Package 'RNAither'

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Title Statistical analysis of high-throughput RNAi screens

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Description RNAither analyzes cell-based RNAi screens, and includes
quality assessment, customizable normalization and statistical
tests, leading to lists of significant genes and biological
processes.

License Artistic-2.0

biocViews CellBasedAssays, QualityControl, Preprocessing,
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NeedsCompilation no

R topics documented:

RNAither-package   *Statistical analysis of high-throughput RNAi screens*

Description

RNAither analyzes cell-based RNAi screens, and includes quality assessment, customizable normalization and statistical tests, leading to lists of significant genes and biological processes.

Details

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Author(s)

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BScore   *BScore normalization*

Description

Normalization with BScores (see References).

Usage

```
BScore(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
listOfArgs	a list containing:
	- a character string specifying the column whose values will be used for normalization
	- a flag specifying whether controls should be excluded for the computation of the median polish (1) or not (0)

Value

A list containing:

header	The new header (with an added entry about the normalization procedure in the comments)
dataset	The new dataset with normalized values. The old values are saved in an extra column of the dataset with the suffix ".old"

References

C. Brideau, B. Gunter, B. Pikounis, and A. Liaw. Improved statistical methods for hit selection in high-throughput screening. J Biomol Screen, 8:634-647, 2003

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")
normres <- BScore(header, dataset, list("SigIntensity", 0))
newheader <- normres[[1]]
newdataset <- normres[[2]]
```

channelPlot	*Plot signal channels against each other*

Description

Generates plots allowing pairwise comparison of signal channels. Fits a lowess regression curve into the plots.

Usage

```
channelPlot(header, dataset, vecOfChannels, flag, plotTitle, showPlot, smSpan=2/3)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
vecOfChannels	A vector containing the names of the signal channels to be compared, e.g. "Sig-Intensity"
flag	0, 1, or 2. 0 uses the data from the complete dataset, 1 makes comparisons for each experiment, 2 makes comparisons for each plate.
plotTitle	The plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.
smSpan	The smoother span of the lowess curve. This gives the proportion of points in the plot which influence the smooth at each value. Larger values give more smoothness. Optional, defaults to 2/3

Value

Saves the plots in pdf and png files named after the experiment name specified in the header concatenated with the `plotTitle`, the number of the comparison, and if applicable the experiment number and/or the plate number.

When `flag == 0`, returns the plot name (`plotName`).

When `flag == 1`, returns a list containing:

plotName	The plot name
minOfScreens	The number of the first experiment
numOfScreens	The number of the last experiment

When `flag == 2`, returns a list containing: the plot name, a vector with the number of the first experiment and of the last experiment, and a vector with the number of the first plate and the number of the last plate.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")
plotname <- channelPlot(header, dataset, c("SigIntensity", "NbCells"), 0, "Channel comparison", 1)
```

---

closestToZero   *Return the replicate value closest to zero*

Description

Out of a set of replicate values, returns the one closest to zero.

Usage

```
closestToZero(Ivec, na.rm = T)
```

Arguments

Ivec	All channel values for a specific siRNA/gene
na.rm	Removes NA values

Value

A double giving the value closest to zero out of the given replicate values.

See Also

[rms](), [trim](), [furthestFromZero](), [summarizeReps](), [summarizeRepsNoFiltering]()

Examples

```
data(exampleDataset, package="RNAither")
Indexes <- findReplicates(dataset, "GeneName", "CPSF1")
replicateclosest <- closestToZero(dataset$SigIntensity[Indexes])
```

---

compareHits   *Searching for common hits between different scoring methods*

Description

Searches for common hits between different scoring methods.

Usage

```
compareHits(hitVec1, hitVec2, namesHitVec1, namesHitVec2)
```

Arguments

hitVec1, hitVec2
> the two binary hit vectors to be compared

namesHitVec1, namesHitVec2
> the names of the siRNAs corresponding to the hit vectors

Value

Returns a character vector indicating which siRNAs are identified as hits in two different hit scoring schemes.

See Also

[vennDiag](#) , [Ttest](#), [MannWhitney](#)

Examples

```
data(scoredDataset1, package="RNAither")
data(pValVec1, package="RNAither")

data(scoredDataset2, package="RNAither")
data(pValVec2, package="RNAither")

##for details on the generation of pValVec and scoredDataset,
##see the examples of the functions Ttest and MannWhitney linked above.

scoredHits1 <- hitselectionPval(scoredDataset1, pValVec1, "SigIntensity", "Hits1", 0.05,
"GeneName", "pvalue_testfile1.txt")
scoredHits2 <- hitselectionPval(scoredDataset2, pValVec2, "SigIntensity", "Hits2", 0.05,
"GeneName", "pvalue_testfile2.txt")

hitVector1 <- scoredHits1[[2]]
hitVector2 <- scoredHits2[[2]]

common_hits <- compareHits(hitVector1, hitVector2, names(hitVector1), names(hitVector2))
```

---

compareReplicaPlates   *Compare replica plates*

Description

Generates plots comparing the same plates in different experiments pairwise.

Usage

```
compareReplicaPlates(header, dataset, plotTitle, col4val, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
plotTitle	the plot title
col4val	a character string specifying the column whose values will be used for the plot
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

For each plate, plots of pairwise comparisons between replicate intensities are generated and saved as a pdf file named after the experiment name specified in the header concatenated with the plotTitle .

See Also

   [compareReplicates](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

compareReplicaPlates(header, dataset, "Comparison of replica plate", "SigIntensity", 1)
```

---

   compareReplicates   *Compare replicate values*

Description

   Plots replicate intensities pairwise for each experiment.

Usage

```
compareReplicates(header, dataset, plotTitle, col4val, col4anno, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
plotTitle	the plot title
col4val	a character string specifying the column whose values will be used for the plot
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

   For each experiment, plots of pairwise comparisons between replicate intensities are generated and saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle, and the number of the experiment.

   The function returns a list containing:

plotName	the plot name
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment
maxCombinationNum	
	the number of replicates to compare

See Also

   [compareReplicaPlates](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

compareReplicates(header, dataset, "Comparison of Replicates", "SigIntensity", "GeneName", 1, 0)
```

---

compareReplicateSD   *Plot the standard deviation of replicates*

Description

In the same fashion as spatialDistrib, generates a plot of the standard deviation of replicate values.

Usage

```
compareReplicateSD(header, dataset, plotTitle, colname4SD, col4anno, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
plotTitle	the plot title
colname4SD	a character string specifying the column whose values will be used for the computation of the replicate standard deviation
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

Generates a plot of the standard deviation of replicate values of all experiments. The plot is saved as a png file named after the experiment name specified in the header concatenated with the plotTitle.

Wells showing positive controls sd are marked with a "P", wells showing negative controls sd with an "N".

The plot will also be saved as an html file containing mouse-overs with the siRNA name for each well.

The function returns the plotname.

See Also

spatialDistrib, compareReplicateSDPerScreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

compareReplicateSD(header, dataset, "Replicate standard intensity deviation",
"SigIntensity", "GeneName", 1)
```

---

compareReplicateSDPerScreen

Plot the standard deviation of replicates for each experiment

---

Description

In the same fashion as spatialDistrib, generates plots of the standard deviation of replicate values for each experiment.

Usage

```
compareReplicateSDPerScreen(header, dataset, plotTitle, colname4SD, col4anno, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
plotTitle	the plot title
colname4SD	a character string specifying the column whose values will be used for the computation of the replicate standard deviation
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

Generates plots of the standard deviation of replicate values for each experiment. The plots are saved as png files named after the experiment name specified in the header concatenated with the plotTitle and the number of the experiment.

Wells showing positive controls sd are marked with a "P", wells showing negative controls sd with an "N".

The plots will also be saved as html files containing mouse-overs with the siRNA name for each well.

The function returns a list of length 3 containing:

basicPlotName	the plot name
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment

See Also

spatialDistrib, compareReplicateSD

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

compareReplicateSDPerScreen(header, dataset, "Replicate standard intensity deviation",
"SigIntensity", "GeneName", 1)
```

---

controlDensity   *Plotting the control density*

Description

Plots the density of positive and negative controls (if applicable) for all controls contained in the dataset.

Usage

```
controlDensity(header, dataset, channel, plotTitle, showPlot, supHisto)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the name of the column containing the values for computing the density, e.g. "SigIntensity"
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows
supHisto	0 or 1. 1 will additionally superimpose a colour histogram of the values for the positive and negative controls. Otherwise choose 0.

Value

Plots the density of positive and negative controls (if applicable) for all controls contained in the dataset. The plot is saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns the plot name.

See Also

controlDensityPerScreen, controlDensityPerPlate

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

controlDensity(header, dataset, "SigIntensity", "Control density", 1, 1)
```

controlDensityPerPlate

Plotting the control density per plate

Description

Plots the density of positive and negative controls (if applicable) for each plate.

Usage

```
controlDensityPerPlate(header, dataset, channel, plotTitle, plotDesign, showPlot, supHisto)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
channel	a character string specifying the name of the column containing the values for computing the density, e.g. "SigIntensity"
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows
supHisto	0 or 1. 1 will additionally superimpose a colour histogram of the values for the positive and negative controls. Otherwise choose 0.

Value

Generates a series of plots for each experiment and each plate, showing the density of positive and negative controls (if applicable). The plots are saved as pdf and png files named after the experiment name specified in the header concatenated with the plotTitle.

The function returns a list of length 3 containing:

plotName	the plot name
c(minOfScreens, numOfScreens)	
	a vector with the number of the first experiment and of the last experiment
c(minOfPlates, numOfPlates)	
	a vector with the number of the first plate and the number of the last plate

See Also

[controlDensity](#), [controlDensityPerScreen](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

controlDensityPerPlate(header, dataset, "SigIntensity", "Control density", 1, 1, 1)
```

controlDensityPerScreen

Plotting the control density per experiment

Description

Plots the density of positive and negative controls (if applicable) for each experiment.

Usage

```
controlDensityPerScreen(header, dataset, channel, plotTitle, showPlot, supHisto)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the name of the column containing the values for computing the density, e.g. "SigIntensity"
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows
supHisto	0 or 1. 1 will additionally superimpose a colour histogram of the values for the positive and negative controls. Otherwise choose 0.

Value

Generates a series of plots for each experiment, showing the density of positive and negative controls (if applicable). The plots are saved as pdf and png files named after the experiment name specified in the header concatenated with the plotTitle and the number of the experiment.

The function returns a list of length 3 containing:

plotName	the plotname
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment

See Also

controlDensity, controlDensityPerPlate

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

controlDensityPerScreen(header, dataset, "SigIntensity", "Control density", 1, 1)
```

controlNorm	*Normalization on controls*

Description

Performs a normalization on either positive or negative controls.

Usage

```
controlNorm(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
listOfArgs	a list containing: - a character string specifying the column whose values will be used for normalization - 1 or 2, 1 meaning a normalization per experiment, 2 meaning a normalization per plate - 0 or 1, 0 meaning a normalization on the median of negative controls, 1 meaning a normalization on the median of positive controls. Can also be the GeneName of a specific control siRNA - 1 or 2, 1 meaning the signal values are divided by the median, 2 meaning the median is subtracted from the signal values

Value

Returns a list containing:

header	the new header (with an added entry about the normalization procedure in the comments).
dataset	the new dataset with normalized values. The old values are saved in an extra column with the suffix ".old".

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- controlNorm(header, dataset, list(2, 0, "SigIntensity", 1))

newheader <- normres[[1]]
newdataset <- normres[[2]]
```

---

createSubset *Creating a subset of a dataset according to a certain column value*

Description

Creates a subset of a dataset containing all wells/lines having a certain value in a specified column.

