

# Package ‘PowerExplorer’

April 16, 2019

**Title** Power Estimation Tool for RNA-Seq and proteomics data

**Version** 1.2.2

**URL** <https://gitlab.utu.fi/CompBioMedNGSTools/PowerExplorer>

**Description** Estimate and predict power among groups and multiple sample sizes with simulated data, the simulations are operated based on distribution parameters estimated from the provided input dataset.

**biocViews** ImmunoOncology, RNASeq, Proteomics, DifferentialExpression, MultipleComparison, Sequencing, Coverage, ChIPSeq

**Depends** R (>= 3.5.0), SummarizedExperiment

**Imports** DESeq2, ROTS, vsn, stats, utils, methods, gridExtra, MASS, data.table, ggplot2, Biobase, S4Vectors, BiocParallel

**Suggests** knitr, rmarkdown

**VignetteBuilder** knitr

**License** Artistic-2.0

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 6.1.1

**git\_url** <https://git.bioconductor.org/packages/PowerExplorer>

**git\_branch** RELEASE\_3\_8

**git\_last\_commit** d2cb645

**git\_last\_commit\_date** 2019-01-04

**Date/Publication** 2019-04-15

**Author** Xu Qiao [aut, cre],  
Laura Elo [cph]

**Maintainer** Xu Qiao <xu.qiao@utu.fi>

## R topics documented:

estimatePower . . . . .	2
exampleObject . . . . .	3
exampleProteomicsData . . . . .	4
exampleRNASeqData . . . . .	4
listEstPwr . . . . .	5

listPredPwr . . . . .	5
plotEstPwr . . . . .	6
plotPredPwr . . . . .	7
PowerExplorerStorage-class . . . . .	7
predictPower . . . . .	8
show,PowerExplorerStorage-method . . . . .	9

**Index****10**


---

<b>estimatePower</b>	<i>Estimate Power of the Actual Data</i>
----------------------	--

---

**Description**

Estimate power of comparison between each two groups based on the data simulated from estimated normal distributions of entries in the entire dataset

**Usage**

```
estimatePower(inputObject, groupVec, isLogTransformed = FALSE,
  dataType = c("RNASeq", "Proteomics"), minLFC = 0.5, alpha = 0.05,
  ST = 100, seed = 123, enableROTS = FALSE, paraROTS = list(B =
  1000, K = NULL, paired = FALSE, a1 = NULL, a2 = NULL, progress = FALSE),
  showProcess = FALSE, saveResultData = FALSE, parallel = FALSE,
  BPPARAM = bpparam())
```

**Arguments**

<b>inputObject</b>	a numeric raw data matrix or SummarizedExperiment object
<b>groupVec</b>	a vector indicating the grouping of samples
<b>isLogTransformed</b>	logical; set to TRUE, if the input data is log transformed.
<b>dataType</b>	"RNASeq" or "Proteomics" indicates the data type of the input data matrix.
<b>minLFC</b>	the threshold for log2 fold change, entries with lower LFC are not included in the power calculation, set to 0 if no threshold is needed.
<b>alpha</b>	controlled false positive rate.
<b>ST</b>	the number of simulations of abundance data generation and repeated times of statistical test for each entry (>=100 recommended).
<b>seed</b>	an integer seed for the random number generator.
<b>enableROTS</b>	logical; if TRUE, Reproducibility-Optimized Test Statistic (ROTS) will be used as the statistical model. used as the statistical model.
<b>paraROTS</b>	a list object containing additional parameters passed to ROTS (if enabled), see <a href="#">ROTS</a> .
<b>showProcess</b>	logical; if TRUE, show the detailed information of each simulation, used for debugging only.
<b>saveResultData</b>	logical; if TRUE, save the simulated data into RData with name pattern "simulated_Data_numRep_X_numSim_XXX_XXXXXX.RData"
<b>parallel</b>	logical; if FALSE parallelization is disabled; if TRUE, parallelize calculations using <a href="#">BiocParallel</a> .
<b>BPPARAM</b>	an optional argument object passed <a href="#">bplapply</a> to indicate the registered cores, if parallel=TRUE.

**Value**

a list of power estimates grouped in comparisons between each two groups

**See Also**

[predictPower](#) predict power with increasing sample sizes

**Examples**

```
# Example 1: a random generated Proteomics dataset (10 DE, 100 non-DE)
# Note: Simulation times(ST) is specified as 10 for shorter example runtime,
# ST > 50 is recommended
data(exampleProteomicsData)
dataMatrix <- exampleProteomicsData$dataMatrix
groupVec <- exampleProteomicsData$groupVec

# Run estimation without LFC filtration
resObject <- estimatePower(dataMatrix, groupVec,
                           dataType="Proteomics",
                           isLogTransformed=FALSE,
                           minLFC=0, alpha=0.05,
                           ST=10, seed=123)
```

---

exampleObject

*An Example Predicted Power Object For Tests*

---

**Description**

This is an example PowerExplorerStorage resulted from an example run on a publically available RNA-Seq database containing the gene expression in C57BL/6J and DBA/2J Mouse Striatum

**Usage**

```
data(exampleObject)
```

**Format**

An object of class PowerExplorerStorage with 12152 rows and 21 columns.

