## Package 'cytoMEM'

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Type Package

Title Marker Enrichment Modeling (MEM)

Version 1.13.0

Description MEM, Marker Enrichment Modeling, automatically generates and displays quantitative labels for cell populations that have been identified from single-cell data. The input for MEM is a dataset that has pre-clustered or pre-gated populations with cells in rows and features in columns. Labels convey a list of measured features and the features' levels of relative enrichment on each population. MEM can be applied to a wide variety of data types and can compare between MEM labels from flow cytometry, mass cytometry, single cell RNA-seq, and spectral flow cytometry using RMSD.

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Imports gplots, tools, flowCore, grDevices, stats, utils, matrixStats, methods

## **Collate**

 $"MEM\_function.R" create\_labels\_txt.R" IQR\_thresh.R" build\_heatmaps.R" add\_cluster\_ID.R" add\_file ID\_to\_cluster ID.R" characteristics and the support of the property of the$ 

biocViews Proteomics, SystemsBiology, Classification, FlowCytometry,

DataRepresentation, DataImport, CellBiology, SingleCell, Clustering

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Suggests knitr, rmarkdown

VignetteBuilder knitr

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2 build\_heatmaps

```
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## Description

Takes matrix list generated from MEM as input and outputs MEM labels, heatmap of population median values, and heatmap of MEM scores.

## Usage

```
build_heatmaps(
    MEM_values,
    cluster.MEM = "both",
    cluster.medians = "none",
    cluster.IQRs = "none",
    display.thresh = 1,
    output.files = FALSE,
    labels = FALSE,
    only.MEMheatmap = FALSE)
```

## Arguments

MEM\_values

cluster.MEM

"none","row","col",or "both"; which dimension(s) of the MEM heatmap to hierarchically cluster. See Details for more information.

cluster.medians

"none","row","col",or "both"; which dimension(s) of the median heatmap to cluster. If "none", the median row and column order will match order of the MEM heatmap.

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"none", "row", "col", or "both"; which dimension(s) of the IQR heatmap to cluster.IQRs cluster. If "none", the IQR row and column order will match order of the MEM heatmap. display.thresh Numeric; 0-10. Markers with enrichment scores that are equal to or greater than display. thresh will be displayed as row names on MEM heatmap. TRUE or FALSE; If TRUE, output of build\_heatmaps will be written to folder output.files output folder that is created as a subdirectory of the working directory. Written files include a PDF of the MEM heatmap as well as txt files containing MEM labels (row names), MEM scores, medians, and IQR values. labels TRUE or FALSE; whether or not to include MEM labels in the MEM heatmap only.MEMheatmap TRUE or FALSE; whether to only show MEM heatmap or all relevant heatmaps (MEM, median, IQR)

#### **Details**

Heatmaps are clustered using the default complete linkage hierarchical clustering in the hclust function. See heatmap. 2 and hclust for more information.

#### Value

Heatmaps of median, IQR, and MEM values on each population; optionally written to file.

## Author(s)

Kirsten Diggins, Sierra Lima, Jonathan Irish

#### References

Diggins et al., Nature Methods, 2017

## See Also

```
MEM, heatmap. 2, hclust
```

#### **Examples**

```
# Use output from MEM function or use example data with
data(MEM_values)

build_heatmaps(
    MEM_values,
    cluster.MEM = "both",
    cluster.medians = "none",
    cluster.IQRs = "none",
    display.thresh = 1,
    output.files = TRUE,
    labels = FALSE,
    only.MEMheatmap = FALSE)
```

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MEM

Marker Enrichment Modeling

## **Description**

The MEM function takes pre-clustered, single-cell data as input and calculates relative enrichment scores for each marker on each population.

## Usage