Usage

```
createSubset(dataset, listIDs, equalTo)
```

Arguments

dataset	an R data frame generated with generateDatasetFile
listIDs	a character string and one of the following: Spotnumber, Internal_GeneID, GeneName, SpotTyp
equalTo	A value or character string specifying the value in the chosen column, e.g. all wells on plate 2

Value

A subset of the dataset containing only the wells/lines having a certain value in a specified column.

See Also

indexSubset

Examples

```
data(exampleDataset, package="RNAither")

subset <- createSubset(dataset, dataset$LabtekNb, 2)
```

---

dataset *a typical example RNAi dataset*

Description

See generateDatasetFile for details

Usage

```
dataset
```

Format

See generateDatasetFile

---

datasetDrosophila   *Genome-wide RNAi screen of cell viability in Drosophila Kc167 cells by Boutros et al.*

Description

M. Boutros et al., Genome-wide RNAi analysis of growth and viability in Drosophila cells, Science, 303(5659):832-835, 2004. 3, 18

Usage

```
datasetDrosophila
```

Format

see [generateDatasetFile](#) for details

---

discardLabtek   *Remove a complete plate from the analysis*

Description

Removes a plate/LabTek from the analysis by setting its spot type in the dataset to -1.

Usage

```
discardLabtek(data, screenNr, labtekNr)
```

Arguments

data	an R data frame generated with [generateDatasetFile](#)
screenNr	the number of the experiment that contains the plate to discard
labtekNr	the number of the plate to discard

Value

A new dataset that stil contains the specified plate/LabTek, but excludes it from the further analysis by setting its SpotTypes to -1.

See Also

[discardWells](#)

Examples

```
data(exampleDataset, package="RNAither")

newdataset <- discardLabtek(dataset, 2, 2)
```

---

discardWells   *Remove wells from the analysis*

Description

Removes wells from the analysis by setting their spot type in the dataset to -1.

Usage

```
discardWells(data, screenNr, labtekNr, vecPositions)
```

Arguments

data	an R data frame generated with [generateDatasetFile](#)
screenNr	the number of the experiment that contains the plate to discard
labtekNr	the number of the plate to discard
vecPositions	a vector specifying the numbers of the wells to discard

Value

A new dataset that does not contain the specified wells. A new dataset that stil contains the specified wells/spots, but excludes them from the further analysis by setting their SpotTypes to -1.

See Also

[discardLabtek](#)

Examples

```
data(exampleDataset, package="RNAither")

newdataset <- discardWells(dataset, 2, 1, c(1, 10, 15))
```

---

divideChannels   *Divide channel values*

Description

Replace two channels by their ratio.

Usage

```
divideChannels(ch1, ch2)
```

Arguments

ch1	a vector giving all values from channel 1
ch2	a vector giving all values from channel 2

Value

A vector of the ratio of channel 1 and channel 2.

See Also

[sumChannels](#)

Examples

```
data(exampleDataset, package="RNAither")

newch <- divideChannels(dataset$SigIntensity, dataset$NbCells)
```

---

divNorm   *Mean, median, ... , normalization*

Description

Normalization with the mean, median, or any other function.

Usage

```
divNorm(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
listOfArgs	a list containing:
	- a character string specifying the column whose values will be used for normalization
	- a function to be used for the normalization, e.g. mean, median, ...
	- 1 or 2, 1 meaning a normalization per experiment, 2 meaning a normalization per plate
	- 1 or 2, 1 meaning the normalization is achieved by a division of the intensity values by the outcome of funname, 2, meaning by a substraction
	- a flag specifying whether controls should be excluded for the computation of the result of the function specified in the first element (1) or not (0).

Value

Returns a list containing:

header	the new header (with an added entry about the normalization procedure in the comments)
dataset	the new dataset with normalized values. The old values are saved in an extra column with the suffix ".old"

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- divNorm(header, dataset, list(median, 2, 1, "SigIntensity", 1))

newheader <- normres[[1]]
newdataset <- normres[[2]]
```

DRQualControl   *Computing the dynamic range*

Description

Computes the dynamic range per plate for a complete dataset file and plots the results.

Usage

```
DRQualControl(header, data, nbLinesHeader, channel, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
data	an R data frame generated with [generateDatasetFile](#)
nbLinesHeader	typically 3
channel	A character string specifying the name of the column containing the values for computing the dynamic range, e.g. "SigIntensity"
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

Returns the dynamic range for each plate in the shell and saves them in a text file. The name of the text file will be the concatenation of the experiment name specified in the header and the character string "DR.txt".

Shows a plot of the dynamic range values and saves it as a pdf file under the experiment name specified in the header concatenated with the function argument plotTitle.

References

M. Boutros, L. Bras, and W. Huber. Analysis of cell-based RNAi screens. *Genome Biol*, 7(7): R66, 2006.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

DRQualControl(header, dataset, 3, "SigIntensity", "DR per plate", 1)
```

eraseDataSetColumn   *Remove columns from dataset*

Description

Removes a specified column from a dataset.

Usage

```
eraseDataSetColumn(dataset, colname)
```

Arguments

dataset   an R data frame generated with generateDatasetFile

colname   a character string specifying the name of the column to be removed

Value

An R data frame with the specified column removed.

Examples

```
data(exampleDataset, package="RNAither")

newdataset <- eraseDataSetColumn(dataset, "SDSIntensity")
```

findReplicates   *Find all replicates of a certain siRNA/gene in a dataset*

Description

Returns which lines in the dataset correspond to a given siRNA/gene ID.

Usage

```
findReplicates(dataset, whichCol, replicateID)
```

Arguments

dataset   an R data frame generated with generateDatasetFile

whichCol   a character string specifying the name of the column containing the ID, either
                 Internal_GeneID or GeneName

replicateID   the siRNA/gene ID of interest

Value

An integer vector containing the indexes in the main dataset of all wells corresponding to a given
siRNA/gene ID

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

Indexes <- findReplicates(dataset, "GeneName", "CPSF1")
```

---

furthestFromZero	*Return the replicate value furthest from zero*

Description

Out of a set of replicate values, returns the one furthest from zero.

Usage

```
furthestFromZero(Ivec, na.rm = T)
```

Arguments

Ivec	All channel values for a specific siRNA/gene
na.rm	Removes NA values

Value

A double giving the value furthest from zero out of the given replicate values.

See Also

[rms](#), [trim](#), [closestToZero](#), [summarizeReps](#), [summarizeRepsNoFiltering](#)

Examples

```
data(exampleDataset, package="RNAither")

Indexes <- findReplicates(dataset, "GeneName", "CPSF1")
replicateclosest <- furthestFromZero(dataset$SigIntensity[Indexes])
```

---

generateDatasetFile	*Generate Dataset File*

Description

Generates a text file containing all experimental data. Needed for all subsequent analysis functions.

Usage

```
generateDatasetFile(externalExperimentName, typeOfData, comments, outputFile,
plateLayoutInternal, plateLayoutNCBI, nbRowsPerPlate, nbColsPerPlate, screenNb_pre,
emptyWells, poorWells, controlCoordsOutput, backgroundValOutput, meanSignalOutput,
SDmeanSignal, objNumOutput, cellNumOutput)
```

Arguments

externalExperimentName

A character string specifying the experiment name, e.g. "Johns Experiment Nb. 1"

typeOfData   A character string specifying the type of data, e.g. "364 well plate data for virus screens"

comments   A character string specifying comments. NA if not available.

outputFile   A character string specifying the name of the text file containing the dataset.

plateLayoutInternal

A matrix of internal siRNA IDs specifying their position on the plate (row-wise). Each column of the matrix stands for one plate.

plateLayoutNCBI

A matrix of gene names specifying their position on the plate (row-wise). Each column of the matrix stands for one plate.

nbRowsPerPlate   The number of rows per plate

nbColsPerPlate   The number of columns per plate

screenNb_pre   The screen/experiment number

emptyWells   A list containing, for each plate, an integer vector of the positions of empty wells. NA if there are no empty wells on the plate.

poorWells   A list containing, for each plate, an integer vector of the positions of wells that, for a certain reason, should not be taken into account during the analysis. NA if there are no such wells on the plate.

controlCoordsOutput

A list containing, for each plate, a list of integer vectors specifying the positions of positive (first element in sublist) and negative (second element in sublist) controls. NA if there are no positive/negative controls on the plate.

backgroundValOutput

A list containing, for each plate, a vector of background values per well

meanSignalOutput

A list containing, for each plate, a vector of intensity values for each well

SDmeanSignal   A list containing, for each plate, a vector of standard deviations of intensity values for each well

objNumOutput   A list containing, for each plate, a vector of the number of identified objects for each well

cellNumOutput   A list containing, for each plate, a vector of intensity values for each well, e.g. a vector of the number of identified cells for each well.

Details

Positions on plates are specified with one integer only. For example, the position of the well in row 2 and column 5 is (RowNo-1)*(Number of columns on plate)+ColNo.

Value

The function generates a text file consisting of a header and a 'dataset'. The header contains the experiment description (ExternalExperimentName, TypeOfData and Comments). The dataset is an R data frame, each row corresponding to one well, with the following columns:

Spotnumber   The position of the well on the plate

Internal_GeneID
           The ID of the siRNA

GeneName	The gene name
SpotType	Can be -1, 0, 1 or 2.
	Type -1 wells (e.g. emtpy wells, wells with poor quality) are not considered in subsequent analyses but are kept in the data set for the sake of completeness.
	Type 0 wells correspond to negative controls, type 1 wells to positive controls.
	Type 2 wells correspond to the standard data wells.
SigIntensity	The signal intensity (channel 1)
SDSIntensity	The standard deviation of the signal intensity, if available
Background	The background per well, if available
LabtekNb	The plate number
RowNb	The row number
ColNb	The column number
ScreenNb	The screen number
NbCells	E.g. the number of cells identified in the well (channel 2)
PercCells	The ratio (number of identified cells)/(number of identified objects)

See Also

[joinDatasetFiles](), [joinDatasets]()

Examples

```
##gene names
plateLayout1 <- c("test1", "empty", "test3", "test4", "test5",
"test6", "test7", "empty", "test9", "test10", "test11", "test12")

plateLayout2 <- c("test1", "test2", "test3", "test4", "test5",
"test6", "test7", "test8", "test9", "test10", "test11", "test12")

plateLayout <- cbind(plateLayout1, plateLayout2)

emptyWells <- list(c(2, 8), NA_integer_)
##the first plate has two empty wells at position 2 and 8,
##the second plate does not have any empty wells

poorWells <- NA_integer_
##no wells of poor quality

controlCoordsOutput <- list(list(NA_integer_, NA_integer_), list(NA_integer_, c(9,10)))
##the first plate does not have any control siRNAs,
##the second plate has two negative controls at position 9 and 10

backgroundValOutput<-NA_integer_
##no background signal intensities available

sigPlate1<-c(2578, NA_integer_, 3784, 3784, 2578, 5555, 5555, NA_integer_, 8154, 2578, 3784, 2578)
sigPlate2<-c(8154, 3784, 5555, 3784, 11969, 2578, 1196, 5555, 17568, 2578, 5555, 2578)
##the signal intensities on the plates
```

```
meanSignalOutput<-list(sigPlate1, sigPlate2)

SDmeansignal<-NA_integer_
##no standard deviation available

objnumOutput<-NA_integer_
##no cell count available

cellnumOutput<-NA_integer_

generateDatasetFile("First test screen", "RNAi in virus-infected cells",
NA_character_, "testscreen_output.txt", plateLayout, plateLayout, 3, 4,
1, emptyWells, poorWells, controlCoordsOutput, backgroundValOutput,
meanSignalOutput, SDmeansignal, objnumOutput, cellnumOutput)

