

# Package ‘StructuralVariantAnnotation’

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**Type** Package

**Title** Variant annotations for structural variants

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**Description** StructuralVariantAnnotation provides a framework for analysis of structural variants within the Bioconductor ecosystem. This package contains contains useful helper functions for dealing with structural variants in VCF format. The packages contains functions for parsing VCFs from a number of popular callers as well as functions for dealing with breakpoints involving two separate genomic loci encoded as GRanges objects.

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**Depends** GenomicRanges, rtracklayer, VariantAnnotation, BiocGenerics, R (>= 4.1.0)

**Imports** assertthat, Biostrings, palign, stringr, dplyr, methods, rlang, GenomicFeatures, IRanges, S4Vectors, SummarizedExperiment, GenomeInfoDb,

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---

align_breakpoints	<i>Adjusting the nominal position of a pair of partnered breakpoint.</i>
-------------------	--

---

### Description

Adjusting the nominal position of a pair of partnered breakpoint.

### Usage

```
align_breakpoints(
  vcf,
  align = c("centre"),
  is_higher_breakend = names(vcf) < info(vcf)$PARID
)
```

**Arguments**

vcf                    A VCF object.  
align                 The alignment type.  
is\_higher\_breakend   Breakpoint ID ordering.

**Value**

A VCF object with adjusted nominal positions.

---

breakendRanges	<i>Extracting unpartnered breakend structural variants as a GRanges</i>
----------------	---

---

**Description**

Extracting unpartnered breakend structural variants as a GRanges

**Usage**

```
breakendRanges(x, ...)

## S4 method for signature 'VCF'
breakendRanges(x, ...)
```

**Arguments**

x                    A VCF object.  
...                  Parameters of `.breakpointRanges()`. See `breakpointRanges` for more details.

**Details**

The VCF standard supports single breakends where a breakend is not part of a novel adjacency and lacks a mate. This function supports parsing single breakends to GRanges, where a dot symbol is used in the ALT field to annotate the directional information. Single breakends provide insights to situations when one side of the structural variant is not observed, due to e.g. low mappability, non-reference contigs, complex multi-break operations, etc. See Section 5.4.9 of <https://samtools.github.io/hts-specs/VCFv4.3.pdf> for details of single breakends.

**Value**

A GRanges object of SVs.

**Methods (by class)**

- VCF: Extracting unpartnered structural variants as GRanges.

**Examples**

```
vcf.file <- system.file("extdata", "gridss.vcf",
                       package = "StructuralVariantAnnotation")
vcf <- VariantAnnotation::readVcf(vcf.file, "hg19")
breakendRanges(vcf)
breakendRanges(vcf, nominalPosition=TRUE)
```

---

breakpointgr2bedpe      *Converting breakpoint GRanges to BEDPE-like dataframe*

---

### Description

Converting breakpoint GRanges to BEDPE-like dataframe

### Usage

```
breakpointgr2bedpe(gr)
```

### Arguments

`gr`                      A GRanges object.

### Details

`breakpointgr2bedpe` converts a breakpoint GRanges to a BEDPE-formatted dataframe. The BEDPE format consists of two sets of genomic loci, optional columns of name, score, strand1, strand2 and any user-defined fields. See <https://bedtools.readthedocs.io/en/latest/content/general-usage.html> for more details on the BEDPE format.

### Value

A BEDPE-formatted data frame.

### Examples

```
#converting a GRanges object to BEDPE-like dataframe
vcf.file <- system.file("extdata", "gridss.vcf", package = "StructuralVariantAnnotation")
vcf <- VariantAnnotation::readVcf(vcf.file, "hg19")
gr <- breakpointRanges(vcf)
breakpointgr2bedpe(gr)
```

---

breakpointgr2pairs      *Converts a breakpoint GRanges object to a Pairs object*

---

### Description

Converts a breakpoint GRanges object to a Pairs object

Converts a BEDPE Pairs containing pairs of GRanges loaded using to a breakpoint GRanges object.