```
MEM(exp_data,
    transform=FALSE,
    cofactor=1,
    choose.markers=FALSE,
    markers="all",
    choose.ref=FALSE,
    zero.ref=FALSE,
    rename.markers=FALSE,
    new.marker.names="none",
    file.is.clust=FALSE,
    add.fileID=FALSE,
    IQR.thresh=NULL,
    output.prescaled.MEM=FALSE,
    scale.matrix = "linear",
    scale.factor = 0)
```

## **Arguments**

exp_data l	list of file names or a matrix or o	data.frame object where the l	ast column con-

tains a numeric cluster ID for each cell (row). If exp\_data is a list of files, either each file must be one cluster or each file must contain a cluster channel (column) that specifies a numeric cluster ID for each cell (row). See details for

more information.

transform TRUE or FALSE; whether or not to apply asinh transformation to the data. De-

fault is FALSE.

cofactor numeric; if transform is TRUE, what cofactor should be applied. Default is 1.

Arcsinh transformed value = arcsinh(raw value/cofactor)

choose.markers TRUE or FALSE; whether or not the user wants to choose the markers (columns)

for analysis in the console. If data contains markers that will not be used in the analysis (e.g. SSC or FSC channels in flow data), should be set to TRUE. If FALSE, either all of the markers in the experiment data will be used in MEM or the user can pass a character string of the markers to be used in the analysis

using the function call (markers) below.

markers "all" or ex."1:2,7,11:12,25"; if the user wants to choose markers to be used

in the MEM analysis without having the console ask for a user input, enter a character string similar to the one shown in the example. The markers chosen

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> should be separated with colons or commas, without spaces spaces between. If "all", all of the markers will be used in MEM.

choose.ref

TRUE or FALSE; Default reference for each population is all other populations in the dataset. For example, in a dataset containing 7 clusters, reference for population 1 would include clusters 2-7. If set to TRUE, user will be prompted in the console to enter which cluster(s) should be used as reference instead of the default bulk non-population reference.

zero.ref

TRUE or FALSE; If set to TRUE, a zero, or synthetic negative, reference will be used for all populations. MAGref therefore is 0 and IQRref is the median IQR across all markers chosen. MEM scores will go from 0 to +10 instead of -10 to +10.

rename.markers TRUE or FALSE; if TRUE, user will be prompted to enter new column names in the console. Default FALSE. If FALSE, either the column names will not be changed or the user can pass a character string of the new column names using the function call new marker names below.

new.marker.names

"none" or ex. "CD4, CD19, HLA-DR, CD8, CD14, CD16"; if user wants new column names for channels without having the console ask for a user input, enter a character string like the one shown in the example. Each new column name should be separated by a comma, without spaces between names. If "none", the column names will not be changed.

file.is.clust

TRUE or FALSE; if multiple files are entered as input and each file contains cells from only one cluster, should be set to TRUE. This prompts function to merge data into one matrix for analysis and to add a file ID for each file that will stand in as the cluster ID. A text file indicating which file corresponds to which cluster number will be written to the output files folder (by default will be created as a subdirectory in your working directory).

add.fileID

TRUE or FALSE; if multiple files are entered but file.is.clust is FALSE, this indicates that there are multiple files but each contains cells from multiple clusters and that there is already a cluster channel included as the last column in each file. If add. fileID is TRUE, a file ID will be appended to the cluster ID so user can identify the file as well as cluster from which each population came.

IQR.thresh

Default NULL. Optionally can be set to a numeric value. See Details for more information.

output.prescaled.MEM

Default FALSE. If TRUE, creates folder in working directory called "output files" containing a TXT file with pre-scaled MEM values. The MEM matrix output by build\_heatmaps contains post-scaled (-10 to +10 scale) MEM values.

scale.matrix

Default "linear". Choose how to scale the MEM matrix. Can choose from "linear" "log" or "arcsinh" for the MEM matrix scale, apply scale, and then transform from -10 to 10 or 0 to 10.

scale.factor

Default 0. Choose the factor for the MEM matrix scaling. For example, choosing 2 will apply a log2 scale if "log" is chosen for scale.matrix, if "arcsinh"" is chosen then choosing 2 for scale. factor will use arcsinh scale with a cofactor of 2.

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#### **Details**

For each population and its reference, MEM first calculates median marker levels and marker interquartile ranges (IQR), and then calculates MEM scores according to the equation

