Package 'singleCellTK'

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Type Package

Title Comprehensive and Interactive Analysis of Single Cell RNA-Seq Data

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Description The Single Cell Toolkit (SCTK) in the singleCellTK package provides an interface to popular tools for importing, quality control, analysis, and visualization of single cell RNA-seq data. SCTK allows users to seamlessly integrate tools from various packages at different stages of the analysis workflow. A general ``a la carte" workflow gives users the ability access to multiple methods for data importing, calculation of general QC metrics, doublet detection, ambient RNA estimation and removal, filtering, normalization, batch correction or integration, dimensionality reduction, 2-D embedding, clustering, marker detection, differential expression, cell type labeling, pathway analysis, and data exporting. Curated workflows can be used to run Seurat and Celda. Streamlined quality control can be performed on the command line using the SCTK-QC pipeline. Users can analyze their data using commands in the R console or by using an interactive Shiny Graphical User Interface (GUI). Specific analyses or entire workflows can be summarized and shared with comprehensive HTML reports generated by Rmarkdown. Additional documentation and vignettes can be found at camplab.net/sctk.

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Encoding UTF-8

biocViews SingleCell, GeneExpression, DifferentialExpression, Alignment, Clustering, ImmunoOncology, BatchEffect, Normalization, QualityControl, DataImport, GUI

LazyData FALSE

Imports ape, anndata, AnnotationHub, batchelor, BiocParallel, celldex, colourpicker, colorspace, cowplot, cluster, ComplexHeatmap, data.table, DelayedMatrixStats, DESeq2, dplyr, DT, ExperimentHub, ensembldb, fields, ggplot2, ggplotify, ggrepel, ggtree, gridExtra, GSVA (>= 1.50.0), GSVAdata, igraph, KernSmooth, limma, MAST, Matrix (>= 1.6-1), matrixStats, methods, msigdbr, multtest, plotly, plyr, ROCR, Rtsne,

S4Vectors, scater, scMerge (>= 1.2.0), scran, Seurat (>= 3.1.3), shiny, shinyjs, SingleR, SoupX, sva, reshape2, shinyalert, circlize, enrichR (>= 3.2), celda, shinycssloaders, DropletUtils, scds (>= 1.2.0), reticulate (>= 1.14), tools, tximport, eds, withr, GSEABase, R.utils, zinbwave, scRNAseq (>= 2.0.2), TENxPBMCData, yaml, rmarkdown, magrittr, scDblFinder, metap, VAM (>= 0.5.3), tibble, rlang, TSCAN, TrajectoryUtils, scuttle, utils, stats, zellkonverter

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Contents

`calcEffectSizes` *Finds the effect sizes for all genes in the original dataset, regardless of significance.*

Description

Finds the effect sizes for all genes in the original dataset, regardless of significance.

Usage

```
calcEffectSizes(countMatrix, condition)
```

Arguments

`countMatrix`	Matrix. A simulated counts matrix, sans labels.
`condition`	Factor. The condition labels for the simulated cells. If more than 2 conditions are given, the first will be compared to all others by default.

Value

A vector of cohen's d effect sizes for each gene.

Examples

```
data("mouseBrainSubsetSCE")
res <- calcEffectSizes(assay(mouseBrainSubsetSCE, "counts"),
                       condition = colData(mouseBrainSubsetSCE)$level1class)
```

`combineSCE` *Combine a list of SingleCellExperiment objects as one SingleCellExperiment object*

Description

Combine a list of SingleCellExperiment objects as one SingleCellExperiment object

Usage

```
combineSCE(sceList, by.r = NULL, by.c = NULL, combined = TRUE)
```

Arguments

sceList	A list contains SingleCellExperiment objects. Currently, combineSCE function only support combining SCE objects with assay in dgCMatrix format. It does not support combining SCE with assay in delayedArray format.
by.r	Specifications of the columns used for merging rowData. If set as NULL, the rownames of rowData tables will be used to merging rowData. Default is NULL.
by.c	Specifications of the columns used for merging colData. If set as NULL, the rownames of colData tables will be used to merging colData. Default is NULL.
combined	logical; if TRUE, it will combine the list of SingleCellExperiment objects and return a SingleCellExperiment. If FALSE, it will return a list of SingleCellExperiment whose rowData, colData, assay and reducedDim data slot are compatible within SCE objects in the list. Default is TRUE.

Value

A SingleCellExperiment object which combines all objects in sceList. The colData is merged.

Examples

```
data(scExample, package = "singleCellTK")
combinedsce <- combineSCE(list(sce,sce), by.r = NULL, by.c = NULL, combined = TRUE)
```

---

computeHeatmap *Computes heatmap for a set of features against dimensionality reduction components*

Description

The computeHeatmap method computes the heatmap visualization for a set of features against a set of dimensionality reduction components. This method uses the heatmap computation algorithm code from `Seurat` but plots the heatmap using `ComplexHeatmap` and `cowplot` libraries.