**Usage**

```
breakpointgr2pairs(
  bpgr,
  writeQualAsScore = TRUE,
  writeName = TRUE,
  bedpeName = NULL,
  firstInPair = NULL
)

pairs2breakpointgr(
  pairs,
  placeholderName = "bedpe",
  firstSuffix = "_1",
  secondSuffix = "_2",
  nameField = "name",
  renameScoreToQUAL = TRUE
)
```

**Arguments**

<code>bpgr</code>	breakpoint GRanges object
<code>writeQualAsScore</code>	write the breakpoint GRanges QUAL field as the score fields for compatibility with BEDPE rtracklayer export
<code>writeName</code>	write the breakpoint GRanges QUAL field as the score fields for compatibility with BEDPE rtracklayer export
<code>bedpeName</code>	function that returns the name to use for the breakpoint. Defaults to the sourceId, name column, or row names (in that priority) of the first breakend of each pair.
<code>firstInPair</code>	function that returns TRUE for breakends that are considered the first in the pair, and FALSE for the second in pair breakend. By default, the first in the pair is the breakend with the lower ordinal in the breakpoint GRanges object.
<code>pairs</code>	a Pairs object consisting of two parallel genomic loci.
<code>placeholderName</code>	prefix to use to ensure each entry has a unique ID.
<code>firstSuffix</code>	first in pair name suffix to ensure breakend name uniqueness
<code>secondSuffix</code>	second in pair name suffix to ensure breakend name uniqueness
<code>nameField</code>	Fallback field for row names if the Pairs object does not contain any names. BEDPE files loaded using rtracklayer use the "name" field.
<code>renameScoreToQUAL</code>	renames the 'score' column to 'QUAL'. Performing this rename results in a consistent variant quality score column name for variant loaded from BEDPE and VCF.

**Details**

Breakpoint-level column names will override breakend-level column names.

**Value**

Pairs GRanges object suitable for export to BEDPE by rtracklayer

Breakpoint GRanges object.

**Examples**

```
vcf.file <- system.file("extdata", "gridss.vcf", package = "StructuralVariantAnnotation")
bpgr <- breakpointRanges(VariantAnnotation::readVcf(vcf.file))
pairgr <- breakpointgr2pairs(bpgr)
#rtracklayer::export(pairgr, con="example.bedpe")
bedpe.file <- system.file("extdata", "gridss.bedpe", package = "StructuralVariantAnnotation")
bedpe.pairs <- rtracklayer::import(bedpe.file)
bedpe.bpgr <- pairs2breakpointgr(bedpe.pairs)
```

---

**breakpointGRangesToVCF**

*Converts the given breakpoint GRanges object to VCF format in breakend notation.*

---

**Description**

Converts the given breakpoint GRanges object to VCF format in breakend notation.

**Usage**

```
breakpointGRangesToVCF(gr, ...)
```

**Arguments**

<code>gr</code>	breakpoint GRanges object. Can contain both breakpoint and single breakend SV records.
<code>...</code>	For <code>cbind</code> and <code>rbind</code> a list of VCF objects. For all other methods ... are additional arguments passed to methods. See VCF class in VariantAnnotation for more details.

**Value**

A VCF object.

---

**breakpointRanges**

*Extracting the structural variants as a GRanges.*

---

**Description**

Extracting the structural variants as a GRanges.

`.breakpointRanges()` is an internal function for extracting structural variants as GRanges.

**Usage**

```
breakpointRanges(x, ...)

## S4 method for signature 'VCF'
breakpointRanges(x, ...)

.breakpointRanges(
  vcf,
  nominalPosition = FALSE,
  placeholderName = "svrecord",
  suffix = "_bp",
  info_columns = NULL,
  unpartneredBreakends = FALSE,
  inferMissingBreakends = FALSE,
  ignoreUnknownSymbolicAlleles = FALSE
)
```

**Arguments**

x	A VCF object
...	Parameters of .breakpointRanges(). See below.
vcf	A VCF object.
nominalPosition	Determines whether to call the variant at the nominal VCF position, or to call the confidence interval (incorporating any homology present). Default value is set to FALSE, where the interval is called based on the CIPOS tag. When set to TRUE, the ranges field contains the nominal variant position only.
placeholderName	Variant name prefix to assign to unnamed variants.
suffix	The suffix to append to variant names.
info_columns	VCF INFO columns to include in the GRanges object.
unpartneredBreakends	Determining whether to report unpartnered breakends. Default is set to FALSE.
inferMissingBreakends	Infer missing breakend records from ALT field of records without matching partners
ignoreUnknownSymbolicAlleles	Ignore unknown symbolic alleles. StructuralVariantAnnotation currently handles INS, INV, DEL, DUP as well as the VCF specifications non-compliant RPL, TRA symbolic alleles.