**Examples**

```
data(exampleObject)

show(exampleObject)
plotEstPwr(exampleObject)
```

---

exampleProteomicsData *Randomly Generated Proteomics Dataset*

---

### Description

This is a randomly generated proteomics dataset with 130 protein entries (rows) and 15 samples (columns) in 3 sample groups A, B and C, the log2 fold change (LFC) between group B and A is specified as 1, between C and B is also 1, thus the LFC is 2 between C and A.

### Usage

```
data(exampleProteomicsData)
```

### Format

An list contains "dataMatrix" and "groupVec"

### Examples

```
data(exampleProteomicsData)
head(exampleProteomicsData$dataMatrix)
```

---

exampleRNASeqData *Randomly Generated RNASeq Dataset*

---

### Description

This is a randomly generated RNASeq dataset with 130 gene entries (rows) and 15 samples (columns) in 3 sample groups A, B and C, the log2 fold change (LFC) between group B and A is specified as 1, between C and B is also 1, thus the LFC is 2 between C and A.

### Usage

```
data(exampleRNASeqData)
```

### Format

An list contains "dataMatrix" and "groupVec"

### Examples

```
data(exampleRNASeqData)
head(exampleRNASeqData$dataMatrix)
```

---

**listEstPwr***List Estimated Power*

---

**Description**

show a top-list of power in numerical order, or list the power selected genes/proteins.

**Usage**

```
listEstPwr(inputObject, decreasing = TRUE, top = 20, selected = NA)
```

**Arguments**

- |             |   |
|-------------|---|
| inputObject | the input inputObject.  |
| decreasing  | logical; TRUE, decreasing order; FALSE, increasing order.   |
| top         | the number of genes/proteins in the top list  |
| selected    | default as NA; specify as a list of geneID or protein ID to show power of a list of interested records. |

**Value**

a top list of power / power of a list of interested genes or proteins

**Examples**

```
data(exampleObject)
# show 10 top genes with high power (decreasing order)
listEstPwr(exampleObject, decreasing = TRUE, top = 10)
# show a list of interested genes
listEstPwr(exampleObject,
           selected = c("ENSMUSG00000000303",
                        "ENSMUSG00000087272",
                        "ENSMUSG00000089921"))
```

---

**listPredPwr***List Predicted Power*

---

**Description**

show a top-list of predicted power in numerical order, or list the power selected genes/proteins.

**Usage**

```
listPredPwr(inputObject, decreasing = TRUE, top = 20, selected = NA)
```

**Arguments**

<code>inputObject</code>	the input <code>inputObject</code> .
<code>decreasing</code>	logical; TRUE, decreasing order; FALSE, increasing order.
<code>top</code>	the number of genes/proteins in the top list
<code>selected</code>	default as NA; specify as a list of geneID or protein ID to show power of a list of interested records.

**Value**

a top list of power / power of a list of interested genes or proteins for each sample size

**Examples**

```
data(exampleObject)
# show 10 top genes with high power (decreasing order)
listPredPwr(exampleObject, decreasing = TRUE, top = 10)

# show a list of interested genes
listPredPwr(exampleObject,
            selected = c("ENSMUSG0000000303",
                         "ENSMUSG00000087272",
                         "ENSMUSG00000089921"))
```

plotEstPwr

*Plot A Summary of Estimated Power***Description**

Produce a plot to summary the power estimated by function `estimatePower`, the plot function is called in `estimatePower`, but using it manually is possible

**Usage**

```
plotEstPwr(inputObject)
```

**Arguments**

<code>inputObject</code>	a result container object <code>PowerExplorerStorage</code> returned from <code>estimatePower</code> .
--------------------------	--

**Value**

plot(s) of the summarised information on the estimated power

**Examples**

```
data(exampleObject)
plotEstPwr(exampleObject)
```

**plotPredPwr***Observe Predicted Power Within Different LFC Scales***Description**

With a complete power list and LFC list returned from [predictPower](#), power estimates can be observed dynamically within specified LFC ranges.

**Usage**

```
plotPredPwr(inputObject, minLFC, maxLFC, LFCscale = 1)
```

**Arguments**

<code>inputObject</code>	a result container object <a href="#">PowerExplorerStorage</a>
<code>minLFC</code>	default as 0, the left edge of the LFC range within which genes will be included in the graph.
<code>maxLFC</code>	default as NA (the maximum value in data will be used) the right edge of the LFC range within which genes will be included in the graph.
<code>LFCscale</code>	the size of each unit when segmenting predicted power by LFC

**Value**

plot(s) of power density under multiple sample sizes

**Examples**

```
# load an example object containing
# predicted power under different sample sizes
data(exampleObject)
plotPredPwr(exampleObject)
plotPredPwr(exampleObject)
# It is possible to observe power trend in different scales and ranges of LFCs
plotPredPwr(exampleObject, minLFC=0, maxLFC=2, LFCscale=0.5)
```

**PowerExplorerStorage-class***PowerExplorer Object***Description**

An extended `SummarizedExperiment` object to contain input `dataMatrix`, grouping information, estimated power, predicted power, fold change estimates and other estimation parameters.