Usage

```
computeHeatmap(
  inSCE,
  useAssay,
  dims = 10,
  nfeatures = 30,
  cells = NULL,
  reduction = "pca",
  disp.min = -2.5,
  disp.max = 2.5,
  balanced = TRUE,
  nCol = NULL,
  externalReduction = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Specify the name of the assay that will be scaled by this function for the features that are used in the heatmap.
dims	Specify the number of dimensions to use for heatmap. Default 10.
nfeatures	Specify the number of features to use for heatmap. Default is 30.
cells	Specify the samples/cells to use for heatmap computation. Default is NULL which will utilize all samples in the assay.
reduction	Specify the reduction slot in the input object. Default is "pca".
disp.min	Specify the minimum dispersion value to use for floor clipping of assay values. Default is -2.5.
disp.max	Specify the maximum dispersion value to use for ceiling clipping of assay values. Default is 2.5.
balanced	Specify if the number of of up-regulated and down-regulated features should be balanced. Default is TRUE.
nCol	Specify the number of columns in the output plot. Default is NULL which will auto-compute the number of columns.
externalReduction	Specify an external reduction if not present in the input object. This external reduction should be created using CreateDimReducObject function.

Value

Heatmap plot object.

---

computeZScore *Compute Z-Score*

Description

Computes Z-Score from an input count matrix using the formula ((x-mean(x))/sd(x)) for each gene across all cells. The input count matrix can either be a base matrix, dgCMatrix or a DelayedMatrix. Computations are performed using DelayedMatrixStats package to efficiently compute the Z-Score matrix.

Usage

```
computeZScore(counts)
```

Arguments

counts	matrix (base matrix, dgCMatrix or DelayedMatrix)

Value

z-score computed counts matrix (DelayedMatrix)

Examples

```
data(sce_chcl, package = "scds")
assay(sce_chcl, "countsZScore") <- computeZScore(assay(sce_chcl, "counts"))
```

`constructSCE` *Create SingleCellExperiment object from csv or txt input*

Description

Create SingleCellExperiment object from csv or txt input

Usage

```
constructSCE(data, samplename)
```

Arguments

`data`	A data.table object containing the count matrix.
`samplename`	The sample name of the data.

Value

A SingleCellExperiment object containing the count matrix.

`convertSCEToSeurat` *convertSCEToSeurat Converts sce object to seurat while retaining all assays and metadata*

Description

convertSCEToSeurat Converts sce object to seurat while retaining all assays and metadata

Usage

```
convertSCEToSeurat(
  inSCE,
  countsAssay = NULL,
  normAssay = NULL,
  scaledAssay = NULL,
  copyColData = FALSE,
  copyReducedDim = FALSE,
  copyDecontX = FALSE,
  pcaReducedDim = NULL,
  icaReducedDim = NULL,
  tsneReducedDim = NULL,
  umapReducedDim = NULL
)
```

Arguments

inSCE	A SingleCellExperiment object to convert to a Seurat object.
countsAssay	Which assay to use from sce object for raw counts. Default NULL.
normAssay	Which assay to use from sce object for normalized data. Default NULL.
scaledAssay	Which assay to use from sce object for scaled data. Default NULL.
copyColData	Boolean. Whether copy 'colData' of SCE object to the 'meta.data' of Seurat object. Default FALSE.
copyReducedDim	Boolean. Whether copy 'reducedDims' of the SCE object to the 'reductions' of Seurat object. Default FALSE.
copyDecontX	Boolean. Whether copy 'decontXcounts' assay of the SCE object to the 'assays' of Seurat object. Default TRUE.
pcaReducedDim	Specify a character value indicating the name of the reducedDim to store as default pca computation in the output seurat object. Default is NULL which will not store any reducedDim as the default pca. This will only work when copyReducedDim parameter is set to TRUE.
icaReducedDim	Specify a character value indicating the name of the reducedDim to store as default ica computation in the output seurat object. Default is NULL which will not store any reducedDim as the default ica. This will only work when copyReducedDim parameter is set to TRUE.
tsneReducedDim	Specify a character value indicating the name of the reducedDim to store as default tsne computation in the output seurat object. Default is NULL which will not store any reducedDim as the default tsne. This will only work when copyReducedDim parameter is set to TRUE.
umapReducedDim	Specify a character value indicating the name of the reducedDim to store as default umap computation in the output seurat object. Default is NULL which will not store any reducedDim as the default umap. This will only work when copyReducedDim parameter is set to TRUE.

Value

Updated seurat object that contains all data from the input sce object

Examples

```
data(scExample, package = "singleCellTK")
seurat <- convertSCEToSeurat(sce)
```

---

convertSeuratToSCE *convertSeuratToSCE Converts the input seurat object to a sce object*

Description

convertSeuratToSCE Converts the input seurat object to a sce object

Usage

```
convertSeuratToSCE(
  seuratObject,
  normAssayName = "seuratNormData",
  scaledAssayName = "seuratScaledData"
)
```

Arguments

seuratObject	Input Seurat object
normAssayName	Name of assay to store the normalized data. Default "seuratNormData".
scaledAssayName	Name of assay to store the scaled data. Default "seuratScaledData".

Value

SingleCellExperiment output object

Examples

```
data(scExample, package = "singleCellTK")
seurat <- convertSCEToSeurat(sce)
sce <- convertSeuratToSCE(seurat)
```

dedupRowNames *Deduplicate the rownames of a matrix or SingleCellExperiment object*

Description

Adds '-1', '-2', ... '-i' to multiple duplicated rownames, and in place replace the unique rownames, store unique rownames in `rowData`, or return the unique rownames as character vecetor.

Usage

```
dedupRowNames(x, as.rowData = FALSE, return.list = FALSE)
```

Arguments

`x`	A matrix like or /linkS4classSingleCellExperiment object, on which we can apply `rownames()` to and has duplicated rownames.
`as.rowData`	Only applicable when `x` is a /linkS4classSingleCellExperiment object. When set to `TRUE`, will insert a new column called `"rownames.uniq"` to `rowData(x)`, with the deduplicated rownames.
`return.list`	When set to `TRUE`, will return a character vector of the deduplicated rownames.

Value

By default, a matrix or /linkS4classSingleCellExperiment object with rownames deduplicated. When `x` is a /linkS4classSingleCellExperiment and `as.rowData` is set to `TRUE`, will return `x` with `rowData` updated. When `return.list` is set to `TRUE`, will return a character vector with the deduplicated rownames.

Examples

