

# Package ‘TSRchitect’

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**Title** Promoter identification from large-scale TSS profiling data

**Description** In recent years, large-scale transcriptional sequence data has yielded considerable insights into the nature of gene expression and regulation in eukaryotes. Techniques that identify the 5' end of mRNAs, most notably CAGE, have mapped the promoter landscape across a number of model organisms. Due to the variability of TSS distributions and the transcriptional noise present in datasets, precisely identifying the active promoter(s) for genes from these datasets is not straightforward. TSRchitect allows the user to efficiently identify the putative promoter (the transcription start region, or TSR) from a variety of TSS profiling data types, including both single-end (e.g. CAGE) as well as paired-end (RAMPAGE, PEAT, STRIPE-seq). In addition, (new with version 1.3.0) TSRchitect provides the ability to import aligned EST and cDNA data. Along with the coordinates of identified TSRs, TSRchitect also calculates the width, abundance and two forms of the Shape Index, and handles biological replicates for expression profiling. Finally, TSRchitect imports annotation files, allowing the user to associate identified promoters with genes and other genomic features. Three detailed examples of TSRchitect's utility are provided in the User's Guide, included with this package.

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**License** GPL-3

**URL** <https://github.com/brendelgroup/tsrchitect>

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`addAnnotationToTSR`      **addAnnotationToTSR**

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**Description**

`addAnnotationToTSR` associates an identified promoter with a given gene, if found upstream and on the same strand within a specified range.

**Usage**

```
addAnnotationToTSR(experimentName, tsrSetType, tsrSet = 1, upstreamDist,
downstreamDist, feature, featureColumnID, writeTable = TRUE)

## S4 method for signature
## 'tssObject,
##   character,
##   numeric,
##   numeric,
##   numeric,
##   character,
##   character,
##   logical'
addAnnotationToTSR(experimentName,
tsrSetType, tsrSet = 1, upstreamDist = 1000, downstreamDist = 200,
feature = "gene", featureColumnID = "ID", writeTable = TRUE)
```

**Arguments**

<code>experimentName</code>	an object of class <i>tssObject</i> with occupied data slots <code>@tsrData</code> (and/or <code>@tsrDataMerged</code> ). The <i>tssObject</i> must already have an annotation attached to the slot <code>@annotation</code> , which is provided by either <code>importAnnotationExternal</code> or <code>importAnnotationHub</code> .
<code>tsrSetType</code>	Specifies the type of TSR set to be processed. Options are "replicates" or "merged".
<code>tsrSet</code>	Number of the data set (of type <i>tsrSetType</i> ) to be processed. (numeric)
<code>upstreamDist</code>	the maximum distance (in bp) upstream of the selected interval necessary to associate a TSR with a given annotation. (numeric)
<code>downstreamDist</code>	the maximum distance (in bp) downstream of the start of the selected interval to associate a TSR with a given annotation. (numeric)
<code>feature</code>	Specifies the feature to be used for annotation (typically "gene" [default] or "mRNA" for GFF3 input); set to "all" if all annotations from the input are to be used. (character)
<code>featureColumnID</code>	Name of the column identifier in the <code>GRanges</code> annotation object. This should be "ID" (default) for GFF3 input or "name" for bed input. (character)
<code>writeTable</code>	logical, specifying whether the output should be written to a tab-delimited file. Defaults to TRUE.

**Value**

`addAnnotationToTSR` adds feature annotation to the (merged) `@tsrData` data frame and returns the updated *tssObject*.

**Note**

An example similar to the this one can be found in the vignette (`/inst/doc/TSRchitect.Rmd`)

## Examples

```
load(system.file("extdata", "tssObjectExample.RData",
package="TSRchitect"))
tssObjectExample <- addAnnotationToTSR(experimentName=tssObjectExample,
tsrSetType="merged", tsrSet=1, upstreamDist=1000, downstreamDist=200,
feature="transcript", featureColumnID="ID", writeTable=FALSE)
#if the object attached to @annotation is a gff/gff3 file
```

addTagCountsToTSR

**addTagCountsToTSR**

## Description

`addTagCountsToTSR` adds a matrix of tag counts to a set of identified TSRs

## Usage

```
addTagCountsToTSR(experimentName, tsrSetType, tsrSet = 1,
tagCountThreshold = 1, writeTable = TRUE)

## S4 method for signature 'tssObject,character,numeric,numeric,logical'
addTagCountsToTSR(experimentName,
tsrSetType, tsrSet = 1, tagCountThreshold = 1, writeTable = TRUE)
```

