function to sample the spatial location information for each single cell</i>
--------------------	---

---

**Description**

The function to sample the spatial location information for each single cell

**Usage**

```
get_high_res_cords(cords, numcell, shape = "Square")
```

**Arguments**

coords	The spatial location information in the measure spatial locations, with the first and second columns represent the 2-D x-y coordinate system
numcell	a numeric value indicating the number of single cells in each measured location, we suggest 20 for ST technology, 7 for 10x Viisum and 2 for Slide-seq
shape	a character indicating whether the sampled spatial coordinates for single cells locating in a Square-like region or a Circle-like region. The center of this region is the measured spatial location in the non-single cell resolution spatial transcriptomics data. The default is 'Square', the other shape is 'Circle'

**Value**

Returns a dataframe with the sampled spatial location information for each single cell

---

get_weight_for_cell	<i>The function to estimate the cell type composition signature for each single cell in the scRNaseq reference data</i>
---------------------	---

---

**Description**

The function to estimate the cell type composition signature for each single cell in the scRNaseq reference data

**Usage**

```
get_weight_for_cell(sc_eset, ct_varname, ct_select, sample_varname, B)
```

**Arguments**

sc_eset	the sc_eset stored in the CARD object
ct_varname	character, the name of the column in metaData that specifies the cell type annotation information, stored in the CARD object
ct_select	vector of cell type names that you are interested in to deconvolute, default as NULL. stored in the CARD object
sample_varname	character, the name of the column in metaData that specifies the sample information. stored in the CARD object
B	reference basis matrix stored in the CARD object.

**Value**

Returns a matrix of the cell type composition signature for each single cell in the scRNaseq reference

---

markerList	<i>marker gene list</i>
------------	-------------------------

---

### Description

The marker gene list is a list format with each element of the list being the cell type specific gene markers.

### Usage

```
data(markerList)
```

### Format

An object of class list of length 20.

---

mvn_cv	<i>Imputation and Construction of High-Resolution Spatial Maps for Cell Type Composition and Gene Expression by the spatial correlation structure between original spatial locations and new grided spatial locations</i>
--------	---

---

### Description

Imputation and Construction of High-Resolution Spatial Maps for Cell Type Composition and Gene Expression by the spatial correlation structure between original spatial locations and new grided spatial locations

### Usage

```
mvn_cv(
  vtrain,
  location_orig,
  train_ind,
  test_ind,
  B,
  xinput_norm,
  optimal_b,
  optimal_phi,
  lambda,
  ineibor
)
```

### Arguments

vtrain	Matrix, estimated V matrix from CARD
location_orig	Data frame, spatial location data frame of the original spatial resolved transcriptomics dataset, stored in the spatialCoords(CARD_object)
train_ind	Vector, index of the original spatial locations

test_ind	Vector, index of the newly grided spatial locations
B	Matrix, used in the deconvolution as the reference basis matrix
xinput_norm	Matrix, used in the deconvolution as the normalized spatial count data
optimal_b	Vector, vector of the intercept for each cell type estimated based on the original spatial resolution
optimal_phi	Numeric, the optimal phi value stored in CARD_object
lambda	Vector, vector of cell type specific scalar in the CAR model
ineibor	Numeric, number of neighbors used in the imputation on newly grided spatial locations, default is 10.

**Value**

Return a list with the imputed Cell type composition Vtest matrix on the newly grided spatial locations and predicted normalized gene expression

---

norm\_coords\_train\_test

*Normalize the new spatial locations without changing the shape and relative positions*

---

**Description**

Normalize the new spatial locations without changing the shape and relative positions

**Usage**

```
norm_coords_train_test(location_orig, train_ind, test_ind)
```

**Arguments**

location_orig	Data frame, spatial location data frame of the original spatial resolved transcriptomics dataset, stored in the spatialCoords(CARD_object)
train_ind	Vector, Index of the original spatial locations
test_ind	Vector, Index of the newly grided spatial locations

**Value**

Return the normalized spatial location data frame

---

sample_grid_within	<i>Make new spatial locations on unmeasured tissue through grids.</i>
--------------------	---

---

### Description

Make new spatial locations on unmeasured tissue through grids.

### Usage

```
sample_grid_within(location, num_sample, concavity = 2)
```

### Arguments

location	Data frame, spatial location data frame of the original spatial resolved transcriptomics dataset, stored in the spatialCoords(CARD_object)
num_sample	Numeric, approximate number of cells in grid within the shape of the spatial location data frame
concavity	Numeric, a relative measure of concavity. The default is 2.0, which can procure detailed enough shapes. Infinity results in a convex hull while 1 results in a more detailed shape.

### Value

Return a list of data frame with newly grided points

---

sc_count	<i>scRNA-seq count data</i>
----------	-----------------------------

---

### Description

The scRNA-seq count data must be in the format of matrix or sparseMatrix, while each row represents a gene and each column represents a cell.

### Usage

```
data(sc_count)
```

### Format

An object of class dgCMatrx with 7000 rows and 1926 columns.