```
data("scExample", package = "singleCellTK")
sce <- dedupRowNames(sce)
```

detectCellOutlier *Detecting outliers within the SingleCellExperiment object.*

Description

A wrapper function for isOutlier. Identify outliers from numeric vectors stored in the SingleCellExperiment object.

Usage

```
detectCellOutlier(
  inSCE,
  slotName,
  itemName,
  sample = NULL,
  nmads = 3,
  type = "both",
  overwrite = TRUE
)
```

Arguments

inSCE	A SingleCellExperiment object.
slotName	Desired slot of SingleCellExperiment used for plotting. Possible options: "assays", "colData", "metadata", "reducedDims". Required.
itemName	Desired vector within the slot used for plotting. Required.
sample	A single character specifying a name that can be found in `colData(inSCE)` to directly use the cell annotation; or a character vector with as many elements as cells to indicates which sample each cell belongs to. Default NULL. decontX will be run on cells from each sample separately.
nmads	Integer. Number of median absolute deviation. Parameter may be adjusted for more lenient or stringent outlier cutoff. Default 3.
type	Character. Type/direction of outlier detection; whether the lower/higher outliers should be detected, or both. Options are "both", "lower", "higher".
overwrite	Boolean. If TRUE, and this function has previously generated an outlier decision on the same itemName, the outlier decision will be overwritten. Default TRUE.

Value

A SingleCellExperiment object with " added to the colData slot. Additionally, the decontaminated counts will be added as an assay called 'decontXCounts'.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runDecontX(sce[,sample(ncol(sce),20)])
sce <- detectCellOutlier(sce, slotName = "colData", sample = sce$sample,
 nmads = 4, itemName = "decontX_contamination", type = "both")
```

`diffAbundanceFET` *Calculate Differential Abundance with FET*

Description

Calculate Differential Abundance with FET

Usage

```
diffAbundanceFET(inSCE, cluster, variable, control, case, analysisName)
```

Arguments

`inSCE`	A `SingleCellExperiment` object.
`cluster`	A single `character`, specifying the name to store the cluster label in `colData`.
`variable`	A single `character`, specifying the name to store the phenotype labels in `colData`.
`control`	`character`. Specifying one or more categories that can be found in the vector specified by `variable`.
`case`	`character`. Specifying one or more categories that can be found in the vector specified by `variable`.
`analysisName`	A single `character`. Will be used for naming the result table, which will be saved in metadata slot.

Details

This function will calculate the cell counting and fraction by dividing all cells to groups specified by the arguments, together with statistical summary by performing Fisher Exact Tests (FET).

Value

The original `SingleCellExperiment` object with `metadata(inSCE)` updated with a list `diffAbundanceFET`, containing a new `data.frame` for the analysis result, named by `analysisName`. The `data.frame` contains columns for number and fraction of cells that belong to different cases, as well as "Odds_Ratio", "PValue" and "FDR".

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- diffAbundanceFET(inSCE = mouseBrainSubsetSCE,
                                        cluster = "tissue",
                                        variable = "level1class",
                                        case = "oligodendrocytes",
                                        control = "microglia",
                                        analysisName = "diffAbundFET")
```

discreteColorPalette *Generate given number of color codes*

Description

Three different generation methods are wrapped, including distinctColors, [randomcoloR](SCTK_PerformingQC_Cell_V and the ggplot default color generation.

Usage

```
discreteColorPalette(
  n,
  palette = c("random", "ggplot", "celda"),
  seed = 12345,
  ...
)
```

Arguments

n	An integer, the number of color codes to generate.
palette	A single character string. Select the method, available options are "ggplot", "celda" and "random". Default "random".
seed	An integer. Set the seed for random process that happens only in "random" generation. Default 12345.
...	Other arguments that are passed to the internal function, according to the method selected.

Value

A character vector of n hex color codes.

Examples

```
discreteColorPalette(n = 3)
```

distinctColors *Generate a distinct palette for coloring different clusters*

Description

Generate a distinct palette for coloring different clusters

Usage

```
distinctColors(
  n,
 hues = c("red", "cyan", "orange", "blue", "yellow", "purple", "green", "magenta"),
  saturation.range = c(0.7, 1),
  value.range = c(0.7, 1)
)
```

Arguments

n	Integer; Number of colors to generate
hues	Character vector of R colors available from the colors() function. These will be used as the base colors for the clustering scheme. Different saturations and values (i.e. darkness) will be generated for each hue.
saturation.range	Numeric vector of length 2 with values between 0 and 1. Default: c(0.25, 1)
value.range	Numeric vector of length 2 with values between 0 and 1. Default: c(0.5, 1)

Value

A vector of distinct colors that have been converted to HEX from HSV.

Examples

```
distinctColors(10)
```

---

downSampleCells *Estimate numbers of detected genes, significantly differentially expressed genes, and median significant effect size*

Description

Estimate numbers of detected genes, significantly differentially expressed genes, and median significant effect size

Usage

```
downSampleCells(
  originalData,
  useAssay = "counts",
  minCountDetec = 10,
  minCellsDetec = 3,
  minCellnum = 10,
  maxCellnum = 1000,
  realLabels,
  depthResolution = 10,
```