##load the dataset into R:
header<-readLines("testscreen_output.txt",3)
dataset<-read.table("testscreen_output.txt", skip=3, colClasses=c(NA, NA, NA, NA,
"factor", NA, NA, NA, NA, NA, NA, NA, NA, NA), stringsAsFactors=FALSE)
```

---

generateReplicateMat   *Generate a matrix of replicates*

Description

Generates a matrix out of a dataset, each row corresponding to an siRNA/gene ID, each column to
a channel value or its index in the dataset.

Usage

```
generateReplicateMat(data, minNbReps, IndexOrInt, col4val, col4anno)
```

Arguments

data	an R data frame generated with generateDatasetFile
minNbReps	set to 2 if you want to exclude replicates occurring only once in the dataset, otherwise 1.
IndexOrInt	a character string - either "Index" or "Intensities" - specifying which values are to be contained in the output matrix.
col4val	a character string specifying the name of the dataset column to be used for the values of the output matrix (if IndexOrIntensities is set to "Intensities"), for example "SigIntensity" or "NbCells"
col4anno	a character string specifying the name of the dataset column to be used for the output matrix' rows, for example "GeneName" or "Internal_GeneID".

Details

The function will omit values or indexes of lines/wells whose value in the column specified by
colname4val is set to NA, (which is the case if the spot type is set to -1). If you do not want to
omit those, use generateRepMatNoFilter.

Value

A matrix with each row corresponding to an siRNA/gene ID (as reflected in rownames), each column to a channel value or its index in the dataset. Missing values (in case of different number of replicates occuring for different siRNAs/genes) are set to NA.

See Also

generateRepMatNoFilter

Examples

```
data(exampleDataset, package="RNAither")

replicatematrix <- generateReplicateMat(dataset, 2, "Index", "SigIntensity",  "GeneName")
```

---

generateRepMatNoFilter

Generate a matrix of replicates (II)

Description

Generates a matrix out of a dataset, each row corresponding to an siRNA/gene ID, each column to a channel value or its index in the dataset.

Usage

```
generateRepMatNoFilter(data, minNbReps, IndexOrInt, col4val, col4anno)
```

Arguments

data	an R data frame generated with generateDatasetFile
minNbReps	set to 2 if you want to exclude replicates occurring only once in the dataset, otherwise 1.
IndexOrInt	a character string - either "Index" or "Intensities" - specifying which values are to be contained in the output matrix.
col4val	a character string specifying the name of the dataset column to be used for the values of the output matrix (if IndexOrIntensities is set to "Intensities"), for example "SigIntensity" or "NbCells"
col4anno	a character string specifying the name of the dataset column to be used for the output matrix' rows, for example "GeneName" or "Internal_GeneID".

Details

The function will not omit values or indexes of lines/wells with spot type -1. If you want to omit those, use generateReplicatematrix.

Value

A matrix with each row corresponding to an siRNA/gene ID (as reflected in rownames), each column to a channel value or its index in the dataset. Missing values (in case of different number of replicates occuring for different siRNAs/genes) are set to NA.

See Also

[generateReplicateMat](#)

Examples

```
data(exampleDataset, package="RNAither")

replicatematrix <- generateRepMatNoFilter(dataset, 2, "Index", "SigIntensity", "GeneName")
```

---

gseaAnalysis   *Perform a GSEA analysis of a list of genes*

Description

Performs a GSEA analysis of a list of genes using the package   topGO (see References).

Usage

```
gseaAnalysis(hitVector, whichOnto)
```

Arguments

hitVector	a named hit vector as generated by [hitselectionZscore](#) or [hitselectionPval](#)
whichOnto	One of the three GO ontologies: "biological_process", "molecular_function" or "cellular_component"

Value

A table containing the enriched GO terms and their significance.

References

A. Alexa, J. Rahnenfuhrer and T. Lengauer. Improved scoring of functional groups from gene expression data by decorrelating GO graph structure. Bioinformatics, 22(13):1600-1607, 2006

Adrian Alexa and Jorg Rahnenfuhrer (2006). topGO: Enrichment analysis for Gene Ontology. R package version 1.4.0.

See Also

[Ttest](#)

Examples

```
data(scoredDataset1, package="RNAither")
data(pValVec1, package="RNAither")

##for details on the generation of pValVec1 and scoredDataset1, see the example of the Ttest function linked a

scoredHits1 <- hitselectionPval(scoredDataset1, pValVec1, "SigIntensity", "Hits1", 0.1,
"GeneName", "pvalue_testfile1.txt")
hitVector1 <- scoredHits1[[2]]
gseaTable <- gseaAnalysis(hitVector1, "biological_process")
```

---

header	*a typical header of an example RNAi dataset*

Description

See [generateDatasetFile](#) for details

Usage

```
header
```

Format

See [generateDatasetFile](#)

---

headerDrosophila	*the header of the genome-wide RNAi screen of cell viability in Drosophila Kc167 cells by Boutros et al.*

Description

M. Boutros et al., Genome-wide RNAi analysis of growth and viability in Drosophila cells, Science, 303(5659):832-835, 2004. 3, 18

Usage

```
headerDrosophila
```

Format

See [generateDatasetFile](#)

---

hitselectionPval	*Selecting hits according to p-values*

Description

Selects significant genes according to their p-value.

Usage

```
hitselectionPval(dataset, pValVec, col4val, col4sel, thresh, col4anno, file4hits)
```

Arguments

dataset	an R data frame generated with [generateDatasetFile](#)
pValVec	a vector of p-values, as generated by one of the test functions [Ttest](#), [MannWhitney](#) or [RankProduct](#)
col4val	a character vector specifying a column of intensity values
col4sel	a character vector specifying the name of the new dataset column where hits will be stored
thresh	the threshold for the p-values, typically 0.05
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
file4hits	the name of the file to store the results in

Details

If there are no p-values under the defined threshold `thresh`, the threshold is increased to `min(pvalvec)`.

Value

A list containing:

dataset	the dataset with an added column defining the hits in the form of a binary vector
hitVector	the binary vector itself
replicaMatrix	a matrix of replicates with corresponding values (as generated by [generateReplicateMat](#))
thresh	the threshold for the p-values

P-values and the intensity values for each siRNA are stored in a text output file.

See Also

[hitselectionZscore](#), [hitselectionZscorePval](#), [Ttest](#)

Examples

```
data(scoredDataset1, package="RNAither")
data(pValVec1, package="RNAither")

##for details on the generation of pValVec1 and scoredDataset1, see the example of the Ttest function linked a

scoredHits1 <- hitselectionPval(scoredDataset1, pValVec1, "SigIntensity", "Pval_hits", 0.05,
"GeneName", "pvalue_testfile1.txt")

newdataset <- scoredHits1[[1]]
hitvector <- scoredHits1[[2]]
```

---

hitselectionZscore *Selecting hits according to ZScores*

Description

Selects significant genes according to their ZScore.

Usage

```
hitselectionZscore(dataset, col4zscore, col4sel, thresh, flag, flag2, col4anno,
sumFunc, file4hits)
```

Arguments

dataset   an R data frame generated with generateDatasetFile

col4zscore   a character vector specifying the name of the column containing the ZScores,
                 usually SigIntensity

col4sel   a character vector specifying the name of the new dataset column where hits will
                 be stored

thresh   the threshold for the ZScores. The interpretation depends on the choice of the
                 parameter flag2.

flag   1 or 2. 1 means the ZScores are kept per well, 2 that they are summarized
                 according to the parameter sumFunc.

flag2   1, 2 or -2.
                 If 1 is chosen and thresh == n, then the n greatest Zscores are chosen as hits.
                 If 1 is chosen and thresh == -n, then the n smallest Zscores are chosen.
                 If 1 is chosen and thresh == 0, all ZScores are chosen and written to the output
                 file.
                 If 2 is chosen, all Zscores greater than or equal to thresh are chosen.
                 If -2 is chosen, all Zscores smaller than or equal to thresh are chosen.

col4anno   a character string specifying the name of the dataset column to be used to define
                 the replicate, for example "GeneName" or "Internal_GeneID"

sumFunc   the function used to summarize ZScore values, e.g. mean or median.

file4hits   the name of the file to store the results in

Details

If flag2 == -2, and there are no ZScores under the defined threshold thresh, the threshold is
increased to min(ZScores).

If flag2 == 2, and there are no ZScores over the defined threshold thresh, the threshold is
increased to max(ZScores).

Value

A list containing:

dataset   the dataset with an added column defining the hits in the form of a binary vector
hitVector   the binary vector itself
thresh   the threshold for the ZScores

ZScores are stored in a text output file.

References

N. Malo et al. Statistical practice in high-throughput screening data analysis. Nature Biotech, 24(2): 167-175, 2006.

See Also

[hitselectionPval](), [hitselectionZscorePval](), [Ttest]()

Examples

```
data(scoredDataset1, package="RNAither")
data(pValVec1, package="RNAither")

##for details on the generation of pValVec1 and scoredDataset1, see the example of the Ttest function linked a

scoredHits1 <- hitselectionZscore(scoredDataset1, "SigIntensity", "Zscore_hits", -10,
2, 1, "GeneName", median, "Zscores_testfile1.txt")

newdataset <- scoredHits1[[1]]
hitvector <- scoredHits1[[2]]
```

---

hitselectionZscorePval

Selecting hits according to ZScores and p-values

Description

Selects significant genes according to their ZScore (summarized with the gene median) and p-values.

Usage

```
hitselectionZscorePval(dataset, pValVec, col4zscore, col4sel, thresh, thresh2,
flag2, col4anno, sumFunc, file4hits)
```

Arguments

dataset	an R data frame generated with [generateDatasetFile]()
pValVec	a vector of p-values, as generated by one of the test functions [Ttest](), [MannWhitney]() or [RankProduct]()
col4zscore	a character vector specifying the name of the column containing the ZScores, usually "SigIntensity"
col4sel	a character vector specifying the name of the new dataset column where hits will be stored
thresh	the threshold for the ZScores. The interpretation depends on the choice of the parameter flag2.
thresh2	the threshold for the p-values
flag2	2 or -2. If 2 is chosen, all Zscores greater than or equal to thresh are chosen. If -2 is chosen, all Zscores smaller than or equal to thresh are chosen.

col4anno   a character string specifying the name of the dataset column to be used to define
             the replicate, for example "GeneName" or "Internal_GeneID".

sumFunc   the function used to summarize ZScore values, e.g. mean or median.

file4hits   the name of the file to store the results in

Details

If there are no p-values under the defined threshold thresh2, it is increased to min(pvalvec).

If flag2 == -2 and there are no ZScores under the defined threshold thresh, it is increased to
min(ZScores).

If flag2 == 2 and there are no ZScores over the defined threshold thresh, it is increased to
max(ZScores).

If there are not hits for the combined threshold of p-values and ZScores, the ZScore threshold is
changed until there is a hit.

Value

A list containing:

dataset   the dataset with an added column defining the hits in the form of a binary vector

hitVector   the binary vector itself

thresh   the threshold for the ZScores

thresh2   he threshold for the p-values

ZScores and p-values are stored in a text output file.

See Also

[hitselectionPval](), [hitselectionZscore](), [Ttest]()

Examples

```
data(scoredDataset1, package="RNAither")
data(pValVec1, package="RNAither")