---

sc_meta	<i>scRNAseq meta data</i>
---------	---------------------------

---

### Description

The scRNAseq meta data must be in the format of data frame while each row represents a cell. The rownames of the scRNAseq meta data should match exactly with the column names of the scRNAseq count data. The sc\_meta data must contain the column indicating the cell type assignment for each cell (e.g., "cellType" column in the example sc\_meta data). Sample/subject information should be provided, if there is only one sample, we can add a column by sc\_meta\$sampleInfo = "sample1".

### Usage

```
data(sc_meta)
```

### Format

An object of class `data.frame` with 1926 rows and 3 columns.

---

sc_QC	<i>Quality control of scRNA-seq count data</i>
-------	--

---

### Description

Quality control of scRNA-seq count data

### Usage

```
sc_QC(
  counts_in,
  metadata,
  ct_varname,
  ct_select,
  sample_varname = NULL,
  min_cells = 0,
  min_genes = 0
)
```

### Arguments

counts_in	Raw scRNAseq count data, each column is a cell and each row is a gene.
metadata	data frame, metadata with "ct_varname" specify the cell type annotation information and "sample_varname" specify the sample information
ct_varname	character, the name of the column in metadata that specifies the cell type annotation information
ct_select	vector of cell type names that you are interested in to deconvolute, default as NULL. If NULL, then use all cell types provided by single cell dataset;



sample_varname	character, the name of the column in metadata that specifies the sample information. If NULL, we just use the whole as one sample.
min_cells	numeric, we filtered out the non-expressed cells.
min_genes	numeric we filtered out the non-expressed genes

**Value**

Return the filtered scRNA-seq data and meta data stored in a S4 class (SingleCellExperiment)

---

select_info	<i>Select Informative Genes used in the deconvolution</i>
-------------	---

---

**Description**

Select Informative Genes used in the deconvolution

**Usage**

```
select_info(basis, sc_eset, commongene, ct_select, ct_varname)
```

**Arguments**

basis	Reference basis matrix.
sc_eset	scRNAseq data along with meta data stored in the S4 class format (SingleCellExperiment).
commongene	common genes between scRNAseq count data and spatial resolved transcriptomics data.
ct_select	vector of cell type names that you are interested in to deconvolute, default as NULL. If NULL, then use all cell types provided by single cell dataset;
ct_varname	character, the name of the column in metaData that specifies the cell type annotation information

**Value**

a vector of informative genes selected

---

show, CARD-method	<i>Show method for the CARD class</i>
-------------------	---------------------------------------

---

**Description**

This method provides a concise summary of an object of class CARD, displaying key information including the project name, the number of spots, the number of cell types, and a sample of the Proportion\_CARD matrix.

**Usage**

```
## S4 method for signature 'CARD'
show(object)
```

**Arguments**

object                      An object of class CARD.

**Value**

A concise summary of the CARD object is printed to the console.

---

show, CARDfree-method    *Show method for the CARDfree class*

---

**Description**

This method provides a concise summary of an object of class CARDfree, displaying key information including the project name, the number of spots, the number of cell types, and a sample of the Proportion\_CARD matrix.

**Usage**

```
## S4 method for signature 'CARDfree'
show(object)
```

**Arguments**

object                      An object of class CARDfree.

**Value**

A concise summary of the CARDfree object is printed to the console.

---

Sigma                              *Calculate the variance covariance matrix used in the imputation of the new grided locations*

---

**Description**

Calculate the variance covariance matrix used in the imputation of the new grided locations

**Usage**

```
Sigma(location_orig, train_ind, test_ind, optimal_phi, ineibor)
```

**Arguments**

location_orig	Data frame, spatial location data frame of the original spatial resolved transcriptomics dataset, stored in the spatialCoords(CARD_object)
train_ind	Vector, index of the original spatial locations
test_ind	Vector, index of the newly grided spatial locations
optimal_phi	Numeric, the optimal phi value stored in CARD_object
ineibor	Numeric, number of neighbors used in the imputation on newly grided spatial locations, default is 10.

**Value**

Return a list with the imputed Cell type composition Vtest matrix on the newly grided spatial locations and predicted normalized gene expression

---

spatial_count	<i>Spatial transcriptomics count data</i>
---------------	---

---

**Description**

The spatial transcriptomics count data must be in the format of matrix or sparseMatrix, while each row represents a gene and each column represents a spatial location. The column names of the spatial data can be in the “XcoordYcoord” (i.e., 10x10) format, but you can also maintain your original spot names, for example, barcode names.

**Usage**

```
data(spatial_count)
```

**Format**

An object of class dgMatrix with 11000 rows and 428 columns.

---

spatial_location	<i>Spatial location data</i>
------------------	------------------------------

---

**Description**

The spatial location data must be in the format of data frame while each row represents a spatial location, the first column represents the x coordinate and the second column represents the y coordinate. The rownames of the spatial location data frame should match exactly with the column names of the spatial\_count.

**Usage**

```
data(spatial_location)
```

**Format**

An object of class data.frame with 428 rows and 2 columns.

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