

Package ‘ggseqalign’

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Type Package

Title Minimal Visualization of Sequence Alignments

Version 1.2.0

Description Simple visualizations of alignments of DNA or AA sequences as well as arbitrary strings. Compatible with Biostrings and ggplot2. The plots are fully customizable using ggplot2 modifiers such as theme().

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BugReports <https://github.com/simeross/ggseqalign/issues>

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`ggseqalign-package` *ggseqalign: Minimal Visualization of Sequence Alignments*

Description

Simple visualizations of alignments of DNA or AA sequences as well as arbitrary strings. Compatible with Biostrings and ggplot2. The plots are fully customizable using ggplot2 modifiers such as `theme()`.

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See Also

Useful links:

- <https://github.com/simeross/ggseqalign>
- Report bugs at <https://github.com/simeross/ggseqalign/issues>

`alignment_table` *Alignment Table*

Description

Generate a table of mismatches and indels between one or many query sequences and a subject sequence.

Usage

```
alignment_table(query = XStringSet, subject = XStringSet, ...)
```

Arguments

<code>query</code>	A string or vector of strings or object of class XStringSet containing the query sequences/strings.
<code>subject</code>	A string or object of class XStringSet containing the subject sequence/string. Must be of length 1.
<code>...</code>	Any additional parameters are passed on to pwalign::pairwiseAlignment() . This allows for adjusting alignment algorithm and parameters.

Value

A list containing tibbles with information on mismatches and indels.

Examples

```
query_seq <- Biostrings::DNAStringSet(c("ACCGTACCTGG", "ACCTTGG"))
subject_seq <- Biostrings::DNAStringSet("ACCGTACCGGG")
alignment_table(query_seq, subject_seq)

# Works with any string
query_string <- c("boo", "fizzbuzz")
subject_string <- "boofizz"
alignment_table(query_string, subject_string)
```

plot_sequence_alignment

Plot sequence alignment

Description

This function generates a sequence alignment plot using ggplot2 based on the input alignment table.

Usage

```
plot_sequence_alignment(
  alignment_tbl = alignment_table(query, subject),
  insertion_color = "#21918c",
  hide_mismatches = FALSE
)
```

Arguments

`alignment_tbl` An alignment table containing query and subject information for sequence alignment. Generated with [alignment_table\(\)](#).

`insertion_color` The color to use for indicating insertions in the alignment. Default is '#21918c'. Can be any output of colors() or hex code.

`hide_mismatches` A logical value indicating whether to hide mismatches in the alignment plot. Default is FALSE.

Value

A ggplot object of the sequence alignment plot.

Examples

```

q <- (c("boo", "fibububuzz", "bozz", "baofuzz"))
s <- "boofizz"

alignment <- alignment_table(q, s)
pl1 <- plot_sequence_alignment(alignment_tbl = alignment)
pl1

# Provide names for (some) query and subject elements to label the y-axis
names(q) <- c("Seq1", NA, "Seq3")
names(s) <- "reference"
pl2 <- plot_sequence_alignment(alignment_table(q, s))
pl2

# Compatible with StringSets from Biostrings
library(Biostrings)

dna <- readDNAStringSet(system.file("extdata", "dm3_upstream2000.fa.gz",
                                     package = "Biostrings"
                                     ))
# The entries dna[2:5] are identical
q <- dna[2:4]
s <- dna[5]
pl3 <- plot_sequence_alignment(alignment_table(q, s))
pl3

# Let's introduce some SNPs, insertions and deletions
q[1] <- as(
  replaceLetterAt(q[[1]], c(5, 200, 400), "AGC"),
  "DNAStringSet"
)
q[2] <- as(
  c(substr(q[[2]], 300, 1500), substr(q[[2]], 1800, 2000)),
  "DNAStringSet"
)
q[3] <- as(
  replaceAt(
    q[[3]], 1500,
    paste(rep("A", 1000), collapse = ""))
),
"DNAStringSet"
)
names(q) <- c("mismatches", "deletions", "insertion")
names(s) <- substr(names(s)[1], 1, 34)

pl4 <- plot_sequence_alignment(alignment_table(q, s))
pl4

# Compatible with ggplot2 theming
library(ggplot2)

pl4 +

```

```
ylab("Sequence variants") +  
xlab("Length in bp") +  
scale_color_viridis_d() +  
theme(  
  axis.text.x = element_text(angle = 90, vjust = 0.5, hjust = 1),  
  axis.title = element_text()  
)
```

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* **internal**

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