## Arguments

<code>experimentName</code>	a S4 object of class <i>tssObject</i> containing information in slot <code>@tssTagData</code>
<code>tsrSetType</code>	specifies the set to be written to file. Options are "replicates" or "merged". (character)
<code>tsrSet</code>	number of the dataset to be processed, where 1 corresponds to the first slot, and so on. (numeric)
<code>tagCountThreshold</code>	number of tags required at a given TSS position in order to be included in the overall count for a TSR (numeric)
<code>writeTable</code>	specifies whether the output should be written to a table. (logical)

## Value

a matrix of tag counts and median-normalized tag counts is appended to the data frame of the selected set of identified TSRs in the returned *tssObject*; note that the number of new columns is twice the number of replicates in the sample

## Note

An example similar to the one provided can be found in the vignette (`/inst/doc/TSRchitect.Rmd`)

## Examples

```
load(system.file("extdata", "tssObjectExample.RData", package="TSRchitect"))
tssObjectExample <- addTagCountsToTSR(experimentName=tssObjectExample,
tsrSetType="merged", tsrSet=1, tagCountThreshold=25, writeTable=FALSE)
```

bedToTSS

**bedToTSS**

## Description

bedToTSS extracts TSS information from each attached .bed file in a tssObject object

## Usage

```
bedToTSS(experimentName)

## S4 method for signature 'tssObject'
bedToTSS(experimentName)
```

## Arguments

experimentName an S4 object of class tssObject with bed files loaded

## Value

produces a **GRangesList** containing separate **GRanges** objects for each .bed file contained within *experimentName*, placing them in the returned *tssObject*.

## Note

An example similar to the one provided can be found in the vignette (/inst/doc/TSRchitect.Rmd).

createSummarizedExperiment

**createSummarizedExperiment**

## Description

createSummarizedExperiment creates and returns a *SummarizedExperiment* object from the tag counts on a selected TSR dataset.

## Usage

```
createSummarizedExperiment(experimentName, tsrSetType = "merged",
tsrSet = 1, samplePrefix)

## S4 method for signature 'tssObject,character,numeric,character'
createSummarizedExperiment(experimentName,
tsrSetType = "merged", tsrSet = 1, samplePrefix)
```

### Arguments

experimentName an S4 object of class *tssObject* containing information in slot @*tssTagData*  
 tsrSetType specifies the TSR set to be converted into a *SummarizedExperiment* object. Options are "replicates" or "merged". (character)  
 tsrSet number of the dataset to be processed (numeric).  
 samplePrefix the prefix (or prefixes) that match the sample identifiers in the tsrData column. (character)

### Value

a summarizedExperiment object from the specified TSR data set that is to be written to your working directory.

### Note

For more information on the SummarizedExperiment class, please visit <https://bioconductor.org/packages/release/bioc/vignettes/SummarizedExperiment/inst/doc/SummarizedExperiment.pdf>

### Examples

```
load(system.file("extdata", "tssObjectExample.RData", package="TSRchitect"))
createSummarizedExperiment(tssObjectExample, tsrSetType="merged", tsrSet=1,
samplePrefix=c("sample1", "sample2"))
```

determineTSR

**determineTSR**

### Description

determineTSR Identifies TSRs from entire TSS datasets as specified.