```
  iterations = 10,
  totalReads = 1e+06
)
```

Arguments

originalData	The SingleCellExperiment object storing all assay data from the shiny app.
useAssay	Character. The name of the assay to be used for subsampling.
minCountDetec	Numeric. The minimum number of reads found for a gene to be considered detected.
minCellsDetec	Numeric. The minimum number of cells a gene must have at least 1 read in for it to be considered detected.
minCellnum	Numeric. The minimum number of virtual cells to include in the smallest simulated dataset.
maxCellnum	Numeric. The maximum number of virtual cells to include in the largest simulated dataset
realLabels	Character. The name of the condition of interest. Must match a name from sample data. If only two factors present in the corresponding colData, will default to t-test. If multiple factors, will default to ANOVA.
depthResolution	Numeric. How many different read depth should the script simulate? Will simulate a number of experimental designs ranging from 10 reads to maxReadDepth, with logarithmic spacing.
iterations	Numeric. How many times should each experimental design be simulated?
totalReads	Numeric. How many aligned reads to put in each simulated dataset.

Value

A 3-dimensional array, with dimensions = c(iterations, depthResolution, 3). [,,1] contains the number of detected genes in each simulated dataset, [,,2] contains the number of significantly differentially expressed genes in each simulation, and [,,3] contains the mediansignificant effect size in each simulation. If no genes are significantly differentially expressed, the median effect size defaults to infinity.

Examples

```
data("mouseBrainSubsetSCE")
subset <- mouseBrainSubsetSCE[seq(100),]
res <- downSampleCells(subset,
                       realLabels = "level1class",
                       iterations=2)
```

`downSampleDepth` *Estimate numbers of detected genes, significantly differentially expressed genes, and median significant effect size*

Description

Estimate numbers of detected genes, significantly differentially expressed genes, and median significant effect size

Usage

```
downSampleDepth(
  originalData,
  useAssay = "counts",
  minCount = 10,
  minCells = 3,
  maxDepth = 1e+07,
  realLabels,
  depthResolution = 10,
  iterations = 10
)
```

Arguments

`originalData`	SingleCellExperiment object storing all assay data from the shiny app.
`useAssay`	Character. The name of the assay to be used for subsampling.
`minCount`	Numeric. The minimum number of reads found for a gene to be considered detected.
`minCells`	Numeric. The minimum number of cells a gene must have at least 1 read in for it to be considered detected.
`maxDepth`	Numeric. The highest number of total reads to be simulated.
`realLabels`	Character. The name of the condition of interest. Must match a name from sample data.
`depthResolution`	Numeric. How many different read depth should the script simulate? Will simulate a number of experimental designs ranging from 10 reads to maxReadDepth, with logarithmic spacing.
`iterations`	Numeric. How many times should each experimental design be simulated?

Value

A 3-dimensional array, with dimensions = c(iterations, depthResolution, 3). [,,1] contains the number of detected genes in each simulated dataset, [,,2] contains the number of significantly differentially expressed genes in each simulation, and [,,3] contains the mediansignificant effect size in each simulation. If no genes are significantly differentially expressed, the median effect size defaults to infinity.

Examples

```
data("mouseBrainSubsetSCE")
subset <- mouseBrainSubsetSCE[seq(1000),]
res <- downSampleDepth(subset,
                       realLabels = "level1class",
                       iterations=2)
```

`expData` *expData Get data item from an input* `SingleCellExperiment` *object. The data item can be an* `assay`*,* `altExp` *(subset) or a* `reducedDim`*, which is retrieved based on the name of the data item.*

Description

expData Get data item from an input `SingleCellExperiment` object. The data item can be an `assay`, `altExp` (subset) or a `reducedDim`, which is retrieved based on the name of the data item.

Usage

```
expData(inSCE, assayName)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`assayName`	Specify the name of the data item to retrieve.

Value

Specified data item.

Examples

```
data(scExample, package = "singleCellTK")
mat <- expData(sce, "counts")
```

expData,ANY,character-method

expData Get data item from an input `SingleCellExperiment` *object. The data item can be an* `assay`*,* `altExp` *(subset) or a* `reducedDim`*, which is retrieved based on the name of the data item.*

Description

expData Get data item from an input `SingleCellExperiment` object. The data item can be an `assay`, `altExp` (subset) or a `reducedDim`, which is retrieved based on the name of the data item.

Usage

```
## S4 method for signature 'ANY,character'
expData(inSCE, assayName)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`assayName`	Specify the name of the data item to retrieve.

Value

Specified data item.

Examples

```
data(scExample, package = "singleCellTK")
mat <- expData(sce, "counts")
```

expData<-

expData Store data items using tags to identify the type of data item stored. To be used as a replacement for assay<- setter function but with additional parameter to set a tag to a data item.

Description

expData Store data items using tags to identify the type of data item stored. To be used as a replacement for assay<- setter function but with additional parameter to set a tag to a data item.

Usage

```
expData(inSCE, assayName, tag = NULL, altExp = FALSE) <- value
```

Arguments

inSCE	Input SingleCellExperiment object.
assayName	Specify the name of the input assay.
tag	Specify the tag to store against the input assay. Default is NULL, which will set the tag to "uncategorized".
altExp	A logical value indicating if the input assay is a altExp or a subset assay.
value	An input matrix-like value to store in the SCE object.

Value

A SingleCellExperiment object containing the newly stored data.

Examples

```
data(scExample, package = "singleCellTK")
mat <- expData(sce, "counts")
expData(sce, "counts", tag = "raw") <- mat
```

---

expData<-,ANY,character,CharacterOrNullOrMissing,logical-method

expData Store data items using tags to identify the type of data item stored. To be used as a replacement for assay<- setter function but with additional parameter to set a tag to a data item.

Description

expData Store data items using tags to identify the type of data item stored. To be used as a replacement for assay<- setter function but with additional parameter to set a tag to a data item.

Usage

```
## S4 replacement method for signature 'ANY,character,CharacterOrNullOrMissing,logical'
expData(inSCE, assayName, tag = NULL, altExp = FALSE) <- value
```

Arguments

inSCE	Input SingleCellExperiment object.
assayName	Specify the name of the input assay.
tag	Specify the tag to store against the input assay. Default is NULL, which will set the tag to "uncategorized".
altExp	A logical value indicating if the input assay is a altExp or a subset assay.
value	An input matrix-like value to store in the SCE object.

Value

A SingleCellExperiment object containing the newly stored data.

Examples

```
data(scExample, package = "singleCellTK")
mat <- expData(sce, "counts")
expData(sce, "counts", tag = "raw") <- mat
```

`expDataNames` *expDataNames Get names of all the data items in the input* `SingleCellExperiment` *object including assays, altExps and reducedDims.*

Description

expDataNames Get names of all the data items in the input `SingleCellExperiment` object including assays, altExps and reducedDims.

Usage

```
expDataNames(inSCE)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.