##for details on the generation of pValVec1 and scoredDataset1, see the example of the Ttest function linked a

scoredHits1 <- hitselectionZscorePval(scoredDataset1, pValVec1, "SigIntensity",
"Zscore_pval_hits", -1.5, 0.05, -2, "GeneName", median, "Zscores_pvals_testfile1.txt")

newdataset <- scoredHits1[[1]]
hitvector <- scoredHits1[[2]]
```

incorporatepValVec   *Incorporate a vector of p-values into a dataset*

Description

Incorporates a vector of p-values into a dataset. Also works with a dataset containing values per well (non summarized), or with a hit vector.

Usage

```
incorporatepValVec(dataset, pValVec, replicaMatrix, col4anno, colname4pval)
```

Arguments

dataset	an R data frame generated with generateDatasetFile
pValVec	a vector of p-values
replicaMatrix	a matrix of replicate values, as generated by generateReplicateMat
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, for example "GeneName" or "Internal_GeneID"
colname4pval	a character string specifying the name of the dataset column the p-values will be stored in

Value

Returns the dataset with an added column of p-values.

See Also

multTestAdjust, Ttest

Examples

```
data(exampleDataset, package="RNAither")

data(scoredDataset1, package="RNAither")
##scoredDataset1 already contains the p-value column
data(pValVec1, package="RNAither")

##for details on the generation of pValVec1 and scoredDataset1, see the example of the Ttest function linked a

temp <- generateReplicateMat(dataset, 1, "Intensities", "SigIntensity",  "GeneName")
replicamatrix <- temp[[1]]
newdataset <- incorporatepValVec(dataset, pValVec1, replicamatrix, "GeneName", "pvals")
##newdataset and scoredDataset1 are now equivalent
```

indexSubset   *Saving the indexes of a subset in the main dataset*

Description

Used together with [createSubset](#), returns the indexes in the main dataset of the wells chosen as a subset by the previous call of [createSubset](#).

Usage

```
indexSubset(listIDs, equalTo)
```

Arguments

listIDs   a character string and one of the following: Spotnumber, Internal_GeneID, GeneName, SpotTyp

equalTo   A value or character string specifying the value in the chosen column, e.g. all
               wells on plate 2

Value

An integer vector containing the indexes in the main dataset of the wells chosen as a subset by the
previous call of [createSubset](#).

See Also

[createSubset](#)

Examples

```
data(exampleDataset, package="RNAither")

subset <- createSubset(dataset, dataset$LabtekNb, 2)
indexOfSubsetInDataset <- indexSubset(dataset$LabtekNb, 2)
```

joinDatasetFiles   *Join dataset files*

Description

Merges two or more dataset files into one, with one common header.

Usage

```
joinDatasetFiles(listOfFiles, nbOfLinesInHeader, newHead, outputFile)
```

Arguments

listOfFiles   a list of the names of the files to join

nbOfLinesInHeader

                 typically 3

newHead   the new header

outputFile   the name of the file to save the header and concatenated dataset in

See Also

generateDatasetFile, joinDatasets

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")
saveDataset(header, dataset, "save_testfile1.txt")

header[[1]] <- "external_experiment_name,Test screen"
header[[2]] <- "comments,contains twice Screen Nb 1"

joinDatasetFiles(list( "save_testfile1.txt", "save_testfile1.txt"), 3, header,
"concatenated_testfile.txt")
```

---

joinDatasets   *Join datasets*

Description

Merges two or more datasets into one.

Usage

```
joinDatasets(listOfDatasets)
```

Arguments

listOfDatasets   a list of the datasets to join

Value

The joined datasets.

See Also

generateDatasetFile, joinDatasetFiles

Examples

```
data(exampleDataset, package="RNAither")
doubledataset <- joinDatasets(list(dataset, dataset))
```

LiWongRank *Li Wong rank / invariant probeset normalization*

Description

Performs a Li Wong rank / invariant probeset normalization (see References).

Usage

```
LiWongRank(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
listOfArgs	a list containing: - a character string specifying the column whose values will be used for normalization - a character string specifying the name of the dataset column to be used for the computation of the siRNA/gene ranks

Details

For each plate type/layout in each experiment, generates a ranked list of siRNAs according to their intensity values. Only siRNAs occuring only once on the plate are allowed in the list. The normalization is performed only if all plate types have a maximum of 20

For each "unique" siRNA on a plate type, the variance of its ranks across plates is computed. A histogram of variances is plotted and allows the user to choose a threshold. A list of siRNAs with rank variances under the given threshold is then returned for each plate type so that the user can choose an siRNA to normalize the plate with.

Value

Returns a list containing:

header	the new header (with an added entry about the normalization procedure in the comments)
dataset	the new dataset with normalized values. The old values are saved in an extra column with the suffix ".old"

References

C. Li and WH Wong. Model-based analysis of oligonucleotide arrays: model validation, design issues and standard error application. Genome Biol, 2(8):research0032.1-0032.11, 2001.

E. Schadt, C. Li, B. Ellis, and WH Wong. Feature Extraction and Normalization Algorithms for High-Density Oligonucleotide Gene Expression Array Data. J Cell Biochem Suppl, 37:120-125, 2001.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- LiWongRank(header, dataset, list("SigIntensity", "GeneName"))
newheader=normres[[1]]
newdataset=normres[[2]]
```

---

lowessNorm   *Lowess normalization*

Description

Performs a plate-wise lowess normalization of the data.

Usage

```
lowessNorm(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
listOfArgs	a list containing:
	- a character string specifying the column used as channel 1 (colname4ch1)
	- a character string specifying the column used as channel 2 (colname4ch2)
	- optionally: the smoother span (smSpan) of the lowess function. This gives the proportion of points which influence the smooth at each value. Larger values give more smoothness. Defaults to 2/3.

Value

Corrects intensity values in case the values of ch2 decrease with the increase of ch1 values.

Returns a list containing:

header	the new header (with an added entry about the normalization procedure in the comments)
dataset	the new dataset with normalized values. The old values are saved in an extra column with the suffix ".old"

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- lowessNorm(header, dataset, list("NbCells","SigIntensity"))
newheader <- normres[[1]]
newdataset <- normres[[2]]
```

---

mainAnalysis   *Wrapper function for full automated analysis*

Description

Performs a standard analysis of the data (quality and statistics) from a dataset file.

Usage

```
mainAnalysis(header, dataset, flagForSameExp, listOfNormalizations, listOfArgs4norm,
listOfStatTests, listOfArgs4stat, multTestAdj, hitScoringVec1, hitScoringVec2,
posNegFlag, flag4Gsea, vecOfChannels, whichOnto)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
flagForSameExp	either 0 or 1. If 1, all experiments defined in the column ScreenNb in the dataset file must have the same design (same type and same number of replicates - exact plate layout is irrelevant) so that Spearman's correlation coefficient can be computed between experiments (each with summarized replicates)
listOfNormalizations	
	a list of the normalization function to apply. Can be LiWongRank, varAdjust, divNorm, quantileNormalization, BScore, ZScore, ZScorePerScreen, subtractBackground, lowessNorm, controlNorm
listOfArgs4norm	
	a list containing, for each element of listofnormalizations, the arguments to be passed on
listOfStatTests	
	a list of the statistical tests to perform. Can be Ttest, MannWhitney, RankProduct
listOfArgs4stat	
	a list containing, for each element of listofstattests, the arguments to be passed on
multTestAdj	indicates the p-value correction for multiple testing - one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", or "none" (Type ?p.adjust for details))
hitScoringVec1	a vector of length 3 indicating (in that order):
	- scoring according to p-value (0: no, 1: yes)
	- scoring according to ZScore with ZScore < threshold (0: no, 1: yes), or according to ZScore < threshold and p-value < hitScoringVec2[1] (2)
	- scoring according to ZScore with ZScore > threshold (0: no, 1: yes), or according to ZScore > threshold and p-value < hitScoringVec2[1] (2).
	If hitScoringVec1[2] or hitScoringVec1[3] are equal to 2, hitScoringVec1[1] must be equal to one, otherwise p-values will not be computed.
hitScoringVec2	a vector of length 3 indicating the thresholds for hitscoringvec1
posNegFlag	either 0 (no controls available) or 1 (controls available)

flag4Gsea   Can be:

                 - either 0: No GSEA analysis is performed

                 - or 1: A GSEA analysis is performed for each hit scoring method

                 - or a binary vector that allows to choose which hit scoring method(s) will be
                 used for a GSEA analysis. Hit scoring methods are sorted as follows: first, hits
                 are scored according to the p-values of each test specified in `listOfStatTests` .
                 Then, if the option of scoring hits according to p-values and Intensities is cho-
                 sen (see `hitScoringVec1` , for each test, a hit vector is generated. Finally, if
                 the option of scoring hits according to Intensities only is chosen, hit vectors are
                 generated for this option.

vecOfChannels   a character vector containing the names of the channels to be used for quality
                 plots, for example `"SigIntensity"` or `"NbCells"`

whichOnto   one of the three GO hierarchies: `"biological_process"` , `"molecular_function"` or
                 `"cellular_component"` - used for the GSEA analysis

Value

Generates the html output files `index.html` and `indexnorm.html` containing the quality analysis
of raw and normalized data, respectively, and   `stats.html`, containing the statistical analysis. If
several normalization methods are applied, an `indexnorm` file is generated after each.

Note

This function is deprecated and kept only for backwards compatibility. Please use the `"rnaiter"` func-
tion instead.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

mainAnalysis(header, dataset, 0, list(controlNorm), list(list(1, 0, "SigIntensity", 1)),
list(Ttest, MannWhitney), list(list("l", 1, "SigIntensity", "GeneName"),
list("l", 1, "SigIntensity", "GeneName")), "none", c(1, 0, 0), c(0.05, 0, 0), 1,
0, c("SigIntensity", "NbCells"), "biological_process")
```

---

makeBoxplot4PlateType   *Generate a boxplot of the data per plate*

Description

Generates a boxplot comparing the same plates in different experiments.

Usage

```
makeBoxplot4PlateType(header, dataset, channel, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the column whose values will be used for the box-plot
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

For each plate type, a boxplot of intensity values per experiment will be saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle and the number of the plate.

The function returns a list containing:

plotName	the plotname
minOfPlates	the number of the first experiment
numOfPlates	the number of the last experiment

See Also

makeBoxplotControls, makeBoxplotControlsPerScreen, makeBoxplotControlsPerPlate, makeBoxplotPerPlate, makeBoxplotPerScreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

makeBoxplot4PlateType(header, dataset, "SigIntensity", "Data vs. Controls", 1)
```

---

makeBoxplotControls   *Generate a boxplot of the data vs. the controls*

Description

Generates a boxplot of intensity values of negative controls, positive controls and experimental data.

Usage

```
makeBoxplotControls(header, dataset, channel, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the column whose values will be used for the box-plot
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

A boxplot of intensity values of negative controls, positive controls and experimental data will be saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns the plot name.

See Also

[makeBoxplotControlsPerScreen](), [makeBoxplotControlsPerPlate](), [makeBoxplotPerPlate](), [makeBoxplotPerScree]()

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

makeBoxplotControls(header, dataset, "SigIntensity", "Data vs. Controls", 1)
```

---

makeBoxplotControlsPerPlate

Generate a boxplot of the data vs. the controls for each plate

---

Description

Generates a boxplot of intensity values of negative controls, positive controls and experimental data for each plate of each experiment available in the dataset.