**Details**

Structural variants are converted to breakend notation. Due to ambiguities in the VCF specifications, structural variants with multiple alt alleles are not supported. The CIPOS tag describes the uncertainty interval around the position of the breakend. See Section 5.4.8 of <https://samtools.github.io/hts-specs/VCFv4.3.pdf> for details of CIPOS. If HOMLEN or HOMSEQ is defined without CIPOS, it is assumed that the variant position is left aligned. A breakend on the '+' strand indicates a break immediately after the given position, to the left of which is the DNA segment involved in the breakpoint. The '-' strand indicates a break immediately before the given position, rightwards of which is the DNA segment involved in the breakpoint. Unpaired variants are removed at this stage.

**Value**

A GRanges object of SVs.

**Methods (by class)**

- VCF: Extracting structural variants as GRanges.

**Examples**

```
vcf.file <- system.file("extdata", "vcf4.2.example.sv.vcf",
                        package = "StructuralVariantAnnotation")
vcf <- VariantAnnotation::readVcf(vcf.file, "hg19")
breakpointRanges(vcf)
breakpointRanges(vcf, nominalPosition=TRUE)
```

---

calculateReferenceHomology

*Calculates the length of inexact homology between the breakpoint sequence and the reference*

---

**Description**

Calculates the length of inexact homology between the breakpoint sequence and the reference

**Usage**

```
calculateReferenceHomology(
  gr,
  ref,
  anchorLength = 300,
  margin = 5,
  match = 2,
  mismatch = -6,
  gapOpening = 5,
  gapExtension = 3
)
```

**Arguments**

gr	reakpoint GRanges
ref	reference BSgenome
anchorLength	Number of bases to consider for homology
margin	Number of additional reference bases include. This allows for inexact homology to be detected even in the presence of indels.
match	see pwalgn::pairwiseAlignment
mismatch	see pwalgn::pairwiseAlignment
gapOpening	see pwalgn::pairwiseAlignment
gapExtension	see pwalgn::pairwiseAlignment

**Value**

A dataframe containing the length of inexact homology between the breakpoint sequence and the reference.

---

`countBreakpointOverlaps`

*Counting overlapping breakpoints between two breakpoint sets*

---

**Description**

Counting overlapping breakpoints between two breakpoint sets

**Usage**

```
countBreakpointOverlaps(  
  querygr,  
  subjectgr,  
  countOnlyBest = FALSE,  
  breakpointScoreColumn = "QUAL",  
  maxgap = -1L,  
  minoverlap = 0L,  
  ignore.strand = FALSE,  
  sizemargin = NULL,  
  restrictMarginToSizeMultiple = NULL  
)
```

**Arguments**

`querygr`, `subjectgr`, `maxgap`, `minoverlap`, `ignore.strand`, `sizemargin`,  
`restrictMarginToSizeMultiple`

See `findBreakpointOverlaps()`.

`countOnlyBest` Default value set to `FALSE`. When set to `TRUE`, the result count each subject breakpoint as overlapping only the best overlapping query breakpoint. The best breakpoint is considered to be the one with the highest `QUAL` score.

`breakpointScoreColumn`

Query column defining a score for determining which query breakpoint is considered the best when `countOnlyBest=TRUE`.

**Details**

`countBreakpointOverlaps()` returns the number of overlaps between breakpoint objects, based on the output of `findBreakpointOverlaps()`. See `GenomicRanges::countOverlaps-methods`

**Value**

An integer vector containing the tabulated query overlap hits.

**Examples**

```
truth_vcf = VariantAnnotation::readVcf(system.file("extdata", "na12878_chr22_Sudmunt2015.vcf",
package = "StructuralVariantAnnotation"))
crest_vcf = VariantAnnotation::readVcf(system.file("extdata", "na12878_chr22_crest.vcf",
package = "StructuralVariantAnnotation"))
caller_bpgr = breakpointRanges(crest_vcf)
caller_bpgr$true_positive = countBreakpointOverlaps(caller_bpgr, breakpointRanges(truth_vcf),
maxgap=100, sizemargin=0.25, restrictMarginToSizeMultiple=0.5, countOnlyBest=TRUE)
```

---

elementExtract	<i>Extracts the element of each element at the given position</i>
----------------	---

---

**Description**

Extracts the element of each element at the given position

**Usage**

```
elementExtract(x, offset = 1)
```

**Arguments**

x	list-like object
offset	offset of list

**Value**

The element of each element at given positions.