### Usage

```
determineTSR(experimentName, n.cores, tssSetType, tssSet,
tagCountThreshold, clustDist, ...)
```

```
## S4 method for signature
## 'tssObject, numeric, character, character, numeric, numeric'
determineTSR(experimentName,
n.cores = 1, tssSetType = c("replicates", "merged"),
tssSet = "all", tagCountThreshold = 1, clustDist = 20,
writeTable = FALSE, mixedorder = FALSE)
```

### Arguments

experimentName an object of class *tssObject* containing information in slot @*tssTagData*  
 n.cores the number of cores to be used for this job. ncores=1 means serial execution of function calls (numeric)

tssSetType	specifies the set to be clustered. Options are "replicates" or "merged". (character)
tssSet	default is "all"; if a single TSS dataset is desired, specify tssSet number (character)
tagCountThreshold	the number of TSSs required at a given position for it to be considered in TSR identification. (numeric)
clustDist	the maximum distance of TSSs between two TSRs in base pairs. (numeric)
writeTable	specifies whether the output should be written to a table. (logical)
mixedorder	a logical specifying whether the sequence names should be ordered alphanumerically in the output table ("10" following "9" rather than "1"). (logical)

**Value**

creates a list of [GenomicRanges](#)-containing TSR positions in slot `@tsrData` of the returned `tssObject` object

**Note**

An example similar to this one can be found in the vignette (/inst/doc/TSRchitect.Rmd)

**Examples**

```
load(system.file("extdata", "tssObjectExample.RData", package="TSRchitect"))
tssObjectExample <- determineTSR(experimentName=tssObjectExample, n.cores=1,
tssSetType="replicates", tssSet="1", tagCountThreshold=25, clustDist=20,
writeTable=FALSE)
```

**Description**

An internal function, which is invoked using the user-level function `determineTSR` that identifies TSRs from the selected tssSet (Internal function)

**Usage**

```
detTSR(experimentName, tssSetType, tssSet = 1, tagCountThreshold,
       clustDist)

## S4 method for signature 'tssObject,character,numeric,numeric,numeric'
detTSR(experimentName,
       tssSetType, tssSet = 1, tagCountThreshold = 1, clustDist)
```

**Arguments**

`experimentName` - a S4 object of class `tssObject` containing information in slot `tssTagData`  
`tssSetType` - specifies the set to be clustered. Options are "replicates" or "merged"  
`tssSet` - number of the dataset to be analyzed  
`tagCountThreshold`  
                   - number of TSSs required at a given position  
`clustDist` - maximum distance of TSSs between two TSRs (in base pairs)

**Value**

via the user-level function `determineTSR`, creates a list of `GenomicRanges` objects containing TSR positions in slot 'tsrData' on the `tssObject` object

`getBamDataFirstRead`    **getBamDataFirstRead**

**Description**

an accessor function that retrieves the contents of a specified slot "bamDataFirstRead" from a given `tssObject`

**Usage**

```
getBamDataFirstRead(experimentName, slot)

## S4 method for signature 'tssObject,numeric'
getBamDataFirstRead(experimentName, slot)
```

**Arguments**

`experimentName` an S4 object of class `tssObject`  
`slot` 'numeric' a number corresponding to the slot in "bamDataFirstRead" to be retrieved.

**Value**

the contents of the specified slot "bamDataFirstRead" are returned

**Examples**

```
load(system.file("extdata", "tssObjectExample.RData",
package="TSRchitect"))
example.bamDataFirstRead <-
getBamDataFirstRead(experimentName=tssObjectExample, slot = 1)
example.bamDataFirstRead
```

---

getBamDataLastRead **getBamDataLastRead**

---

### Description

an accessor function that retrieves the contents of a specified slot "bamDataLastRead" from a given *tssObject*

### Usage

```
getBamDataLastRead(experimentName, slot)

## S4 method for signature 'tssObject,numeric'
getBamDataLastRead(experimentName, slot)
```

### Arguments

experimentName an S4 object of class *tssObject*  
slot 'numeric' a number corresponding to the slot in "bamDataLastRead" to be retrieved.