Usage

```
makeBoxplotControlsPerPlate(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile]()
dataset	an R data frame generated with [generateDatasetFile]()
channel	a character string specifying the column whose values will be used for the box-plot
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

For each experiment, a series of boxplots of intensity values of negative controls, positive controls and experimental data will be saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle and the number of the experiment.

The function returns a list containing:

plotName   the plotname

```
c(minOfScreens, numOfScreens)
                a vector with the number of the first experiment and of the last experiment
c(minOfPlates, numOfPlates)
                a vector with the number of the first plate and the number of the last plate
```

See Also

makeBoxplotControls, makeBoxplotControlsPerScreen, makeBoxplotPerPlate, makeBoxplotPerScreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

makeBoxplotControlsPerPlate(header, dataset, "SigIntensity", "Data vs. Controls", 1, 1)
```

---

makeBoxplotControlsPerScreen

Generate a boxplot of the data vs. the controls for each experiment

---

Description

Generates a boxplot of intensity values of negative controls, positive controls and experimental data for each experiment available in the dataset.

Usage

```
makeBoxplotControlsPerScreen(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the column whose values will be used for the box-plot
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

A series of boxplots of intensity values of negative controls, positive controls and experimental data will be saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns a list containing:

plotName	the plotname
minOfScreens	the number of the first experiment
numOfScreens	he number of the last experiment

See Also

makeBoxplotControls, makeBoxplotControlsPerPlate, makeBoxplotPerPlate, makeBoxplotPerScreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

makeBoxplotControlsPerScreen(header, dataset, "SigIntensity", "Data vs. Controls", 1, 1)
```

---

makeBoxplotPerPlate   *Generate a boxplot of the data per plate*

Description

Generates a boxplot of intensity values per plate for each experiment available in the dataset.

Usage

```
makeBoxplotPerPlate(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the column whose values will be used for the box-plot
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

For each experiment, a boxplot of intensity values per plate will be saved as a pdf and a png file named after the experiment name specified in the header concatenated with the   plotTitle.

The function returns a list containing:

plotName	the plotname
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment

See Also

makeBoxplotControls, makeBoxplotControlsPerPlate, makeBoxplotControlsPerScreen, makeBoxplotPerScree

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

makeBoxplotPerPlate(header, dataset, "SigIntensity", "Data vs. Controls", 1, 1)
```

---

makeBoxplotPerScreen   *Generate a boxplot of the data per experiment*

Description

Generates a boxplot of intensity values per experiment.

Usage

```
makeBoxplotPerScreen(header, dataset, channel, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
channel	a character string specifying the column whose values will be used for the box-plot
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

A boxplot of intensity values per experiment will be saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

See Also

[makeBoxplotControls](#), [makeBoxplotControlsPerPlate](#), [makeBoxplotControlsPerScreen](#), [makeBoxplotPerPlate](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

makeBoxplotPerScreen(header, dataset, "SigIntensity", "Data vs. Controls", 1)
```

MannWhitney   *Perform a Mann-Whitney test*

Description

Performs the non-parametric Mann-Whitney test on the intensity data.

Usage

```
MannWhitney(dataset, listofargs)
```

Arguments

dataset   an R data frame generated with [generateDatasetFile](#)

listofargs   a list containing:

- "g" (greater) for significant increase, "l" (lower) for significant decrease, or "two.sided" for both

- either a number indicating the true value of the mean, or a character string indicating the name of the gene to compare with

- a character string specifying the column whose values will be used for the test

- a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"

Value

Returns a list containing:

pValVec   a named vector of p-values

dataset   the dataset with an added column "p.value.mannwhitney"

paste("pValue.mannwhitney", testType, sep="_")

                 the character string "p.value.mannwhitney" concatenated with the testType (first element of listofargs)

"Mann-Whitney test"

                 the character string "Mann-Whitney test"

See Also

[Ttest](#), [RankProduct](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

pvals1 <- MannWhitney(dataset, list("l", median(dataset$SigIntensity, na.rm=TRUE),
"SigIntensity", "GeneName"))
pValVec1 <- pvals1[[1]]
scoredDataset1 <- pvals1[[2]]
```

---

multTestAdjust   *Adjust p-values for multiple testing*

Description

Adjusts p-values for multiple testing.

Usage

```
multTestAdjust(pValVec, adjustMethod)
```

Arguments

pValVec	a vector of p-values
adjustMethod	one of the following: "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", " For details type ?p.adjust

Value

Returns a vector of corrected p-values. Can be integrated into a dataframe with the function [incorporatepValVec](#).

See Also

[incorporatepValVec](#), [Ttest](#)

Examples

```
data(pValVec1, package="RNAither")

##for details on the generation of pValVec1, see the example of the Ttest function linked above.

newpvalvec <- multTestAdjust(pValVec1,"fdr")
```

---

numCellQualControl   *Quality control of the number of cells*

Description

Plots a histogram of the cell number per well and allows the user to set an upper and a lower threshold so as to exclude wells from the analysis.

Usage

```
numCellQualControl(DataSetFile, nbLinesHeader, plotTitle)
```

Arguments

DataSetFile	a dataset file generated with [generateDatasetFile](#)
nbLinesHeader	typically 3
plotTitle	the plot title

Value

Prints out the list of wells under and over the predefined thresholds in the shell.

Saves a list of discarded siRNA values (if applicable) in a text file named after the experiment name specified in the header concatenated with either "numCellQualControl_discarded_higher.txt" or "numCellQualControl_discarded_lower.txt".

Saves the histogram with the applied thresholds in a pdf file named after the experiment name specified in the header concatenated with the `plotTitle`.

Overwrites the given `DataSetFile` with the new dataset.

See Also

`percCellQualControl`

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")
saveDataset(header, dataset, "save_testfile1.txt")

numCellQualControl("save_testfile1.txt", 3, "Histogram of the number of cells")
```

---

orderGeneIDs   *Order a dataset*

Description

Orders dataset according to one of its columns.

Usage

```
orderGeneIDs(dataset, ID1)
```

Arguments

dataset   an R data frame generated with `generateDatasetFile`

ID1   a character string specifying the name of the column according to which the dataset will be sorted

Value

An R data frame ('dataset') ordered according to its values in the specified column.

See Also

`order`

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

orderedDataset=orderGeneIDs(dataset,"SigIntensity")
```

---

percCellQualControl *Quality control of the percentage of cells*

Description

Plots a histogram of the percentage of cells per well (ratio of the number of identified cells and the number of identified objects) and allows the user to set an upper and a lower threshold so as to exclude wells from the analysis.

Usage

```
percCellQualControl(DataSetFile, nbLinesHeader, plotTitle)
```

Arguments

DataSetFile a dataset file generated with generateDatasetFile

nbLinesHeader typically 3

plotTitle the plot title

Value

Prints out the list of wells under and over the predefined thresholds in the shell.

Saves a list of discarded siRNA values (if applicable) in a text file named after the experiment name specified in the header concatenated with either "percCellQualControl_discarded_higher.txt" or "percCellQualControl_discarded_lower.txt".

Saves the histogram with the applied thresholds in a pdf file named after the experiment name specified in the header concatenated with the plotTitle.

Overwrites the given DataSetFile with the new dataset.

See Also

numCellQualControl

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")
saveDataset(header, dataset, "save_testfile1.txt")

percCellQualControl("save_testfile1.txt", 3, "Histogram of the number of cells")
```

plotBar   *Plot signal intensities per well*

Description

Plots signal intensity values for each well, a blue line showing the median, two green lines showing one median absolute deviation, two red lines showing two median absolute deviations.

Usage

```
plotBar(header, dataset, col4val, flag, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
col4val	a character string specifying the column whose intensity values will be used for the plot
flag	0, 1, or 2. 0 uses the data from the complete dataset, 1 generates one plot for each experiment, 2 generates one plot for each plate.
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the plots in pdf and png files named after the experiment name specified in the header concatenated with the plotTitle and if applicable the experiment number and/or the plate number.

When flag == 0, returns the plot name (plotName).

When flag == 1, returns a list containing:

plotName	The plot name
minOfScreens	The number of the first experiment
numOfScreens	The number of the last experiment

When flag == 2, returns a list containing: the plot name, a vector with the number of the first experiment and of the last experiment, and a vector with the number of the first plate and the number of the last plate.

See Also

ZScorePlot, ZScorePlotTwo

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotname <- plotBar(header, dataset, "SigIntensity", 0, "Data per well", 1)
```

plotControlHisto   *Plot a histogram of the data values and controls*

Description

Plots and saves a histogram of data values and shows the controls, if available, in color.

Usage

```
plotControlHisto(header, dataset, channel, plotTitle, showPlot)
```

Arguments

header   the header of a dataset file generated with generateDatasetFile

dataset   an R data frame generated with generateDatasetFile

channel   a character string specifying the name of the column containing the values to be
                plotted, e.g. "SigIntensity"

plotTitle   the plot title

showPlot   0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save
                the plot(s) without opening windows.

Value

Saves the histogram in a pdf and a png file named after the experiment name specified in the header
concatenated with the plotTitle.

Positive controls are plotted in green, negative controls in red.

The funtion returns the plot name.

See Also

plotControlHistoPerplate, plotControlHistoPerscreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotControlHisto(header, dataset, "SigIntensity", "Distribution of Data and Controls", 1)
```

plotControlHistoPerplate

Plot a histogram of the data values and controls per plate

Description

Plots and saves a histogram of data values per experiment and per plate and shows the controls, if available, in color.

Usage

```
plotControlHistoPerplate(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the name of the column containing the values to be plotted, e.g. "SigIntensity"
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the histograms in a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

Positive controls are plotted in green, negative controls in red.

The function returns a list containing:

histoName	the plotname
c(minOfScreens, numOfScreens)	
	a vector with the number of the first experiment and of the last experiment
c(minOfPlates, numOfPlates)	
	a vector with the number of the first plate and the number of the last plate

See Also

plotControlHisto, plotControlHistoPerscreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotControlHistoPerplate(header, dataset, "SigIntensity", "Distribution of Data and Controls", 1, 1)
```

plotControlHistoPerscreen

Plot a histogram of the data values and controls per experiment

Description

Plots and saves a histogram of data values per experiment and shows the controls, if available, in color.

Usage

```
plotControlHistoPerscreen(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the name of the column containing the values to be plotted, e.g. "SigIntensity"
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the histograms in a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

Positive controls are plotted in green, negative controls in red.

The function returns a list containing:

histoName	the plotname
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment

See Also

plotControlHisto,plotControlHistoPerplate

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotControlHistoPerscreen(header, dataset, "SigIntensity", "Distribution of Data and Controls", 1, 1)
```

plotHisto	*Plot a histogram of the data values*

Description

Plots and saves a histogram of the chosen data values.

Usage

```
plotHisto(header, dataset, channel, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
channel	a character string specifying the name of the column containing the values to be plotted, e.g. `"SigIntensity"`
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the histogram in a pdf and a png file named after the experiment name specified in the header concatenated with the `plotTitle`.

The function returns the plot name.

See Also

plotHistoPerplate, plotHistoPerscreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotHisto(header, dataset, "SigIntensity", "Distribution of Data and Controls", 1)
```

plotHistoPerplate	*Plot a histogram of the data values per plate*

Description

Plots and saves a histogram of the chosen data values per experiment and per plate.

Usage

```
plotHistoPerplate(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
channel	a character string specifying the name of the column containing the values to be plotted, e.g. `"SigIntensity"`
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the histograms in a pdf and a png file named after the experiment name specified in the header concatenated with the `plotTitle`.