Value

A combined `vector` of `assayNames`, `altExpNames` and `reducedDimNames`.

Examples

```
data(scExample, package = "singleCellTK")
expDataNames(sce)
```

`expDataNames,ANY-method` *expDataNames Get names of all the data items in the input* `SingleCellExperiment` *object including assays, altExps and reducedDims.*

Description

expDataNames Get names of all the data items in the input `SingleCellExperiment` object including assays, altExps and reducedDims.

Usage

```
## S4 method for signature 'ANY'
expDataNames(inSCE)
```

Arguments

inSCE	Input SingleCellExperiment object.

Value

A combined vector of assayNames, altExpNames and reducedDimNames.

Examples

```
data(scExample, package = "singleCellTK")
expDataNames(sce)
```

---

expDeleteDataTag *expDeleteDataTag Remove tag against an input data from the stored tag information in the metadata of the input object.*

Description

expDeleteDataTag Remove tag against an input data from the stored tag information in the metadata of the input object.

Usage

```
expDeleteDataTag(inSCE, assay)
```

Arguments

inSCE	Input SingleCellExperiment object.
assay	Name of the assay or the data item against which a tag should be removed.

Value

The input SingleCellExperiment object with tag information removed from the metadata slot.

Examples

```
data(scExample, package = "singleCellTK")
sce <- expSetDataTag(sce, "raw", "counts")
sce <- expDeleteDataTag(sce, "counts")
```

exportSCE *Export data in SingleCellExperiment object*

Description

Export data in SingleCellExperiment object

Usage

```
exportSCE(
  inSCE,
  samplename = "sample",
  directory = "./",
  type = "Cells",
  format = c("SCE", "AnnData", "FlatFile", "HTAN", "Seurat")
)
```

Arguments

inSCE	A SingleCellExperiment object that contains the data. QC metrics are stored in colData of the singleCellExperiment object.
samplename	Sample name. This will be used as name of subdirectories and the prefix of flat file output. Default is 'sample'.
directory	Output directory. Default is './'.
type	Type of data. The type of data stored in SingleCellExperiment object. It can be 'Droplets'(raw droplets matrix) or 'Cells' (cells matrix).
format	The format of output. It currently supports flat files, rds files and python h5 files. It can output multiple formats. Default: c("SCE", "AnnData", "FlatFile", "HTAN").

Value

Generates a file containing data from inSCE, in specified format.

Examples

```
data(scExample)
## Not run:
exportSCE(sce, format = "SCE")

## End(Not run)
```

`exportSCEtoAnnData` *Export a SingleCellExperiment R object as Python annData object*

Description

Writes all assays, colData, rowData, reducedDims, and altExps objects in a SingleCellExperiment to a Python annData object in the .h5ad format All parameters of Anndata.write_h5ad function (https://icb-anndata.readthedocs-hosted.com/en/stable/anndata.AnnData.write_h5ad.html) are available as parameters to this export function and set to defaults. Defaults can be overridden at function call.

Usage

```
exportSCEtoAnnData(
  sce,
  useAssay = "counts",
  outputDir = "./",
  prefix = "sample",
  overwrite = TRUE,
  compression = c("gzip", "lzf", "None"),
  compressionOpts = NULL,
  forceDense = FALSE
)
```

Arguments

`sce`	SingleCellExperiment R object to be exported.
`useAssay`	Character. The name of assay of interests that will be set as the primary matrix of the output AnnData. Default `"counts"`.
`outputDir`	Path to the directory where .h5ad outputs will be written. Default is the current working directory.
`prefix`	Prefix to use for the name of the output file. Default `"sample"`.
`overwrite`	Boolean. Default `TRUE`.
`compression`	If output file compression is required, this variable accepts 'gzip', 'lzf' or "None" as inputs. Default `"gzip"`.
`compressionOpts`	Integer. Sets the compression level
`forceDense`	Default `False` Write sparse data as a dense matrix. Refer `anndata.write_h5ad` documentation for details. Default `NULL`.

Value

Generates a Python anndata object containing data from `inSCE`.

Examples

```
data(sce_chcl, package = "scds")
## Not run:
exportSCEtoAnnData(sce=sce_chcl, compression="gzip")

## End(Not run)
```

`exportSCEtoFlatFile` *Export a SingleCellExperiment object to flat text files*

Description

Writes all assays, colData, rowData, reducedDims, and altExps objects in a SingleCellExperiment to text files. The items in the 'metadata' slot remain stored in list and are saved in an RDS file.

Usage

```
exportSCEtoFlatFile(
  sce,
  outputDir = "./",
  overwrite = TRUE,
  gzipped = TRUE,
  prefix = "SCE"
)
```

Arguments

`sce`	SingleCellExperiment object to be exported.
`outputDir`	Name of the directory to store the exported file(s).
`overwrite`	Boolean. Whether to overwrite the output files. Default `TRUE`.
`gzipped`	Boolean. `TRUE` if the output files are to be gzip compressed. `FALSE` otherwise. Default `TRUE`.
`prefix`	Prefix of file names.

Value

Generates text files containing data from `inSCE`.