### Value

the contents of the specified slot "bamDataLastRead" are returned

---

getFileNames **getFileNames**

---

### Description

an accessor function that retrieves the contents of slot "fileNames" from a given *tssObject*

### Usage

```
getFileNames(experimentName)

## S4 method for signature 'tssObject'
getFileNames(experimentName)
```

### Arguments

experimentName an S4 object of class *tssObject*

### Value

the contents of slot "fileNames" are returned, which are a vector of class "character"

## Examples

```
load(system.file("extdata", "tssObjectExample.RData",
  package="TSRchitect"))
example.file.names <- getFileName(experimentName=tssObjectExample)
example.file.names
```

**getTitle**

**getTitle**

## Description

an accessor function that retrieves the contents of slot "title" from a given *tssObject*

## Usage

```
getTitle(experimentName)

## S4 method for signature 'tssObject'
getTitle(experimentName)
```

## Arguments

experimentName n S4 object of class *tssObject*

## Value

the contents of slot "title" are returned, which are of class "character"

## Examples

```
load(system.file("extdata", "tssObjectExample.RData",
  package="TSRchitect"))
example.title <- getTitle(experimentName=tssObjectExample)
example.title
```

**getTSRdata**

**getTSRdata**

## Description

an accessor function that retrieves the contents of a specified slot "tsrData"/"tsrDataMerged" from a given *tssObject*

## Usage

```
getTSRdata(experimentName, slotType, slot)

## S4 method for signature 'tssObject,character,numeric'
getTSRdata(experimentName,
  slotType = c("replicates", "merged"), slot)
```

**Arguments**

experimentName	an S4 object of class <i>tssObject</i>
slotType	'character' which data type is to be selected. Either "replicates" (tsrCountData) or "merged" (tsrCountDataMerged)
slot	'numeric' a number corresponding to the slot in "tsrData"/"tsrDataMerged" to be retrieved.

**Value**

the contents of the selected slot (either "tsrData" or "tsrDataMerged" are returned)

**Examples**

```
load(system.file("extdata", "tssObjectExample.RData",
  package="TSRchitect"))
ex.tsrData <- getTSRdata(experimentName=tssObjectExample,
  slotType="replicates", slot = 1)
ex.tsrData
```

getTSScountData

**getTSScountData****Description**

an accessor function that retrieves the contents of a specified slot "tssCountData"/"tssCountDataMerged" from a given *tssObject*

**Usage**

```
getTSScountData(experimentName, slotType, slot)

## S4 method for signature 'tssObject,character,numeric'
getTSScountData(experimentName,
  slotType = c("replicates", "merged"), slot)
```

**Arguments**

experimentName	an S4 object of class <i>tssObject</i>
slotType	'character' which data type is to be selected. Either "replicates" (tssCountData) or "merged" (tssCountDataMerged)
slot	'numeric' a number corresponding to the slot in "tssCountData"/"tssCountDataMerged" to be retrieved.

**Value**

the contents of the selected slot (either "tssCountData" or "tssCountDataMerged" are returned)

## Examples

```
load(system.file("extdata", "tssObjectExample.RData",
package="TSRchitect"))
ex.tssCountData <- getTSScountData(experimentName=tssObjectExample,
slotType="replicates", slot = 1)
ex.tssCountData
```

**getTSStagData**

**getTSStagData**

## Description

an accessor function that retrieves the contents of a specified slot "tssTagData" from a given *tssObject*

## Usage

```
getTSStagData(experimentName, slot)

## S4 method for signature 'tssObject,numeric'
getTSStagData(experimentName, slot)
```

## Arguments

experimentName	an S4 object of class <i>tssObject</i>
slot	'numeric' a number corresponding to the slot in "tssTagData" to be retrieved.

## Value

the contents of the specified slot "tssTagData" are returned

## Examples

```
load(system.file("extdata", "tssObjectExample.RData",
package="TSRchitect"))
example.tssTagData <- getTSStagData(experimentName=tssObjectExample, slot=1)
example.tssTagData
```

gsq2bed

**gsq2bed****Description**

gsq2bed converts aligned cDNA data (in .gsq format) to BED format, extracting the 5'-most base.

**Usage**

```
gsq2bed(gsqFile, outfile)

## S4 method for signature 'character,character'
gsq2bed(gsqFile, outfile)
```

**Arguments**

gsqFile	a path to the gsq (GeneSequer) output file (class character)
outfile	the name (class character) of the BED file to be written (default is "gsqOut.bed")

**Value**

a BED file containing a list of the 5'-most base from each of the alignments contained in the GeneSequer (.gsq) output file is written to the user's working directory.