The function returns a list containing:

histoName	the plotname
c(minOfScreens, numOfScreens)	
	a vector with the number of the first experiment and of the last experiment
c(minOfPlates, numOfPlates)	
	a vector with the number of the first plate and the number of the last plate

See Also

[plotHisto](#), [plotHistoPerscreen](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotHistoPerplate(header, dataset, "SigIntensity", "Distribution of Data and Controls", 1, 1)
```

---

plotHistoPerscreen   *Plot a histogram of the data values per experiment*

Description

Plots and saves a histogram of the chosen data values.

Usage

```
plotHistoPerscreen(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
channel	a character string specifying the name of the column containing the values to be plotted, e.g. "SigIntensity"
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the histograms in a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns a list containing:

histoName	the plotname
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment

See Also

[plotHisto](#), [plotHistoPerplate](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotHistoPerscreen(header, dataset, "SigIntensity", "Distribution of Data and Controls", 1, 1)
```

---

plotQQ	*Make a QQ plot*

Description

Shows and saves a QQ plot of the data.

Usage

```
plotQQ(header, dataset, channel, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
channel	a character string specifying the name of the column containing the values to be plotted, e.g. "SigIntensity"
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the QQ plot in a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns the plot name.

See Also

[plotQQperscreen](#), [plotQQperplate](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotQQ(header, dataset, "SigIntensity", "QQplot", 1)
```

---

plotQQperplate   *Make a QQ plot per plate*

Description

Shows and saves a QQ plot of the data for each experiment and each plate in the dataset.

Usage

```
plotQQperplate(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
channel	a character string specifying the name of the column containing the values to be plotted, e.g. "SigIntensity"
plotTitle	the plot title
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the QQ plots in a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns a list containing:

histoName   the plotname

c(minOfScreens, numOfScreens)
                     a vector with the number of the first experiment and of the last experiment

c(minOfPlates, numOfPlates)
                     a vector with the number of the first plate and the number of the last plate

See Also

[plotQQ](#), [plotQQperscreen](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotQQperplate(header, dataset, "SigIntensity", "QQplot", 1, 1)
```

---

plotQQperscreen   *Make a QQ plot per experiment*

Description

Shows and saves a QQ plot of the data for each experiment in the dataset.

Usage

```
plotQQperscreen(header, dataset, channel, plotTitle, plotDesign, showPlot)
```

Arguments

header   the header of a dataset file generated with [generateDatasetFile](#)

dataset   an R data frame generated with [generateDatasetFile](#)

channel   a character string specifying the name of the column containing the values to be plotted, e.g. "SigIntensity"

plotTitle   the plot title

plotDesign   1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.

showPlot   0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the QQ plots in a pdf and a png file named after the experiment name specified in the header concatenated with the `plotTitle`.

The function returns a list containing:

histoName	the plotname
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment

See Also

[plotQQ](#), [plotQQperplate](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotQQperscreen(header, dataset, "SigIntensity", "QQplot", 1, 1)
```

---

pValVec1   *A vector of p-values after a median normalization and a t-test*

Description

See [divNorm](#) and [Ttest](#) for details

Usage

```
pValVec1
```

Format

vector

---

pValVec2   *A vector of p-values after a Mann-Whitney test*

Description

See [MannWhitney](#) for details

Usage

```
pValVec2
```

Format

vector

quantileNormalization   *Quantile normalization*

Description

Quantile normalization (see References)

Usage

```
quantileNormalization(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
listOfArgs	a list containing: - a character string specifying the column whose values will be used for normalization - 1 or 2, 1 meaning a normalization per experiment, 2 meaning a normalization per plate

Value

Returns a list, containing:

header	the new header (with an added entry about the normalization procedure in the comments)
dataset	the new dataset with normalized values. The old values are saved in an extra column with the suffix ".old"

References

B.M. Bolstad, R.A. Irizarry, M. Astrand, and T.P. Speed. A Comparison of Normalization Methods for High Density Oligonucleotide Array Data Based on Variance and Bias. Bioinformatics, 19(2): 185-193, 2003

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- quantileNormalization(header, dataset, list(2, "SigIntensity"))
newheader <- normres[[1]]
newdataset <- normres[[2]]
```

RankProduct   *Perform a Rank Product test*

Description

Performs the non-parametric rank product test on the intensity data.

Usage

```
RankProduct(dataset, listofargs)
```

Arguments

dataset   an R data frame generated with [generateDatasetFile](#)

listofargs   a list containing:
                 - the number of permutations to perform to compute the p-values (usually 100)
                 - 1 or 2, depending if the search is for a significant decrease or increase
                 - a character string specifying the column whose values will be used for the test
                 - a character string specifying the name of the dataset column to be used to define the replicate, for example ″GeneName″ or ″Internal_GeneID″

Value

Returns a list containing

pValVec   a named vector of p-values

dataset   the dataset with an added column ″p.value.rankproduct″

paste("pValue.rankproduct", testType, sep="_")
                     the character string ″p.value.rankproduct″

"Rank product test"
                     the character string "Rank product test"

The p values returned are equivalent to the percentage of false prediction (pfp), which in theory is the equivalent of false discovery rate (FDR). It is possible that they are larger than 1.

See Also

[Ttest](#), [MannWhitney](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

pvals1 <- RankProduct(dataset, list(100, 1, "SigIntensity", "GeneName"))
pValVec1 <- pvals1[[1]]
scoredDataset1 <- pvals1[[2]]
```

replicatesCV	*Compute the correlation of variation (CV)*

Description

Computes the correlation of variation as defined in Tseng et al. (see References)

Usage

```
replicatesCV(header, dataset, PlotTitle, col4val, col4anno, plotDesign, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
PlotTitle	the plot title
col4val	a character string specifying the column whose values will be used to compute the correlation of variation
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
plotDesign	1 or 2. 1 will generate one window containing all plots, 2 will generate a series of plots.
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

The correlation of variation of an siRNA is defined as the standard deviation of its values divided by their mean.

The function generates a plot of the average intensity against the CV for each experiment. The plot will be saved as a pdf and a png file named after the experiment name specified in the header concatenated with the PlotTitle.

The function returns a list containing:

histoName	the plotname
minOfScreens	the number of the first experiment
numOfScreens	the number of the last experiment

References

G. C. Tseng et al. Issues in cDNA microarray analysis: quality filtering, channel normalization, models of variations and assessment of gene effects. Nucleic Acids Res, 29(12): 2549-2557, 2001.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

replicatesCV(header, dataset, "Correlation of Variation versus Mean Intensity",
"SigIntensity", "GeneName", 1, 0)
```

replicatesSpearmancor   *Compute the correlation coefficient betwenn replicates or experiments*

Description

Computes Spearman's rank correlation coefficient for each replicate - either inside each experiment, or between experiments.

Usage

```
replicatesSpearmancor(header, dataset, flag, col4val, col4anno, fileNameSuffix)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
flag	1 or 2. 1 will compute the coefficient for a maximum of 3 replicates, for each experiment available in the dataset. 2 will summarize the replicates from each experiment with their root mean square and compute the correlation coefficient between experiments.
col4val	a character string specifying the column whose values will be used to compute the correlation coefficient
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
fileNameSuffix	a character string that will be used to name the output file containing a table with the correlation coefficients.

Value

For flag==1, the correlation coefficients are printed out to the shell and saved in a text file named after the experiment name specified in the header concatenated with the character string filename-suffix and "Spearmancor.txt".

For flag==2, the correlation coefficients are printed out to the shell and saved in a text file named after the experiment name specified in the header concatenated with the character string filename-suffix and "Spearmancor_AllExp.txt".

The function returns a table containing the correlation coefficients.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

replicatesSpearmancor(header, dataset, 1, "SigIntensity", "GeneName", "testfile1_")
```

rms   *Compute the replicate root mean square*

Description

Computes the root mean square of replicate values

Usage

```
rms(Ivec, na.rm = T)
```

Arguments

Ivec	All channel values for a specific siRNA/gene
na.rm	Removes NA values

Value

A double giving the root mean square of the given replicate values.

See Also

[trim](#), [closestToZero](#), [furthestFromZero](#), [summarizeReps](#), [summarizeRepsNoFiltering](#)

Examples

```
data(exampleDataset, package="RNAither")

Indexes <- findReplicates(dataset, "GeneName", "CPSF1")
rmsval <- rms(dataset$SigIntensity[Indexes])
```

rnaither   *Wrapper function for full automated analysis*

Description

Performs a standard analysis of the data (quality and statistics) from a dataset file.

Usage

```
rnaither(data, expname, excludeCellcounts="none", logtransform=FALSE, normalization=c("lowess","b
```

Arguments

data
: A data frame containing the experimental data to analyze. Each row is corresponding to one well, with the following columns:

  - Spotnumber The position of the well on the plate
  - Internal_GeneID The ID of the siRNA
  - GeneName The gene name
  - SpotType Can be -1, 0, 1 or 2. Type -1 wells (e.g. emtpy wells, wells with poor quality) are not considered in subsequent analyses but are kept in the data set for the sake of completeness. Type 0 wells correspond to negative controls, type 1 wells to positive controls. Type 2 wells correspond to the standard data wells.
  - SigIntensity The signal intensity (channel 1)
  - SDSIntensity The standard deviation of the signal intensity, if available
  - Background The background per well, if available
  - LabtekNb The plate number
  - RowNb The row number
  - ColNb The column number
  - ScreenNb The screen number
  - NbCells E.g. the number of cells identified in the well (channel 2)
  - PercCells The ratio (number of identified cells)/(number of identified objects)

expname
: A character string, assigning a name to the experiment. This will be used as title in the html output generated by rnaither.

excludeCellcounts
: a string constant, one of ″none″, ″lowest″, ″both″, ″lowestperplate″ or ″bothperplate″. The default is ″none″. This parameter can be used to exclude wells from the analysis that have very low or very high numbers of cells.

  - "none" No wells will be excluded based on the number of cells they contain.
  - "lowest", "lowestperplate" The wells with the lowest 5 percent of cellcounts will be excluded from further analysis. "lowest" will consider the entire screen at once, and exclude the wells that are overall the lowest 5 percent. "lowestperplate" will consider each plate separately, excluding on each plate the 5 percent of wells having the lowest cellcounts.
  - "both", "bothperplate" The wells with the lowest and highest 5 percent of cellcounts will be excluded from further analysis. Excluding wells with high cell counts may be useful for image based screens, if it is suspected that cells overlap in images, which might cause problems for image processing. "both" will consider the entire screen at once, and exclude the wells that are overall the lowest and highest 5 percent. "bothperplate" will consider each plate separately, excluding on each plate the 5 percent of wells having the lowest and highest cellcounts.

logtransform
: A logical variable, specifying whether or not the signal intensities should be log-transformed. Default is FALSE.

normalization
: A list of strings containing the normalization steps to carry out. The default are is c(″lowess″,″bscore″). The following normalization procedures are available:

  - "lowess" To carry out lowess normalization. This corrects for effects of cell counts on the signal intensities.

- "liwong"To carry out Li-Wong rank normalization of the signal intensities.
- "varadjust"To divide each signal intensity value by the variance of the signal intensities on the respective plate.
- "divnorm"To divide each signal intensity value by the median signal intensity of the respective plate.
- "quantile"To carry out a quantile normalization on the signal intensities.
- "bscore"To carry out a bscore normalization on the signal intensities (corrects for spatial effects on a plate.
- "zscore"To carry out a zscore normalization (subtract median of plate, divide by median absolute deviation per plate).
- "negcontrol"To normalize on the negative controls - subtract median of negative controls, divide by MAD of negative controls, per plate.
- "percontrol"To do a percentage of controls normalization - Rescale signal intensities so that mean of negative controls is 100, mean of positive controls is 0.
- "percneg"To do a percentage of negative controls normalization - set mean of negative controls to 100, zero signal intensity remains at 0. Normalization routines will be executed in the order as they occur in the list.

test   Specify what statistical test should be used to identify hits. One of

- "ttest"to carry out a t-test if the mean score for a given siRNA / Gene is 0.
- "wilcox"to carry out a Wilcoxon test if the mean score for a given siRNA / Gene is 0.
- "none"to carry out no statistical test.

The default is "ttest".

scorethresh   The threshold on the normalized score to be used to identify hits. The default is 2.0, hence siRNAs with score > +2 or score < -2 are considered hits.

pvalthresh   The threshold on the p-value from the statistical test to be used to identify hits. The default is 0.05

dogo   A logical variable, specifies whether or not a Gene Ontology-based analysis should be carried out. This parameter is currently ignored, GO is presently not supported nby the rnaither wrapper.

outdir   a string specifying the directory in which the results should be stored. Can be an absolute or relative path.

layoutnames   A list of strings, that can be used to assign names to different layouts in the screen. The list should contain the same number of elements as there are different layouts in the screen. These names will be used as labels for the layouts in the html output. If this parameter is not specified, layouts will be numbered in the canonical way.

makeplots   TRUE or FALSE, if set to FALSE, only a subset of the quality control plots will be generated. This speeds up processing, but will result in missing images in the html output.

reorder   logical variable, indicating whether dataset should be reordered prior to processing further. This is recommended if the data frame is incomplete, i.e. if wells or plates are missing completely. reorder=T will considerably slow down the analysis.

Value

Generates the html output file index.html in the directory specified by the outdir parameter.

Examples