**predictPower***Estimate Power Under Increasing Sample Sizes***Description**

Similiar to [estimatePower](#), power estimations are performed under multiple increasing sample sizes

**Usage**

```
predictPower(inputObject, groupVec, isLogTransformed = FALSE,
            dataType = c("RNASeq", "Proteomics"), enableROTS = FALSE,
            paraROTS = list(B = 1000, K = NULL, paired = FALSE, a1 = NULL, a2 =
                           NULL, progress = FALSE), minLFC = 0.5, rangeSimNumRep = NA,
            alpha = 0.05, ST = 100, seed = 123, parallel = FALSE,
            BPPARAM = bpparam(), showProcess = FALSE, saveResultData = FALSE)
```

**Arguments**

<code>inputObject</code>	a numeric raw Proteomics abundance data matrix, in which rows correspond to proteins and columns correspond to samples.
<code>groupVec</code>	a vector indicating the grouping of samples
<code>isLogTransformed</code>	logical; logical; set to TRUE, if the input data is log transformed.
<code>dataType</code>	"RNASeq" or "Proteomics" indicates the data type of the input data matrix.
<code>enableROTS</code>	logical; if TRUE, Reproducibility-Optimized Test Statistic (ROTS) will be used as the statistical model.
<code>paraROTS</code>	a list object containing additional parameters passed to ROTS (if enabled), see <a href="#">ROTS</a> .
<code>minLFC</code>	LFC threshold
<code>rangeSimNumRep</code>	a vector of sample sizes under which power will be estimated
<code>alpha</code>	controlled false positive rate.
<code>ST</code>	the number of simulations of abundance data generation and repeated times of statistical test for each protein (>=100 recommended).
<code>seed</code>	an integer seed for the random number generator.
<code>parallel</code>	logical; if FALSE parallelization is disabled; if TRUE, parallelize calculations using <a href="#">BiocParallel</a> .
<code>BPPARAM</code>	an optional argument object passed <a href="#">bplapply</a> to indicate the registered cores, if <code>parallel=TRUE</code> .
<code>showProcess</code>	logical; if TRUE, show the detailed information of each simulation, used for debugging only.
<code>saveResultData</code>	logical; if TRUE, save the simulated data into RData with name pattern "simulated_Data_numRep_X_numSim_XXX_XXXXX.RData".

**Value**

a list of power predictions for each sample size, grouped in comparisons between each two groups

**See Also**

[estimatePower](#) estimate power based on actual data

**Examples**

```
# Example 1: a random generated Proteomics dataset (10 DE, 100 non-DE)
data(exampleProteomicsData)
dataMatrix <- exampleProteomicsData$dataMatrix
groupVec <- exampleProteomicsData$groupVec

# Run estimation
# Note: Simulation times(ST) is specified as 5 for shorter example runtime
#       For better performance, ST > 50 is recommended
predictedPower <- predictPower(dataMatrix, groupVec,
                                 isLogTransformed=FALSE,
                                 dataType="Proteomics",
                                 minLFC=0,
                                 rangeSimNumRep=c(5, 10, 15),
                                 alpha=0.05, ST=5, seed=123)
```

show,PowerExplorerStorage-method

*show method for PowerExplorerStorage*

**Description**

show method for PowerExplorerStorage

**Usage**

```
## S4 method for signature 'PowerExplorerStorage'
show(object)
```

**Arguments**

object            a PowerExplorerStorage object as input

**Value**

a summary of input PowerExplorerStorage object

**Methods (by class)**

- PowerExplorerStorage: method for PowerExplorerStorage objects

**Examples**

```
data(exampleObject)
show(exampleObject)
```

# Index

\*Topic **datasets**  
  exampleObject, [3](#)  
  exampleProteomicsData, [4](#)  
  exampleRNASeqData, [4](#)

BiocParallel, [2](#), [8](#)  
bplapply, [2](#), [8](#)

estimatePower, [2](#), [6](#), [8](#), [9](#)  
exampleObject, [3](#)  
exampleProteomicsData, [4](#)  
exampleRNASeqData, [4](#)

listEstPwr, [5](#)  
listPredPwr, [5](#)

plotEstPwr, [6](#)  
plotPredPwr, [7](#)  
PowerExplorerStorage, [6](#), [7](#)  
PowerExplorerStorage  
  (PowerExplorerStorage-class), [7](#)  
PowerExplorerStorage-class, [7](#)  
predictPower, [3](#), [7](#), [8](#)

ROTS, [2](#), [8](#)

show, PowerExplorerStorage-method, [9](#)