---

extractBreakpointSequence	<i>Extracts the breakpoint sequence.</i>
---------------------------	--

---

**Description**

Extracts the breakpoint sequence.

**Usage**

```
extractBreakpointSequence(gr, ref, anchoredBases, remoteBases = anchoredBases)
```

**Arguments**

gr	breakpoint GRanges
ref	Reference BSgenome
anchoredBases	Number of bases leading into breakpoint to extract
remoteBases	Number of bases from other side of breakpoint to extract

**Details**

The sequence is the sequenced traversed from the reference anchor bases to the breakpoint. For backward (-) breakpoints, this corresponds to the reverse compliment of the reference sequence bases.

**Value**

Breakpoint sequence around the variant position.

---

extractReferenceSequence

*Returns the reference sequence around the breakpoint position*

---

**Description**

Returns the reference sequence around the breakpoint position

**Usage**

```
extractReferenceSequence(  
  gr,  
  ref,  
  anchoredBases,  
  followingBases = anchoredBases  
)
```

**Arguments**

gr	breakpoint GRanges
ref	Reference BSgenome
anchoredBases	Number of bases leading into breakpoint to extract
followingBases	Number of reference bases past breakpoint to extract

**Details**

The sequence is the sequenced traversed from the reference anchor bases to the breakpoint. For backward (-) breakpoints, this corresponds to the reverse compliment of the reference sequence bases.

**Value**

Reference sequence around the breakpoint position.

---

 findBreakpointOverlaps

*Finding overlapping breakpoints between two breakpoint sets*


---

### Description

Finding overlapping breakpoints between two breakpoint sets

### Usage

```
findBreakpointOverlaps(
  query,
  subject,
  maxgap = -1L,
  minoverlap = 0L,
  ignore.strand = FALSE,
  sizemargin = NULL,
  restrictMarginToSizeMultiple = NULL
)
```

### Arguments

- `query, subject` Both of the input objects should be GRanges objects. Unlike `findOverlaps()`, `subject` cannot be omitted. Each breakpoint must be accompanied with a partner breakend, which is also in the GRanges, with the partner's id recorded in the partner field. See `GenomicRanges::findOverlaps-methods` for details.
- `maxgap, minoverlap` Valid overlapping thresholds of a maximum gap and a minimum overlapping positions between breakend intervals. Both should be scalar integers. `maxgap` allows non-negative values, and `minoverlap` allows positive values. See `GenomicRanges::findOverlaps-methods` for details.
- `ignore.strand` Default value is FALSE. strand information is ignored when set to TRUE. See `GenomicRanges::findOverlaps-methods` for details.
- `sizemargin` Error margin in allowable size to prevent matching of events of different sizes, e.g. a 200bp event matching a 1bp event when `maxgap` is set to 200.
- `restrictMarginToSizeMultiple` Size restriction multiplier on event size. The default value of 0.5 requires that the breakpoint positions can be off by at maximum, half the event size. This ensures that small deletion do actually overlap at least one base pair.

### Details

`findBreakpointOverlaps()` is an efficient adaptation of `findOverlaps-methods()` for breakend ranges. It searches for overlaps between breakpoint objects, and return a matrix including index of overlapping ranges as well as error stats. All breakends must have their partner breakend included in the partner field. A valid overlap requires that breakends on both sides meets the overlapping requirements.

See `GenomicRanges::findOverlaps-methods` for details of overlap calculation.

**Value**

A dataframe containing index and error stats of overlapping breakpoints.