```
data(exampleDataset, package="RNAither")

rnaither(dataset, expname="Example", excludeCellcounts="none", logtransform=FALSE, normalization=c("lowess"
```

---

saveDataset   *Save the normalized dataset into a dataset text file*

Description

Saves the normalized dataset and corresponding header into the specified dataset text file.

Usage

```
saveDataset(header, data, dataSetFile)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
data	an R data frame generated with [generateDatasetFile](#)
dataSetFile	the name of the text file the data will be saved in; can be the same as the old file (will be overwritten without prompting)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- divNorm(header, dataset, list(median, 2, 1, "SigIntensity", 1))
newheader <- normres[[1]]
newdataset <- normres[[2]]
saveDataset(newheader, newdataset, "save_testfile1.txt")
```

---

saveOldIntensityColumns

                               Save old intensity value columns

---

Description

Duplicates the specified column and adds it to the end of the dataset.

Usage

```
saveOldIntensityColumns(dataset, col4val)
```

Arguments

dataset	an R data frame generated with [generateDatasetFile](#)
col4val	a character string specifying the column whose values will be saved as an extra column before normalization

Value

The values in the chosen column are saved in an extra column with the suffix ".old".

Examples

```
data(exampleDataset, package="RNAither")
newdataset <- saveOldIntensityColumns(dataset, "SigIntensity")
```

---

savepValVec   *Save p-values to file*

Description

Saves a vector of p-values to a text file.

Usage

```
savepValVec(pValVec, filename)
```

Arguments

pValVec	a vector of p-values
filename	the name of the text file to save the p-values to.

See Also

[Ttest](#)

Examples

```
data(pValVec1, package="RNAither")

##for details on the generation of pValVec1, see the example of the Ttest function linked above.

savepValVec(pValVec1, "pvals_testfile1.txt")
```

---

scoredDataset1   *A dataset containing an additional column showing the p-values, after*
                               a median normalization and a t-test

---

Description

See [divNorm](#) and [Ttest](#) for details

Usage

```
scoredDataset1
```

Format

see [generateDatasetFile](#) for details

scoredDataset2	*A dataset containing an additional column showing the p-values after a Mann-Whitney test*

Description

See [MannWhitney](#) for details

Usage

```
scoredDataset1
```

Format

see [generateDatasetFile](#) for details

SNRQualControl	*Computing the SNR*

Description

Computes the signal to noise ratio for all data, per experiment and per plate for a complete dataset file and plots histograms of the results.

Usage

```
SNRQualControl(dataSetFile, nbLinesHeader, channel, noise, plotTitle, showPlot)
```

Arguments

dataSetFile	a dataset file generated with [generateDatasetFile](#)
nbLinesHeader	typically 3
channel	a character string specifying the name of the column containing the values for computing the SNR, e.g. `"SigIntensity"`
noise	A character string specifying the name of the column containing the values for computing the SNR, e.g. `"Background"`
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

Shows histogram plots of the SNR for the whole dataset file, per experiment and per plate and saves them in a pdf file. The name of the file will be the concatenation of the experiment name specified in the header and the function argument `plotTitle`.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")
saveDataset(header, dataset, "save_testfile1.txt")

SNRQualControl("save_testfile1.txt", 3, "SigIntensity", "Background", "SNR", 1)
```

---

spatialDistrib   *Generate spatial plots of intensity values*

Description

Generate plots of plates and their intensity values.

Usage

```
spatialDistrib(header, dataset, plotTitle, col4plot, col4anno, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
plotTitle	the plot title
col4plot	a character string specifying the column whose values will be used for the plot
col4anno	a character string specifying the column whose values will be used for the annotation of the plot
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

For each plate, the plot will be saved as a png file named after the experiment name specified in the header concatenated with the   plotTitle, the number of the experiment, and the number of the plate.

Wells containing positive controls are marked with a "P", wells containing negative controls with an "N".

Each plate will also be saved as an html file containing mouse-overs with the siRNA name for each well.

The function returns a list containing:

histoName	the plotname
c(minOfScreens, numOfScreens)	
	a vector with the number of the first experiment and of the last experiment
c(minOfPlates, numOfPlates)	
	a vector with the number of the first plate and the number of the last plate

See Also

compareReplicateSD, compareReplicateSDPerScreen

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

spatialDistrib(header, dataset, "Spatial distribution of cell counts", "NbCells", "GeneName", 1)
```

---

spatialDistribHits   *Plotting the spatial distribution of the hits*

Description

Plots the plates showing the spatial distribution of the hits using the `plotPlate` function of the `prada` package.

Usage

```
spatialDistribHits(header, dataset, plotTitle, col4hits, col4anno, showPlot)
```

Arguments

header	the header of a dataset file generated with `generateDatasetFile`
dataset	an R data frame generated with `generateDatasetFile`
plotTitle	the plot title
col4hits	a character vector specifying the name of the dataset column containing the binary hit vector
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. `"GeneName"` or `"Internal_GeneID"`
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

For each plate, the plot will be saved as a png file named after the experiment name specified in the header concatenated with the `plotTitle`, the number of the experiment, and the number of the plate.

Wells containing positive controls are marked with a "P", wells containing negative controls with an "N".

Each plate will also be saved as an html file containing mouse-overs with the siRNA name for each well.

The function returns a list containing:

histoName	the plotname
c(minOfScreens, numOfScreens)	
	a vector with the number of the first experiment and of the last experiment
c(minOfPlates, numOfPlates)	
	a vector with the number of the first plate and the number of the last plate

See Also

Ttest

Examples

```
data(exampleHeader, package="RNAither")
data(pValVec1, package="RNAither")
data(scoredDataset1, package="RNAither")

##for details on the generation of pValVec1 and scoredDataset1, see the example of the Ttest function linked a

scoredHits1 <- hitselectionPval(scoredDataset1, pValVec1, "SigIntensity", "Hits1", 0.05,
"GeneName", "pvalue_testfile1.txt")

hitDataset1 <- scoredHits1[[1]]

spatialDistribHits(header, hitDataset1, "Spatial distribution of hits", "Hits1", "GeneName", 1)
```

---

subtractBackground   *Background substraction*

Description

Substracts a specified background value from the intensity values.

Usage

```
subtractBackground(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
listOfArgs	a list containing:
	- a character string specifying the column whose values will be used for background substraction
	- a character string specifying the column whose values will be used as background

Value

A list containing:

header	The new header (with an added entry about the normalization procedure in the comments)
dataset	The new dataset with normalized values. The old values are saved in an extra column of the dataset with the suffix ".old"

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- subtractBackground(header, dataset, list("SigIntensity", "Background"))
newheader <- normres[[1]]
newdataset <- normres[[2]]
```

---

sumChannels   *Summarize channels*

Description

Summarizes two channels, for example by computing their ratio.

Usage

```
sumChannels(header, dataset, funName, colname4ch1, colname4ch2)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
funName	the function used to summarize the two channels, for example divideChannels
colname4ch1	a character string specifying the name of the dataset column containing the first channel
colname4ch2	a character string specifying the name of the dataset column containing the second channel

Details

The original dataset columns are saved as extra columns with the suffix ".old" by the function saveOldIntensityColumns.

Value

A list containing:

header	the header with an entry about the channel summarization added in the comments section
newDataset	the new dataset

See Also

eraseDataSetColumn, divideChannels, saveOldIntensityColumns

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

newdataset=sumChannels(header, dataset, divideChannels, "SigIntensity", "NbCells")
```

summarizeReps   *Generate a new dataset with summarized replicates*

Description

Generates a new dataset with summarized replicates.

Usage

```
summarizeReps(data, funSum, col4val, col4anno, cols2del)
```

Arguments

data	an R data frame generated with generateDatasetFile
funSum	a function used to summarize the values of a replicate, e.g. mean, median, rms, trim, max, min, closestToZero, furthestFromZero, ...
col4val	a character vector (containing for example "SigIntensity", Background, NbCells, PercCells, ...) specifying the columns that will be summarized by funSum
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
cols2del	a character vector containing the columns to delete, for example "SDSIntensity"

Details

All columns containing replicate values will be summarized by funSum. For all columns containing positions, screen numbers, plate numbers, etc., all information for different replicates will be kept, comma-separated. All columns containing standard deviations of channels should be specified in colnames2delete.

Value

Returns the summarized dataset.

See Also

summarizeRepsNoFiltering, eraseDataSetColumn, generateReplicateMat, generateRepMatNoFilter, mean, median, rms, trim, max, min, closestToZero, furthestFromZero

Examples

```
data(exampleDataset, package="RNAither")

colname4val <- c("SigIntensity", "Background", "NbCells", "PercCells")
summarizeddataset <- summarizeReps(dataset, mean, colname4val, "GeneName", "SDSIntensity")
```

summarizeRepsNoFiltering

Generate a new dataset with summarized replicates

Description

Generates a new dataset with summarized replicates. Keeps wells/spots with SpotType -1 in the dataset, but intensity values are replaced with NA.

Usage

```
summarizeRepsNoFiltering(data, funSum, col4val, col4anno, cols2del)
```

Arguments

data	an R data frame generated with [generateDatasetFile](#)
funSum	a function used to summarize the values of a replicate, e.g. mean, median, [rms](#), [trim](#), max, min, [closestToZero](#), [furthestFromZero](#), ...
col4val	a character vector (containing for example "SigIntensity", Background, NbCells, PercCells, ...) specifying the columns that will be summarized by funSum
col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID"
cols2del	a character vector containing the columns to delete, for example "SDSIntensity"

Details

All columns containing replicate values will be summarized by funSum. For all columns containing positions, screen numbers, plate numbers, etc., all information for different replicates will be kept, comma-separated. All columns containing standard deviations of channels should be specified in colnames2delete.

Value

Returns the summarized dataset.