```
MEM = |Median_Pop - Median_Ref| + IQR_Ref/IQR_Pop -1; if Median_Pop - Median_ref < 0, -MEM
```

A dataset is provided as an example to be used with MEM and build\_heatmaps. Please see dataset PBMC for more details.

Input data can be file type .txt, .fcs, or .csv. A matrix or data.frame object where the last column contains cluster identy per cell is also accepted. In all cases, the expected data structure is cells (datapoints) in rows and measured markers (i.e. features, parameters) in columns of the input data.

IQR threshold: The MEM equation takes the ratio of population and reference IQRs and adds this value to the difference in medians. Low IQR values below 1, like those resulting from background noise level measurements, can therefore artificially inflate the overall MEM score. In order to correct this, a threshold of 0.5 is automatically applied. However, the function can calculate an IQR threshold using the input data. If IQR\_thresh is set to "auto", the threshold will be calculated as the IQR associated with the 2nd quartile median value across all populations and corresponding reference populations. This should be used if the user anticipates that 0.5 will not be an adequate threshold for the particular dataset.

## Value

MAGpop	Matrix; Median ex	xpression level	of markers on	each population
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MAGref Matrix; Median expression on each population's corresponding reference popu-

ation

IQRpop Matrix; IQR of markers on each population

IQRref Matrix; IQR on each population's corresponding reference population

#### Note

The object generated from MEM is meant to be passed to build\_heatmaps which will generate MEM labels and heatmaps.

## Author(s)

Kirsten Diggins, Sierra Lima, and Jonathan Irish

## References

Diggins et al., Nature Methods, 2017

#### See Also

build\_heatmaps

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## **Examples**

```
## For multiple file input, set working directory to folder containing files, then
## infiles <- dir()
## For single file or object input (e.g. PBMC), input data directly into MEM function
## User inputs
data(PBMC)
MEM_values = MEM(
              PBMC,
              transform=TRUE,
              cofactor=15,
              choose.markers=FALSE,
              markers="all",
              choose.ref=FALSE,
              zero.ref = FALSE,
              rename.markers=FALSE,
              new.marker.names="none",
              IQR.thresh=NULL,
              output.prescaled.MEM=FALSE,
              scale.matrix = "linear",
              scale.factor = 0)
```

MEM\_matrix

MEM matrix

## **Description**

This matrix is the output generated from MEM analysis of the PBMC dataset. It is meant to be used as input for the MEM\_RMSD function to generate RMSD scores of similarity.

## Usage

```
data(MEM_matrix)
```

## **Format**

The format is the 7 populations in rows and the MEM scores for all 25 measured markers in columns. See PBMC dataset for more details.

## **Examples**

```
data(MEM_matrix)
```

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MEM\_RMSD

MEM RMSD similarity between populations

## **Description**

MEM\_RMSD calculates a normalized average RMSD score pairwise between populations given their MEM scores as input. This is meant to serve as a metric of similarity between populations.

The function calculates the sum of squares for all shared markers between two populations, then takes the square root of the average.

For "a" through n markers, the sum of squares is calculated as: sum of squares =  $(a2-a1)^2 + (b2-b1)^2 ...(n2-n1)^2$ 

Root-mean-square deviation (RMSD) is calculated as: RMSD = sqrt(sum of squares/number of markers)

The RMSD values are then converted to percentages with the maximum RMSD in the matrix set as 100 percent, so that the final RMSD score is the percent of the maximum RMSD.

 $Percent_max_RMSD = 100-RMSD/max_RMSD*100$ 

The function then outputs a clustered heatmap of Percent\_max\_RMSD values and the matrix of numerical values used to build the heatmap.

## Usage

```
MEM_RMSD(
   MEM_matrix,
   format=NULL,
   output.matrix=FALSE)
```

## **Arguments**

MEM\_matrix The input to MEM\_RMSD can be either 1) a matrix of values, where popula-

tions are in rows and their MEM scores are in columns, 2) the list of matrices output by MEM, or 3) a file path pointing to a folder containing tab-delimited text files, one file for each population, where each file lists marker names in the first

column and the corresponding MEM scores in the second column.

format Default is NULL. When format is equal to "pop files", the function expects a file

path as input where the designated folder contains one file for each population's

set of MEM scores.

output.matrix If TRUE, the RMSD heatmap in PDF format and txt file with matrix of values

calculated by the function will be output and located in a folder called "output

files" that is generated in the working directory.

## **Details**

If you are calculating MEM\_RMSD on population files, populations do not have to include all of the same markers. The function will determine which markers each pair of populations has in common

MEM\_values 9

and will use those common markers to calculate RMSD. If the populations have no markers in common, the function will terminate with an error. Note that population names must match exactly between files in order for them to be considered the same.

#### Value

RMSD\_vals Matrix of the calculated pairwise percent max RMSD scores

RMSD heatmap Hierarchically clustered heatmap of RMSD\_vals

## Author(s)

Kirsten Diggins, Sierra Lima, Jonathan Irish

## References

Diggins et al., Nature Methods, 2017

## **Examples**