**Examples**

```
#reading in VCF files
query.file <- system.file("extdata", "gridss-na12878.vcf", package = "StructuralVariantAnnotation")
subject.file <- system.file("extdata", "gridss.vcf", package = "StructuralVariantAnnotation")
query.vcf <- VariantAnnotation::readVcf(query.file, "hg19")
subject.vcf <- VariantAnnotation::readVcf(subject.file, "hg19")
#parsing vcfs to GRanges objects
query.gr <- breakpointRanges(query.vcf)
subject.gr <- breakpointRanges(subject.vcf)
#find overlapping breakpoint intervals
findBreakpointOverlaps(query.gr, subject.gr)
findBreakpointOverlaps(query.gr, subject.gr, ignore.strand=TRUE)
findBreakpointOverlaps(query.gr, subject.gr, maxgap=100, sizemargin=0.5)
```

---

findInsDupOverlaps	<i>Finds duplication events that are reported as inserts. As sequence alignment algorithms do not allow backtracking, long read-based variant callers will frequently report small duplication as insertion events. Whilst both the duplication and insertion representations result in the same sequence, this representational difference is problematic when comparing variant call sets.</i>
--------------------	--

---

**Description**

WARNING: this method does not check that the inserted sequence actually matched the duplicated sequence.

**Usage**

```
findInsDupOverlaps(query, subject, maxgap = -1L, maxsizedifference = 0L)
```

**Arguments**

query	a breakpoint GRanges object
subject	a breakpoint GRanges object
maxgap	maximum distance between the insertion position and the duplication
maxsizedifference	maximum size difference between the duplication and insertion.

**Value**

Hits object containing the ordinals of the matching breakends in the query and subject

---

findTransitiveCalls     *Identifies potential transitive imprecise calls that can be explained by traversing multiple breakpoints.*

---

### Description

Transitive calls are imprecise breakpoints or breakpoints with inserted sequence that can be explained by a sequence of breakpoints. That is, A-C calls in which additional sequence may be between A and C that can be explained by A-B-C.

### Usage

```
findTransitiveCalls(
  transitiveGr,
  subjectGr,
  maximumImpreciseInsertSize = 700,
  minimumTraversedBreakpoints = 2,
  maximumTraversedBreakpoints = 6,
  positionalMargin = 8,
  insertionLengthMargin = 50,
  insLen = transitiveGr$insLen,
  impreciseTransitiveCalls = (transitiveGr$HOMLEN == 0 | is.null(transitiveGr$HOMLEN))
    & start(transitiveGr) != end(transitiveGr),
  impreciseSubjectCalls = (subjectGr$HOMLEN == 0 | is.null(subjectGr$HOMLEN)) &
    start(subjectGr) != end(subjectGr),
  allowImprecise = FALSE
)
```

### Arguments

transitiveGr	a breakpoint GRanges object containing imprecise calls
subjectGr	breakpoints to traverse
maximumImpreciseInsertSize	Expected number of bases to traverse imprecise calls.
minimumTraversedBreakpoints	Minimum number of traversed breakpoints to consider a transitive
maximumTraversedBreakpoints	Maximum number of breakpoints to traverse when looking for an explanation of the transitive calls
positionalMargin	Allowable margin of error when matching call positional overlaps. A non-zero margin allows for matching of breakpoint with imperfect homology.
insertionLengthMargin	Allowable difference in length between the inserted sequence and the traversed path length. Defaults to 50bp to allow for long read indel errors.
insLen	Integer vector of same length as ‘transitiveGr’ indicating the number of bases inserted at the breakpoint. Defaults to transitiveGr\$insLen which will be present if the GRanges was loaded from a VCF using breakpointRanges()

- `impreciseTransitiveCalls` Boolean vector of same length as `'transitiveGr'` indicating which calls are imprecise calls. Defaults to calls with a non-zero interval size that have no homology.
- `impreciseSubjectCalls` Boolean vector of same length as `'subjectGr'` indicating which calls are imprecise calls. Defaults to calls with a non-zero interval size that have no homology.
- `allowImprecise` Allow traversal of imprecise calls. Defaults to `FALSE` as to prevent spurious results which skip some breakpoints when traversing multiple breakpoints E.g. An A-D transitive from an underlying A-B-C-D rearrangement will include A-B-D and A-C-D results if `allowImprecise=TRUE`.

**Value**

'DataFrame' containing the transitive calls traversed with the following columns: | column | meaning || ——— | ——— || `transitive_breakpoint_name` | Name of the transitive breakpoint a path was found for || `total_distance` | Total length (in bp) of the path || `traversed_breakpoint_names` | 'CharacterList' of names of breakpoint traversed in the path || `distance_to_traversed_breakpoint` | 'IntegerList' of distances from start of path to end of traversing breakpoint |

---

<code>hasPartner</code>	<i>Determines whether this breakend has a valid partner in this GRanges</i>
-------------------------	---

---

**Description**

Determines whether this breakend has a valid partner in this GRanges

**Usage**

```
hasPartner(gr, selfPartnerSingleBreakends = FALSE)
```

**Arguments**

`gr` GRanges object of SV breakends  
`selfPartnerSingleBreakends` treat single breakends as their own partner.