Examples

```
data(sce_chcl, package = "scds")
## Not run:
exportSCEtoFlatFile(sce_chcl, "sce_chcl")

## End(Not run)
```

`exportSCEToSeurat` *Export data in Seurat object*

Description

Export data in Seurat object

Usage

```
exportSCEToSeurat(
  inSCE,
  prefix = "sample",
  outputDir = "./",
  overwrite = TRUE,
  copyColData = TRUE,
  copyReducedDim = TRUE,
  copyDecontX = TRUE
)
```

Arguments

`inSCE`	A SingleCellExperiment object that contains the data. QC metrics are stored in colData of the singleCellExperiment object.
`prefix`	Prefix to use for the name of the output file. Default `"sample"`.
`outputDir`	Path to the directory where outputs will be written. Default is the current working directory.
`overwrite`	Boolean. Whether overwrite the output if it already exists in the outputDir. Default `TRUE`.
`copyColData`	Boolean. Whether copy 'colData' of SCE object to the 'meta.data' of Seurat object. Default `TRUE`.
`copyReducedDim`	Boolean. Whether copy 'reducedDims' of the SCE object to the 'reductions' of Seurat object. Default `TRUE`.
`copyDecontX`	Boolean. Whether copy 'decontXcounts' assay of the SCE object to the 'assays' of Seurat object. Default `TRUE`.

Value

Generates a Seurat object containing data from `inSCE`.

expSetDataTag *expSetDataTag Set tag to an assay or a data item in the input SCE object.*

Description

expSetDataTag Set tag to an assay or a data item in the input SCE object.

Usage

```
expSetDataTag(inSCE, assayType, assays)
```

Arguments

inSCE	Input SingleCellExperiment object.
assayType	Specify a character(1) value as a tag that should be set against a data item.
assays	Specify name(s) character() of data item(s) against which the tag should be set.

Value

The input SingleCellExperiment object with tag information stored in the metadata slot.

Examples

```
data(scExample, package = "singleCellTK")
sce <- expSetDataTag(sce, "raw", "counts")
```

expTaggedData *expTaggedData Returns a list of names of data items from the input* SingleCellExperiment *object based upon the input parameters.*

Description

expTaggedData Returns a list of names of data items from the input SingleCellExperiment object based upon the input parameters.

Usage

```
expTaggedData(
  inSCE,
  tags = NULL,
  redDims = FALSE,
  recommended = NULL,
  showTags = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
tags	A character() value indicating if the data items should be returned separated by the specified tags. Default is NULL indicating that returned names of the data items are simply returned as a list with default tag as "uncategorized".
redDims	A logical value indicating if reducedDims should be returned as well separated with 'redDims' tag.
recommended	A character() vector indicating the tags that should be displayed as recommended. Default is NULL.
showTags	A logical value indicating if the tags should be shown. If FALSE, output is just a simple list, not separated by tags.

Value

A list of names of data items specified by the other parameters.

Examples

```
data(scExample, package = "singleCellTK")
sce <- expSetDataTag(sce, "raw", "counts")
tags <- expTaggedData(sce)
```

---

featureIndex *Retrieve row index for a set of features*

Description

This will return indices of features among the rownames or rowData of a data.frame, matrix, or a SummarizedExperiment object including a SingleCellExperiment. Partial matching (i.e. grepping) can be used by setting exactMatch = FALSE.

Usage

```
featureIndex(
  features,
  inSCE,
  by = "rownames",
  exactMatch = TRUE,
  removeNA = FALSE,
  errorOnNoMatch = TRUE,
  warningOnPartialMatch = TRUE
)
```

Arguments

features	Character vector of feature names to find in the rows of inSCE.
inSCE	A data.frame, matrix, or SingleCellExperiment object to search.
by	Character. Where to search for features in inSCE. If set to "rownames" then the features will be searched for among rownames(inSCE). If inSCE inherits from class SummarizedExperiment, then by can be one of the fields in the row annotation data.frame (i.e. one of colnames(rowData(inSCE))).
exactMatch	Boolean. Whether to only identify exact matches or to identify partial matches using grep.
removeNA	Boolean. If set to FALSE, features not found in inSCE will be given NA and the returned vector will be the same length as features. If set to TRUE, then the NA values will be removed from the returned vector. Default FALSE.
errorOnNoMatch	Boolean. If TRUE, an error will be given if no matches are found. If FALSE, an empty vector will be returned if removeNA is set to TRUE or a vector of NA if removeNA is set to FALSE. Default TRUE.
warningOnPartialMatch	Boolean. If TRUE, a warning will be given if some of the entries in features were not found in inSCE. The warning will list the features not found. Default TRUE.

Value

A vector of row indices for the matching features in inSCE.

Author(s)

Yusuke Koga, Joshua D. Campbell

See Also

'retrieveFeatureInfo' from package 'scater' and link{regex} for how to use regular expressions when exactMatch = FALSE.

Examples

```
data(scExample)
ix <- featureIndex(features = c("MT-CYB", "MT-ND2"),
                           inSCE = sce,
                           by = "feature_name")
```

`generateHTANMeta` *Generate HTAN manifest file for droplet and cell count data*

Description

Generate HTAN manifest file for droplet and cell count data

Usage

```
generateHTANMeta(
  dropletSCE = NULL,
  cellSCE = NULL,
  samplename,
  htan_biospecimen_id,
  dir,
  dataType = c("Droplet", "Cell", "Both")
)
```

Arguments

`dropletSCE`	A SingleCellExperiment object containing droplet count matrix data
`cellSCE`	A SingleCellExperiment object containing cell count matrix data
`samplename`	The sample name of the SingleCellExperiment objects
`htan_biospecimen_id`	The HTAN biospecimen id of the sample in SingleCellExperiment object
`dir`	The output directory of the SCTK QC pipeline.
`dataType`	Type of the input data. It can be one of "Droplet", "Cell" or "Both".

Value

A SingleCellExperiment object which combines all objects in sceList. The colData is merged.

`generateMeta` *Generate HTAN manifest file for droplet and cell count data*

Description

Generate HTAN manifest file for droplet and cell count data

Usage