```
## For single matrix, input data directly into RMSD function
## User inputs
data(MEM_matrix)

MEM_RMSD(
    MEM_matrix,
    format=NULL,
    output.matrix=FALSE)
```

MEM\_values

MEM values

## Description

This list of 5 matrices is the output generated from MEM analysis of the PBMC dataset. It is meant to be used as input for the build\_heatmaps function to generate population-specific MEM labels and clustered median and MEM score heatmaps.

## Usage

```
data(MEM_values)
```

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#### **Format**

#### **Examples**

data(MEM\_values)

**PBMC** 

Normal Human Peripheral Blood Mononuclear Cells (PBMCs)

## **Description**

This dataset contains 49651 events from a 25-marker panel CyTOF analysis of normal human PBMCs. Expression values are the raw (pre-transformation) median intensity (MI) values.

Data has been pre-gated to include only DNA-intercalator (Iridium) positive and CD45-high events. Expert biaxial gating was used to separate these events into 7 major blood cell populations: CD4+ T cells (cluster 1), CD8+ T cells (cluster 2), IgM+ B cells (cluster 5), IgM- B cells (cluster 4), dendritic cells (DCs) (cluster 3), natural killer (NK) cells (cluster 7), and monocytes (cluster 6). Per-cell population identity is specified in the cluster channel (variable).

This dataset is meant to be used as an example with the MEM package.

See refs for experimental protocol and further details.

## Usage

```
data("PBMC")
```

## **Format**

A data frame with 49651 observations on the following 26 variables.

The following 25 surface markers were measured by CyTOF.

CD19 B cell receptor (BCR)

CD117 c-Kit; RTK expressed by stem and progenitor cells

CD11b ITGAM, macrophage-1 antigen; complement receptor 3

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- CD4 T cell receptor (TCR) co-receptor; binds antigens presented by MHC II
- CD8 T cell receptor (TCR) co-receptor; binds antigens presented by MHC I
- CD20 B cell surface protein
- CD34 surface protein expressed by hemotopoeitic stem cells and lost over course of differentiation
- CD61 surface marker expressed on platelets
- CD123 Interleukin-3 receptor; expressed by progenitor cells
- CD45RA CD45 isoform expressed by Naive T lymphocytes
- CD45 protein tyrosine phosphatase; expressed by all mature leukocytes
- CD10 membrane metallo-endopeptidase expressed by common lymphoid progenitors
- CD33 Siglec-3; expressed by myeloid cells
- CD11c complement receptor; highly expressed on dendritic cells and myeloid cells
- CD14 pattern recognition receptor expressed by innate lymphoid cells
- CD69 involved in signaling and proliferation of activated t-lymphocytes and natural killer cells
- CD15 plays role in phagocytosis and chemotaxis; expressed in multiple blood cell malignancies
- CD16 low affinity Fc receptor for IgG; expressed by natural killer cells, neutrophils, and myeloid cells
- CD44 cell adhesion molecule and hyaluronic acid receptor
- CD38 highly expressed on germinal center B cells and plasma cells
- CD25 IL-2 receptor; expressed by activated T cells
- CD3 T cell receptor (TCR)
- IgM heavy chain isoform of BCR
- HLADR MHC class II receptor
- CD56 NCAM; expressed by natural killer cells
- cluster 1: CD4+ T cells 2: CD8+ T cells 3: Dendritic cells (DCs) 4: IgM- B cells 5: IgM+ B cells 6: Monocytes 7: Natural killer (NK) cells

## **Details**

The dataset should be arcsinh transformed with cofactor of 15. See MEM for more details.

#### Source

Leelatian et al., Methods Mol Biol, 2015.

## References

Diggins et al., Methods, 2016. Diggins et al., Nature Methods, 2017

## **Examples**

data(PBMC)

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