**Value**

True/False for each row in the breakpoint GRanges

**Examples**

```
#Subset to chromosome 6 intra-chromosomal events \code{vcf}
vcf.file <- system.file("extdata", "COL0829T.purple.sv.ann.vcf.gz",
  package = "StructuralVariantAnnotation")
vcf <- VariantAnnotation::readVcf(vcf.file)
gr <- breakpointRanges(vcf)
gr <- gr[seqnames(gr) == "6"]
# We now need to filter out inter-chromosomal events to ensure
# our GRanges doesn't contain any breakpoints whose partner
# has already been filtered out and no longer exists in the GRanges.
gr <- gr[hasPartner(gr)]
```

---

isStructural                      *Determining whether the variant is a structural variant*

---

### Description

Determining whether the variant is a structural variant

### Usage

```
isStructural(x, ...)

## S4 method for signature 'CollapsedVCF'
isStructural(x, ..., singleAltOnly = TRUE)

## S4 method for signature 'ExpandedVCF'
isStructural(x, ...)

## S4 method for signature 'VCF'
isStructural(x, ...)
```

### Arguments

x	A VCF object.
...	Internal parameters.
singleAltOnly	Whether only single ALT values are accepted. Default is set to TRUE.

### Details

The function takes a VCF object as input, and returns a logical value for each row, determining whether the variant is a structural variant.

### Value

A logical list of which the length is the same with the input object.

### Methods (by class)

- CollapsedVCF: Determining whether a CollapsedVCF object is a structural variant. Only single ALT values are accepted.
- ExpandedVCF: Determining whether a ExpandedVCF object is a structural variant.
- VCF: Determining whether a VCF object is a structural variant.

### Examples

```
vcf.file <- system.file("extdata", "gridss.vcf", package = "StructuralVariantAnnotation")
vcf <- VariantAnnotation::readVcf(vcf.file, "hg19")
isStructural(vcf)
```

---

isSymbolic	<i>Determining whether the variant is a symbolic allele.</i>
------------	--

---

### Description

Determining whether the variant is a symbolic allele.

### Usage

```
isSymbolic(x, ...)  
  
## S4 method for signature 'CollapsedVCF'  
isSymbolic(x, ..., singleAltOnly = TRUE)  
  
## S4 method for signature 'ExpandedVCF'  
isSymbolic(x, ...)
```

### Arguments

x	A VCF object.
...	Internal parameters.
singleAltOnly	Whether only single ALT values are accepted. Default is set to TRUE.

### Details

The function takes a VCF object as input, and returns a logical value for each row, determining whether the variant is a symbolic allele.

### Value

A logical list of which the length is the same with the input object.

### Methods (by class)

- `CollapsedVCF`: Determining whether a `CollapsedVCF` object is a symbolic allele. Only single ALT values are accepted.
- `ExpandedVCF`: Determining whether a `ExpandedVCF` object is a symbolic allele

### Examples

```
vcf.file <- system.file("extdata", "gridss.vcf", package = "StructuralVariantAnnotation")  
vcf <- VariantAnnotation::readVcf(vcf.file, "hg19")  
isSymbolic(vcf)
```

---

numtDetect	<i>Detecting nuclear mitochondria fusion events.</i>
------------	--

---

### Description

Detecting nuclear mitochondria fusion events.

### Usage

```
numtDetect(gr, nonStandardChromosomes = FALSE, max_ins_dist = 1000)
```

### Arguments

gr	A GRanges object
nonStandardChromosomes	Whether to report insertion sites on non-standard reference chromosomes. Default value is set to FALSE.
max_ins_dist	The maximum distance allowed on the reference genome between the paired insertion sites. Only intra-chromosomal NUMT events are supported. Default value is 1000.

### Details

Nuclear mitochondrial fusion (NUMT) is a common event found in human genomes. This function searches for NUMT events by identifying breakpoints supporting the fusion of nuclear chromosome and mitochondrial genome. Only BND notations are supported at the current stage. Possible linked nuclear insertion sites are reported using SV IDs in the candidatePartnerId metadata column.