**Examples**

```
extdata.dir <- system.file("extdata", package="TSRchitect")
tssObjectExample <- gsq2bed(gsqFile=paste(extdata.dir,"AtEST.gsq",sep="/"),
                             outfile="")
```

importAnnotationExternal

**importAnnotationExternal****Description**

importAnnotationExternal imports an annotation from an external file and attaches it to the *tssObject*

**Usage**

```
importAnnotationExternal(experimentName, fileType, annotFile)

## S4 method for signature 'tssObject,character,character'
importAnnotationExternal(experimentName,
                        fileType = c("bed", "gff", "gff3"), annotFile)
```

### Arguments

- `experimentName` - an S4 object of class *tssObject* that contains information about the experiment  
`fileType` - the format of the annotation file to be imported. Must be one of: "bed", "gff" or "gff3".  
`annotFile` - a path (full or relative) to the annotation file to be imported.

### Value

fills the slot `@annotation` in the returned *tssObject* with a [GRanges](#) object containing a parsed annotation file of the selected type.

### Note

`importAnnotationExternal` makes use of three functions from the *rtracklayer* package: [import.bed](#), [import.gff](#), and [import.gff3](#).

An example similar to the one provided can be found in *Example 2* from the vignette (run `TSRchitectUsersGuide()` to view the documentation).

To import directly from the AnnotationHub client, please refer to [importAnnotationHub](#).

### Examples

```
load(system.file("extdata", "tssObjectExample.RData", package="TSRchitect"))
extdata.dir <- system.file("extdata", package="TSRchitect")
annotation <- dir(extdata.dir, pattern="\\.gff3$", full.names=TRUE)
tssObjectExample <- importAnnotationExternal(experimentName=tssObjectExample,
                                              fileType="gff3", annotFile=annotation)
```

[importAnnotationHub](#)    **importAnnotationHub**

### Description

imports an annotation directly from AnnotationHub and attaches it to the *tssObject*

### Usage

```
importAnnotationHub(experimentName, provider, annotType, species, annotID)
```

```
## S4 method for signature 'tssObject,character,character,character,character'
importAnnotationHub(experimentName,
                   provider, annotType, species, annotID)
```

### Arguments

- `experimentName` - an S4 object of class *tssObject* that contains information about the experiment  
`provider` - 'character' the source of the annotation in AnnotationHub (e.g. 'gencode')  
`annotType` - 'character' the format of the annotationHub annotation to be imported. Using 'gff' will find annotations in both gff or gff3 formats  
`species` - 'character' the species identifier in AnnotationHub (e.g. 'human')  
`annotID` - 'character' the AnnotationHub identifier to be retrieved

**Value**

fills the slot `@annotation` in the returned `tssObject` with an `AnnotationHub` record. The record retrieved must be an object of class [GRanges](#).

**Note**

An example similar to the one provided can be found in the vignette ([/inst/doc/TSRchitect.Rmd](#))  
Please consult the available records in `AnnotationHub` beforehand using [AnnotationHub-class](#)

---

**inputToTSS****inputToTSS**

---

**Description**

`inputToTSS` extracts TSS information from each attached .bam or .bed file in a `tssObject` object

**Usage**

```
inputToTSS(experimentName)

## S4 method for signature 'tssObject'
inputToTSS(experimentName)
```

**Arguments**

`experimentName` an S4 object of class `tssObject` with bam files loaded

**Value**

produces a [GRangesList](#) containing separate [GRanges](#) objects for each .bam file contained within `experimentName`, placing them them in the returned `tssObject`.

**Note**

An example similar to the one provided can be found in the vignette ([/inst/doc/TSRchitect.Rmd](#)).

**Examples**

```
load(system.file("extdata", "tssObjectExample.RData",
package="TSRchitect"))
tssObjectExample <- inputToTSS(experimentName=tssObjectExample)
```

---

**loadTSSobj****loadTSSobj**

---

**Description**

`loadTSSobj` processes alignment files in .bam or .bed formats from the local directory supplied.