```
generateMeta(
  dropletSCE = NULL,
  cellSCE = NULL,
  samplename,
  dir,
  HTAN = TRUE,
  dataType = c("Droplet", "Cell", "Both")
)
```

Arguments

dropletSCE	A SingleCellExperiment object containing droplet count matrix data
cellSCE	A SingleCellExperiment object containing cell count matrix data
samplename	The sample name of the SingleCellExperiment objects
dir	The output directory of the SCTK QC pipeline.
HTAN	Whether generates manifest file including HTAN specific ID (HTAN Biospecimen ID, HTAN parent file ID and HTAN patient ID). Default is TRUE.
dataType	Type of the input data. It can be one of "Droplet", "Cell" or "Both".

Value

A SingleCellExperiment object which combines all objects in sceList. The colData is merged.

---

generateSimulatedData *Generates a single simulated dataset, bootstrapping from the input counts matrix.*

Description

Generates a single simulated dataset, bootstrapping from the input counts matrix.

Usage

```
generateSimulatedData(totalReads, cells, originalData, realLabels)
```

Arguments

totalReads	Numeric. The total number of reads in the simulated dataset, to be split between all simulated cells.
cells	Numeric. The number of virtual cells to simulate.
originalData	Matrix. The original raw read count matrix. When used within the Shiny app, this will be assay(SCEsetObject, "counts").
realLabels	Factor. The condition labels for differential expression. If only two factors present, will default to t-test. If multiple factors, will default to ANOVA.

Value

A simulated counts matrix, the first row of which contains the 'true' labels for each virtual cell.

Examples

```
data("mouseBrainSubsetSCE")
res <- generateSimulatedData(
         totalReads = 1000, cells=10,
         originalData = assay(mouseBrainSubsetSCE, "counts"),
         realLabels = colData(mouseBrainSubsetSCE)[, "level1class"])
```

`getBiomarker` *Given a list of genes and a SingleCellExperiment object, return the binary or continuous expression of the genes.*

Description

Given a list of genes and a SingleCellExperiment object, return the binary or continuous expression of the genes.

Usage

```
getBiomarker(
  inSCE,
  gene,
  binary = "Binary",
  useAssay = "counts",
  featureLocation = NULL,
  featureDisplay = NULL
)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`gene`	gene list
`binary`	"Binary" for binary expression or "Continuous" for a gradient. Default: "Binary"
`useAssay`	Indicates which assay to use. The default is "counts".
`featureLocation`	Indicates which column name of rowData to query gene.
`featureDisplay`	Indicates which column name of rowData to use to display feature for visualization.

Value

getBiomarker(): A data.frame of expression values

Examples

```
data("mouseBrainSubsetSCE")
getBiomarker(mouseBrainSubsetSCE, gene="C1qa")
```

`getDEGTopTable` *Get Top Table of a DEG analysis*

Description

Users have to run `runDEAnalysis()` first, any of the wrapped functions of this generic function. Users can set further filters on the result. A `data.frame` object, with variables of `Gene`, `Log2_FC`, `Pvalue`, and `FDR`, will be returned.

Usage

```
getDEGTopTable(
  inSCE,
  useResult,
  labelBy = S4Vectors::metadata(inSCE)$featureDisplay,
  onlyPos = FALSE,
  log2fcThreshold = 0.25,
  fdrThreshold = 0.05,
  minGroup1MeanExp = NULL,
  maxGroup2MeanExp = NULL,
  minGroup1ExprPerc = NULL,
  maxGroup2ExprPerc = NULL
)
```

Arguments

`inSCE`	SingleCellExperiment inherited object, with of the singleCellTK DEG method performed in advance.
`useResult`	character. A string specifying the `analysisName` used when running a differential expression analysis function.
`labelBy`	A single character for a column of `rowData(inSCE)` as where to search for the labeling text. Leave `NULL` for `rownames`. Default `metadata(inSCE)$featureDisplay` (see `setSCTKDisplayRow`).
`onlyPos`	logical. Whether to only fetch DEG with positive log2_FC value. Default `FALSE`.
`log2fcThreshold`	numeric. Only fetch DEGs with the absolute values of log2FC larger than this value. Default `0.25`.
`fdrThreshold`	numeric. Only fetch DEGs with FDR value smaller than this value. Default `0.05`.

minGroup1MeanExp
: numeric. Only fetch DEGs with mean expression in group1 greater then this value. Default NULL.

maxGroup2MeanExp
: numeric. Only fetch DEGs with mean expression in group2 less then this value. Default NULL.

minGroup1ExprPerc
: numeric. Only fetch DEGs expressed in greater then this fraction of cells in group1. Default NULL.

maxGroup2ExprPerc
: numeric. Only fetch DEGs expressed in less then this fraction of cells in group2. Default NULL.

Value

A data.frame object of the top DEGs, with variables of Gene, Log2_FC, Pvalue, and FDR.