### Value

A GRanges object of possible NUMT loci.

### Examples

```
vcf.file <- system.file("extdata", "MT.vcf", package = "StructuralVariantAnnotation")
vcf <- VariantAnnotation::readVcf(vcf.file, "hg19")
gr <- breakpointRanges(vcf, nominalPosition=TRUE)
numt.gr <- numtDetect(gr)
```

---

partner	<i>GRanges representing the breakend coordinates of structural variants #@export Partner breakend for each breakend.</i>
---------	--

---

### Description

GRanges representing the breakend coordinates of structural variants #@export Partner breakend for each breakend.

**Usage**

```
partner(gr, selfPartnerSingleBreakends = FALSE)
```

**Arguments**

```
gr                GRanges object of SV breakends
selfPartnerSingleBreakends
                  treat single breakends as their own partner.
```

**Details**

All breakends must have their partner breakend included in the GRanges.

**Value**

A GRanges object in which each entry is the partner breakend of those in the input object.

**Examples**

```
#reading in a VCF file as \code{vcf}
vcf.file <- system.file("extdata", "gridss.vcf", package = "StructuralVariantAnnotation")
vcf <- VariantAnnotation::readVcf(vcf.file, "hg19")
#parsing \code{vcf} to GRanges object \code{gr}
gr <- breakpointRanges(vcf)
#output partner breakend of each breakend in \code{gr}
partner(gr)
```

---

rtDetect

*Detecting retrotranscript insertion in nuclear genomes.*


---

**Description**

Detecting retrotranscript insertion in nuclear genomes.

**Usage**

```
rtDetect(gr, genes, maxgap = 100, minscore = 0.3)
```

**Arguments**

```
gr                A GRanges object
genes             TxDb object of genes. hg19 and hg38 are supported in the current version.
maxgap           The maximum distance allowed on the reference genome between the paired exon
                 boundaries.
minscore         The minimum proportion of intronic deletions of a transcript should be identi-
                 fied.
```

**Details**

This function searches for retroposed transcripts by identifying breakpoints supporting intronic deletions and fusions between exons and remote loci. Only BND notations are supported at the current stage.

**Value**

A GRangesList object, named insSite and rt, reporting breakpoints supporting insert sites and retroposed transcripts respectively. 'exon' and 'txs' in the metadata columns report exon\_id and transcript\_name from the 'genes' object.

---

simpleEventLength	<i>Length of event if interpreted as an isolated breakpoint.</i>
-------------------	--

---

**Description**

Length of event if interpreted as an isolated breakpoint.

**Usage**

```
simpleEventLength(gr)
```

**Arguments**

gr                    breakpoint GRanges object

**Value**

Length of the simplest explanation of this breakpoint/breakend.

---

simpleEventType	<i>Type of simplest explanation of event. Possible types are:   Type   Description     BND   Single breakend     CTX   Interchromosomal translocation     INV   Inversion.     DUP   Tandem duplication     INS   Insertion     DEL   Deletion  </i>
-----------------	--

---

**Description**

Note that both ++ and – breakpoint will be classified as inversions regardless of whether both breakpoint that constitute an actual inversion exists or not

**Usage**

```
simpleEventType(gr, insertionLengthThreshold = 0.5)
```

**Arguments**

gr                    breakpoint GRanges object  
insertionLengthThreshold  
                      portion of inserted bases compared to total event size to be classified as an insertion. For example, a 5bp deletion with 5 inserted bases will be classified as an INS event.

**Value**

Type of simplest explanation of event

---

StructuralVariantAnnotation

*StructuralVariantAnnotation: a package for SV annotation*

---

## Description

StructuralVariantAnnotation contains useful helper functions for reading and interpreting structural variants calls. The package contains functions for parsing VCFs from a number of popular callers as well as functions for dealing with breakpoints involving two separate genomic loci. The package takes a 'GRanges' based breakend-centric approach.

## Details

\* Parse VCF objects with the 'breakpointRanges()' and 'breakendRanges()' functions. \* Find breakpoint overlaps with the 'findBreakpointOverlaps()' and 'countBreakpointOverlaps()' functions. \* Generate BEDPE files for circos plot with 'breakpointgr2pairs()' function. \* ...

For more details on the features of StructuralVariantAnnotation, read the vignette: 'browseVignettes(package = "StructuralVariantAnnotation")'

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