**Usage**

```
loadTSSobj(experimentTitle, inputDir, ...)

## S4 method for signature 'character,character'
loadTSSobj(experimentTitle, inputDir,
  n.cores = 1, isPairedBAM = TRUE, isPairedBED = TRUE,
  sampleSheet = NA, sampleNames = NA, replicateIDs = NA)
```

**Arguments**

<code>experimentTitle</code>	a descriptive title for the experiment (character).
<code>inputDir</code>	path to the directory containing the alignment files (in either .bam or .bed formats) (character). Note that all the paths to all files in <code>inputDir</code> with the extension .bam or .bed will be imported with this function.
<code>n.cores</code>	the number of cores to be used for this job. (numeric)
<code>isPairedBAM</code>	if the input is in BAM format, specifies whether the TSS profiling experiment is paired-end (if TRUE) or single-end (if FALSE). Set to TRUE by default. (logical)
<code>isPairedBED</code>	if the input is in BED format, specifies whether the TSS profiling experiment is paired-end (if TRUE) or single-end (if FALSE). Set to TRUE by default. (logical) Note: if TRUE, the input data must be in bedpe format, as described here: <a href="http://bedtools.readthedocs.io/en/latest/content/general-usage.html">http://bedtools.readthedocs.io/en/latest/content/general-usage.html</a>
<code>sampleSheet</code>	file providing TSS sample information; if provided, <code>sampleNames</code> and <code>replicateIDs</code> are ignored; input format is tab-delimited 3-column rows with sample name, replicate ID, and sample data file name; the file has to include column headers "SAMPLE ReplicateID FILE" and can be either tab-delimited or an Excel spreadsheet (file extension ".xls" or ".xlsx" (character)
<code>sampleNames</code>	unique labels of class character for each TSS sample within the experiment (character).
<code>replicateIDs</code>	identifiers indicating which samples are biological replicates. Note that <code>loadTSSobj</code> imports alignment data in ascending alphanumeric order, so the arguments to <code>replicateIDs</code> must be arranged in this order also so that they directly correspond to the intended file (numeric).

**Value**

`loadTSSobj` fills the slot `bamDataFirstRead` and/or `bedData` on the returned `tssObject` with `GAlignments` objects (for .bam files), or `GRanges` objects (for .bed files).

### Note

An example similar to the one provided can be found in the vignette (*/inst/doc/TSRchitect.Rmd*).

All files found in *inputDir* will be retrieved and written in ascending alphanumeric order to the *@fileNamesBAM* and/or *@fileNamesBED* slot(s) on the *tssObject* that is created.

### Examples

```
extdata.dir <- system.file("extdata/bamFiles", package="TSRchitect")
test.Obj <- loadTSSobj(experimentTitle="Code example", inputDir=extdata.dir,
n.cores=2, isPairedBAM=TRUE, sampleNames=c("sample1rep1", "sample1rep2",
"sample2rep1", "sample2rep2"), replicateIDs=c(1,1,2,2))
```

**makeGRangesFromTSR**

**makeGRangesFromTSR**

### Description

`makeGRangesFromTSR` creates a *GRanges* object from a specified TSR data set

### Usage

```
makeGRangesFromTSR(experimentName, tsrSetType, tsrSet = 1)

## S4 method for signature 'tssObject,character,numeric'
makeGRangesFromTSR(experimentName,
tsrSetType, tsrSet = 1)
```

### Arguments

<code>experimentName</code>	an S4 object of class <i>tssObject</i> containing information in slot <i>@tssTagData</i>
<code>tsrSetType</code>	specifies the set to be written to file. Options are "replicates" or "merged". (character)
<code>tsrSet</code>	number of the dataset to be processed (numeric).