See Also

[summarizeReps](#), [eraseDataSetColumn](#), [generateReplicateMat](#), [generateRepMatNoFilter](#), mean, median, [rms](#), [trim](#), max, min, [closestToZero](#), [furthestFromZero](#)

Examples

```
data(exampleDataset, package="RNAither")

colname4val <- c("SigIntensity", "Background", "NbCells", "PercCells")
summarizeddataset <- summarizeRepsNoFiltering(dataset, mean, colname4val, "GeneName", "SDSIntensity")
```

---

trim   *Compute the replicate mean with trimmed values*

Description

Computes the mean of replicate values, omitting the highest and the lowest 5

Usage

```
trim(Ivec, na.rm = T)
```

Arguments

Ivec   All channel values for a specific siRNA/gene

na.rm   Removes NA values

Value

A double giving the trimmed mean of the given replicate values, i.e. omitting the highest and the lowest 5

See Also

[rms](#), [closestToZero](#), [furthestFromZero](#), [summarizeReps](#), [summarizeRepsNoFiltering](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

Indexes <- findReplicates(dataset, "GeneName", "CPSF1")
replicatemean <- trim(dataset$SigIntensity[Indexes])
```

Ttest   *Perform a Student's t-test*

Description

Performs a Student's t-test on the intensity data.

Usage

```
Ttest(dataset, listofargs)
```

Arguments

dataset an R data frame generated with generateDatasetFile

listofargs a list containing:

- ″g″ (greater) for significant increase, ″l″ (lower) for significant decrease, or ″two.sided″ for both

- either a number indicating the true value of the mean, or a character string indicating the name of the gene to compare with

- a character string specifying the column whose values will be used for the test

- a character string specifying the name of the dataset column to be used to define the replicate, e.g. ″GeneName″ or ″Internal_GeneID″

Value

Returns a list containing:

pValVec a named vector of p-values

dataset the dataset with an added column ″p.value.mannwhitney″

paste(″pValue.ttest″, testType, sep=″_″)

the character string ″pValue.ttest″ concatenated with the testType (first element of listofargs)

″t test″ the character string ″t test″

See Also

MannWhitney, RankProduct

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

pvals1 <- Ttest(dataset, list("l", median(dataset$SigIntensity, na.rm=TRUE), "SigIntensity", "GeneName"))
pValVec1 <- pvals1[[1]]
scoredDataset1 <- pvals1[[2]]
```

---

varAdjust *Variance adjustment*

Description

Divides the intensity values by their median absolute deviation (of the experiment or of the plate)

Usage

```
varAdjust(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
listOfArgs	a list containing:
	- a character string specifying the column whose values will be used for normalization
	- 1 or 2, 1 meaning a normalization per screen, 2 a normalization per plate
	- a flag specifying whether controls should be excluded for the computation of the median absolute deviation (1) or not (0).

Value

Divides the intensity values by their median absolute deviation (of the experiment or of the plate).

Returns a list containing:

header	The new header (with an added entry about the normalization procedure in the comments)
dataset	The new dataset with normalized values. The old values are saved in an extra column of the dataset with the suffix ".old"

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- varAdjust(header, dataset, list(1, "SigIntensity", 1))
newheader <- normres[[1]]
newdataset <- normres[[2]]
```

---

vennDiag	*Plotting a Venn Diagram to compare hits*

Description

Plots a Venn Diagram of up to three binary hit vectors.

Usage

```
vennDiag(header, listOfCols, listOfNames, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
listOfCols	a list of binary hit vectors to compare
listOfNames	a list of character strings for the annotation of the Venn Diagram
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

The plot is saved in a pdf and a png file named after the experiment name specified in the header concatenated with the `plotTitle`.

The function returns the plot name.

See Also

[Ttest](), [MannWhitney]()

Examples

```
data(exampleHeader, package="RNAither")

data(pValVec1, package="RNAither")
data(pValVec2, package="RNAither")
data(scoredDataset1, package="RNAither")
data(scoredDataset2, package="RNAither")

##for details on the generation of pValVec and scoredDataset,
##see the examples of the functions Ttest and MannWhitney linked above.

scoredHits1 <- hitselectionPval(scoredDataset1, pValVec1, "SigIntensity", "pValue.ttest_l", 0.05,
"GeneName", "pvalue_testfile1.txt")

scoredHits2 <- hitselectionPval(scoredDataset2, pValVec2, "SigIntensity", "pValue.mannwhitney_l", 0.05,
"GeneName", "pvalue_testfile2.txt")

hitvector1 <- scoredHits1[[2]]
hitvector2 <- scoredHits2[[2]]

plot_name <- vennDiag(header, list(hitvector1, hitvector2), list("t test", "Mann-Whitney test"),
"Venn diagram", 1)
```

---

volcanoPlot	*Making a volcano plot*

Description

Makes a volcano plot of the data.

Usage

```
volcanoPlot(header, dataset, col4plotx, col4ploty, col4anno, plotTitle, sigLevel, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile]()
dataset	an R data frame generated with [generateDatasetFile]()
col4plotx	a character vector specifying the name of the column containing the intensity values, usually `SigIntensity`
col4ploty	a character vector specifying the name of the dataset column containing the corresponding p-values

col4anno	a character string specifying the name of the dataset column to be used to define the replicate, e.g. "GeneName" or "Internal_GeneID".
plotTitle	the plot title
sigLevel	the significance level for the p-value, indicating where a horizontal green line will be drawn
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows

Value

Plots the intensity values against the negative decadic logarithm of the p-values. A green horizontal line is drawn at the specified significance level.

The plot is saved in a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns the plot name.

See Also

[Ttest](#)

Examples

```
data(exampleHeader, package="RNAither")
data(pValVec1, package="RNAither")
data(scoredDataset1, package="RNAither")

##for details on the generation of pValVec1 and scoredDataset1, see the example of the Ttest function linked a

scoredHits1 <- hitselectionPval(scoredDataset1, pValVec1, "SigIntensity", "pValue.ttest_l", 0.05,
"GeneName", "pvalue_testfile1.txt")

hitDataset1 <- scoredHits1[[1]]
hitvector1 <- scoredHits1[[2]]

volcano_name <- volcanoPlot(header, hitDataset1, "SigIntensity", "pValue.ttest_l", "GeneName",
"Volcano Plot", 0.05, 1)
```

ZPRIMEQualControl   *Computing the Z' factor*

Description

Computes the Z' factor per plate for a complete dataset file and plots the results.

Usage

```
ZPRIMEQualControl(header, data, channel, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
data	an R data frame generated with [generateDatasetFile](#)
channel	a character string specifying the name of the column containing the values for computing the Z' factor, e.g. "SigIntensity"
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Returns the Z' values in the shell for each plate and saves them in a text file. The name of the text file will be the concatenation of the experiment name specified in the header and the character string "Z'Scores.txt".

Shows a plot of the Z' factor values and saves it as a png and a pdf file under the experiment name specified in the header concatenated with the function argument plotTitle.

The function returns a list containing:

plotName	the plot name
ZPrimeTabelle	table containing the Z' values

References

J. Zhang, T. Chung, and K. Oldenburg. A simple statistical parameter for use in evaluation and validation of high throughput screening assays. J Biomol Screen, 4:67-73, 1999.

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

res <- ZPRIMEQualControl(header, dataset, "SigIntensity", "Z' factors per plate", 1)
zprime_plot <- res[[1]]
zprime_table <- res[[2]]
```

ZScore   *ZScore normalization*

Description

ZScore normalization (see Value and References)

Usage

```
ZScore(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
listOfArgs	a list containing: - a character string specifying the column whose values will be used for normalization - a flag specifying whether controls should be excluded for the computation of the median and median absolute deviation (1) or not (0).

Value

The ZScore is defined as the quotient of the difference between an intensity value and the median of the plate, and of the median absolute deviation.

Returns a list containing:

header	The new header (with an added entry about the normalization procedure in the comments)
dataset	The new dataset with normalized values. The old values are saved in an extra column of the dataset with the suffix ".old"

References

N. Malo et al. Statistical practice in high-throughput screening data analysis. Nature Biotech, 24(2): 167-175, 2006.

See Also

[ZScorePerScreen](#), [BScore](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- ZScore(header, dataset, list("SigIntensity", 1))
newheader <- normres[[1]]
newdataset <- normres[[2]]
```

ZScorePerScreen   *ZScore normalization per experiment*

Description

ZScore normalization not per plate, but per experiment (see Value and References)

Usage

```
ZScorePerScreen(header, dataset, listOfArgs)
```

Arguments

header	the header of a dataset file generated with `generateDatasetFile`
dataset	an R data frame generated with `generateDatasetFile`
listOfArgs	a list containing:
	- a character string specifying the column whose values will be used for normalization
	- a flag specifying whether controls should be excluded for the computation of the median and median absolute deviation (1) or not (0).

Value

The ZScore is defined as the quotient of the difference between an intensity value and the median of the experiment, and of the median absolute deviation.

Returns a list containing:

header	The new header (with an added entry about the normalization procedure in the comments)
dataset	The new dataset with normalized values. The old values are saved in an extra column of the dataset with the suffix ".old"

References

N. Malo et al. Statistical practice in high-throughput screening data analysis. Nature Biotech, 24(2): 167-175, 2006.

See Also

ZScore, BScore

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normres <- ZScorePerScreen(header, dataset, list("SigIntensity", 1))
newheader <- normres[[1]]
newdataset <- normres[[2]]
```

ZScorePlot	*Plot normalized intensity values per well*

Description

Plots the normalized intensity values for each well, together with a black line showing the mean, two green lines showing the standard deviation, and two red lines showing 2 standard deviations.

Usage

```
ZScorePlot(header, dataset, flag, col4plot, col4anno, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with generateDatasetFile
dataset	an R data frame generated with generateDatasetFile
flag	either 1 or 2. 1 if the dataset contains values per well, 2 if the dataset contains summarized values for each siRNA (e.g. a dataset summarized with summarizeReps).
col4plot	a character string specifying the column whose values will be used for the plot
col4anno	a character string specifying the column that will be used for the plot annotation
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Plots the normalized intensity values for each well, together with a black line showing the mean, and two red lines showing 2 standard deviations. Clicking on the points shows the gene/siRNA name.

The plot is saved as a pdf and a png file named after the experiment name specified in the header concatenated with the plotTitle.

The function returns the plot name.

See Also

plotBar, ZScorePlotTwo

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

normedvals <- ZScore(header, dataset, list("SigIntensity", 1))
ZScorePlot(normedvals[[1]], normedvals[[2]], 1, "SigIntensity", "GeneName",
"Normed intensity values per well", 1)
```

ZScorePlotTwo   *Plot signal intensities per well (II)*

Description

Plots signal intensity values for each well, a black line showing the median, two green lines showing one median absolute deviation, two red lines showing two median absolute deviations.

Usage

```
ZScorePlotTwo(header, dataset, flag, flag2, col4plot, col4anno, plotTitle, showPlot)
```

Arguments

header	the header of a dataset file generated with [generateDatasetFile](#)
dataset	an R data frame generated with [generateDatasetFile](#)
flag	0, 1, or 2. 0 uses the data from the complete dataset, 1 generates one plot for each experiment, 2 generates one plot for each plate.
flag2	0 draws lines using mean and sd, 1 draws lines using median and mad.
col4plot	a character string specifying the column whose intensity values will be used for the plot
col4anno	in case showPlot == 1, a character string specifying the column used for identifying points
plotTitle	the plot title
showPlot	0 or 1. 1 will open one or several plot windows in the R GUI, 0 will only save the plot(s) without opening windows.

Value

Saves the plots in pdf and png files named after the experiment name specified in the header concatenated with the plotTitle and if applicable the experiment number and/or the plate number.

When flag == 0, returns the plot name (plotName).

When flag == 1, returns a list containing:

plotName	The plot name
minOfScreens	The number of the first experiment
numOfScreens	The number of the last experiment

When flag == 2, returns a list containing: the plot name, a vector with the number of the first experiment and of the last experiment, and a vector with the number of the first plate and the number of the last plate.

See Also

[plotBar](#), [ZScorePlot](#)

Examples

```
data(exampleHeader, package="RNAither")
data(exampleDataset, package="RNAither")

plotname <- ZScorePlotTwo(header, dataset, 0, 1, "SigIntensity", "GeneName", "Data per well", 0)
```

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