### Value

An object of class *GRanges* containing the specified TSR data set. Headers include 'seqnames', 'ranges' (including start and end), 'strand', 'name' (TSR ID) and 'score' (Shape Index/SI) value

### Note

For more information on the *GRanges* class, please visit: [http://web.mit.edu/~r/current/arch/i386\\_linux26/lib/R/library/GRanges-class.html](http://web.mit.edu/~r/current/arch/i386_linux26/lib/R/library/GRanges-class.html)

### Examples

```
load(system.file("extdata", "tssObjectExample.RData", package="TSRchitect"))
makeGRangesFromTSR(experimentName=tssObjectExample, tsrSetType="replicates",
tsrSet=1)
```

mergeSampleData

**mergeSampleData****Description**

`mergeSampleData` combines samples from multiple TSS experiments into a single [GRanges](#) object

**Usage**

```
mergeSampleData(experimentName, n.cores, tagCountThreshold)

## S4 method for signature 'tssObject,numeric,numeric'
mergeSampleData(experimentName,
  n.cores = 1, tagCountThreshold = 1)
```

**Arguments**

<code>experimentName</code>	an S4 object of class <i>tssObject</i> that contains information about the experiment.
<code>n.cores</code>	the number of cores to be used for this job. ncores=1 means serial execution of function calls (numeric)
<code>tagCountThreshold</code>	the number of TSSs required at a given position for it to be considered in sample data merging. (numeric) Note: Merged data sets can become very large when the tagCountThreshold is set low (leading to inclusion of a lot of "noise" and resulting in long execution times in the necessary TSS position ordering step).

**Value**

`tssCountData` datasets are merged (according to the *sampleIDs*) and put in the `tssCountDataMerged` slot in the returned *tssObject*.

**Note**

An example similar to the one provided can be found in the vignette (*/inst/doc/TSRchitect.Rmd*).

**Examples**

```
load(system.file("extdata", "tssObjectExample.RData",
  package="TSRchitect"))
tssObjectExample <- mergeSampleData(experimentName=tssObjectExample,
  n.cores=1, tagCountThreshold=1)
```

## processTSS

## processTSS

## Description

`processTSS` calculates the number of observed reads at a given TSS coordinate across an entire dataset.

## Usage

```
processTSS(experimentName, n.cores, tssSet, writeTable)

## S4 method for signature 'tssObject,numeric,character,logical'
processTSS(experimentName,
           n.cores = 1, tssSet = "all", writeTable = FALSE)
```

## Arguments

<code>experimentName</code>	an S4 object of class <i>tssObject</i> containing information in slot <code>@tssTagData</code>
<code>n.cores</code>	the number of cores to be used for this job. <code>n.cores=1</code> means serial execution of function calls (numeric)
<code>tssSet</code>	default is "all"; to select a single <i>tssSet</i> , specify it (as character)
<code>writeTable</code>	specifies whether the output should be written to a file. (logical)

## Value

Creates a list of [GenomicRanges](#) containing TSS positions in slot *tssTagData* of the returned *tssObject*.

Note

Note that the `tssSet` parameter must be of class `character`, even when selecting an individual dataset. An example similar to the one provided can be found in the vignette (`/inst/doc/TSRchitect.Rmd`).

## Examples

```
load(system.file("extdata", "tssObjectExample.RData",
  package="TSRchitect"))
tssObjectExample <- processTSS(experimentName=tssObjectExample, n.cores=1,
  tssSet="all", writeTable=FALSE)
```

**TSRchitectUsersGuide** **TSRchitectUsersGuide**

### Description

Opens the TSRchitect User's Guide in a pdf viewer on the user's system.

### Usage

```
TSRchitectUsersGuide(view = TRUE)
```

### Arguments

view	(logical) if TRUE (default) the User's Guide is opened in the local pdf viewer. If FALSE then the full path to the User's Guide is returned.
------	---

### Value

if view=FALSE, then the full path to the User's Guide is returned.

### Examples

```
myPath <- TSRchitectUsersGuide(view=FALSE)
```

**tssObject**

**tssObject**

### Description

S4 constructor function for *tssObject*

### Usage

```
tssObject(title = NA, bamDataFirstRead = NA, bamDataLastRead = NA,
          bedData = NA)
```

### Arguments

title	'character' A short descriptive title for the experiment. Is set to NA by default.
bamDataFirstRead	'list' the name of a list of <b>GAlignments</b> objects (originating from .bam files) in the workspace. Set to NA by default.
bamDataLastRead	'list' the name of a list of <b>GAlignments</b> objects (originating from paired-read .bam files) in the workspace. Set to NA by default.
bedData	'list' the name of a list of <b>GRanges</b> or <b>Pairs</b> objects (originating from .bed files) in the workspace. Set to NA by default.

**Value**

a new *tssObject* is returned to the user's workspace.

**Examples**

```
new.tssObj <- tssObject(title="Example")
```

---

writeTSR	writeTSR
----------	----------

---

**Description**

`writeTSR` writes identified TSRs from a specified data set to a file in either tab or BED formats

**Usage**

```
writeTSR(experimentName, tsrSetType, tsrSet = 1, tsrLabel = "TSR_",
         mixedorder = FALSE, fileType = "tab")

## S4 method for signature
## 'tssObject,character,numeric,character,logical,character'
writeTSR(experimentName,
        tsrSetType, tsrSet = 1, tsrLabel = "TSR_", mixedorder = FALSE,
        fileType = "tab")
```

**Arguments**

experimentName	an S4 object of class <i>tssObject</i> containing information in slot <code>@tssTagData</code>
tsrSetType	specifies the set to be written to file. Options are "replicates" or "merged". (character)
tsrSet	number of the dataset to be processed (numeric).
tsrLabel	specifies the label to be used in the name column of BED format output
mixedorder	a logical specifying whether the sequence names should be ordered alphanumerically ("10" following "9" rather than "1"). (logical)
fileType	the format of the file to be written. Possible choices are "tab" for tab-delimited output, "bed" for BED format, and "gff" for GFF3 format (character).

**Value**

A table containing the specified TSR data set that is to be written to your working directory.

**Note**

The .bed file written adheres to the standard six-column BED format, while "tab" format is identical to that of the data.frames containing TSR data.

For more information on the BED format, please visit <https://genome.ucsc.edu/FAQ/FAQformat#format1>

## Examples

```
load(system.file("extdata", "tssObjectExample.RData", package="TSRchitect"))
writeTSS(experimentName=tssObjectExample, tsrSetType="replicates",
         tsrSet=1, tsrLabel="TSRsampel1_", mixedorder=FALSE, fileType="tab")
```

**writeTSS**

**writeTSS**

## Description

`writeTSS` writes identified TSSs from a specified data set to a file in either tab or BED formats

## Usage

```
writeTSS(experimentName, tsrSetType, tsrSet = 1, tsrLabel = "TSS_",
         mixedorder = FALSE, fileType = "tab")

## S4 method for signature
## 'tssObject,character,numeric,character,logical,character'
writeTSS(experimentName,
        tsrSetType, tsrSet = 1, tsrLabel = "TSS_", mixedorder = FALSE,
        fileType = "tab")
```

## Arguments

<code>experimentName</code>	an S4 object of class <i>tssObject</i> containing information in slot <code>@tssCountData</code>
<code>tsrSetType</code>	specifies the set to be written to file. Options are "replicates" or "merged". (character)
<code>tsrSet</code>	number of the dataset to be processed (numeric).
<code>tsrLabel</code>	specifies the label to be used in the name column of BED format output
<code>mixedorder</code>	a logical specifying whether the sequence names should be ordered alphanumerically ("10" following "9" rather than "1"). (logical)
<code>fileType</code>	the format of the file to be written. Possible choices are "tab" for tab-delimited output, "bed" for BED format, and "bedGraph" for GFF3 format (character).

## Value

A table containing the specified TSS data set that is to be written to your working directory.

## Note

The .bed file written adheres to the standard six-column BED format, while "tab" format is identical to that of the data.frames containing TSS data.

For more information on the BED format, please visit <https://genome.ucsc.edu/FAQ/FAQformat#format1>

**Examples**

```
load(system.file("extdata", "tssObjectExample.RData", package="TSRchitect"))
writeTSS(experimentName=tssObjectExample, tssSetType="replicates",
         tssSet=1, tssLabel="TSSsample1_", mixedorder=FALSE, fileType="tab")
```

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