Examples

```
data("sceBatches")
sceBatches <- scaterlogNormCounts(sceBatches, "logcounts")
sce.w <- subsetSCECols(sceBatches, colData = "batch == 'w'")
sce.w <- runWilcox(sce.w, class = "cell_type",
                   classGroup1 = "alpha", classGroup2 = "beta",
                   groupName1 = "w.alpha", groupName2 = "w.beta",
                   analysisName = "w.aVSb")
getDEGTopTable(sce.w, "w.aVSb")
```

---

getDiffAbundanceResults

Get/Set diffAbundanceFET result table

Description

Get/Set diffAbundanceFET result table

Usage

```
getDiffAbundanceResults(x, analysisName)

## S4 method for signature 'SingleCellExperiment'
getDiffAbundanceResults(x, analysisName)

getDiffAbundanceResults(x, analysisName) <- value

## S4 replacement method for signature 'SingleCellExperiment'
getDiffAbundanceResults(x, analysisName) <- value
```

Arguments

x	A SingleCellExperiment object.
analysisName	A single character string specifying an analysis performed with diffAbundanceFET
value	The output table of diffAbundanceFET

Value

The differential abundance table for getter method, or update the SCE object with new result for setter method.

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- diffAbundanceFET(inSCE = mouseBrainSubsetSCE,
                                        cluster = "tissue",
                                        variable = "level1class",
                                        case = "oligodendrocytes",
                                        control = "microglia",
                                        analysisName = "diffAbund")
result <- getDiffAbundanceResults(mouseBrainSubsetSCE, "diffAbund")
```

---

getEnrichRResult<- *Get or Set EnrichR Result*

Description

Get or Set EnrichR Result

Usage

```
getEnrichRResult(inSCE, analysisName) <- value

getEnrichRResult(inSCE, analysisName)

## S4 method for signature 'SingleCellExperiment'
getEnrichRResult(inSCE, analysisName)

## S4 replacement method for signature 'SingleCellExperiment'
getEnrichRResult(inSCE, analysisName) <- value
```

Arguments

inSCE	A SingleCellExperiment object.
analysisName	A string that identifies each specific analysis
value	The EnrichR result table

Value

For getter method, a data.frame of the EnrichR result; For setter method, `inSCE` with EnrichR results updated.

See Also

`runEnrichR`

Examples

```
data("mouseBrainSubsetSCE")
if (Biobase::testBioCConnection()) {
  mouseBrainSubsetSCE <- runEnrichR(mouseBrainSubsetSCE, features = "Cmtm5",
                                    db = "GO_Cellular_Component_2017",
                                    analysisName = "analysis1")
  result <- getEnrichRResult(mouseBrainSubsetSCE, "analysis1")
}
```

---

getFindMarkerTopTable *Fetch the table of top markers that pass the filtering*

Description

Fetch the table of top markers that pass the filtering

Usage

```
getFindMarkerTopTable(
  inSCE,
  log2fcThreshold = 0,
  fdrThreshold = 0.05,
  minClustExprPerc = 0.5,
  maxCtrlExprPerc = 0.5,
  minMeanExpr = 0,
  topN = 1
)

findMarkerTopTable(
  inSCE,
  log2fcThreshold = 1,
  fdrThreshold = 0.05,
  minClustExprPerc = 0.7,
  maxCtrlExprPerc = 0.4,
  minMeanExpr = 1,
  topN = 10
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
log2fcThreshold	Only use DEGs with the absolute values of log2FC larger than this value. Default 1
fdrThreshold	Only use DEGs with FDR value smaller than this value. Default 0.05
minClustExprPerc	A numeric scalar. The minimum cutoff of the percentage of cells in the cluster of interests that expressed the marker gene. Default 0.7.
maxCtrlExprPerc	A numeric scalar. The maximum cutoff of the percentage of cells out of the cluster (control group) that expressed the marker gene. Default 0.4.
minMeanExpr	A numeric scalar. The minimum cutoff of the mean expression value of the marker in the cluster of interests. Default 1.
topN	An integer. Only to fetch this number of top markers for each cluster in maximum, in terms of log2FC value. Use NULL to cancel the top N subscription. Default 10.

Details

Users have to run runFindMarker prior to using this function to extract a top marker table.

Value

An organized data.frame object, with the top marker gene information.

See Also

runFindMarker, plotFindMarkerHeatmap

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runFindMarker(mouseBrainSubsetSCE,
                                     useAssay = "logcounts",
                                     cluster = "level1class")
getFindMarkerTopTable(mouseBrainSubsetSCE)
```

---

getGenesetNamesFromCollection

List geneset names from geneSetCollection

Description

List geneset names from geneSetCollection

Usage

```
getGenesetNamesFromCollection(inSCE, geneSetCollectionName)
```

Arguments

inSCE	Input SingleCellExperiment object.
geneSetCollectionName	The name of an imported geneSetCollection.

Value

A character vector of available genesets from the collection.

---

getMSigDBTable *Shows MSigDB categories*

Description

Returns a data.frame that shows MSigDB categories and subcategories as well as descriptions for each. The entries in the ID column in this table can be used as input for importGeneSetsFromMSigDB.

Usage

```
getMSigDBTable()
```

Value

data.frame, containing MSigDB categories

Author(s)

Joshua D. Campbell

See Also

importGeneSetsFromMSigDB for importing MSigDB gene sets.

Examples

```
getMSigDBTable()
```

getPathwayResultNames *List pathway analysis result names*

Description

List pathway analysis result names

Usage

```
getPathwayResultNames(inSCE, stopIfNone = FALSE, verbose = FALSE)
```

Arguments

inSCE	Input SingleCellExperiment object.
stopIfNone	Whether to stop and raise an error if no results found. If FALSE, will return an empty character vector.
verbose	Show warning if no result found. Default FALSE

Details

Pathway analysis results will be stored as matrices in reducedDims slot of inSCE. This function lists the result names stored in metadata slot when analysis is performed.

Value

A character vector of valid pathway analysis result names.

Examples

```
data(scExample)
getPathwayResultNames(sce)
```

getSampleSummaryStatsTable

Stores and returns table of SCTK QC outputs to metadata.

Description

Stores and returns table of QC metrics generated from QC algorithms within the metadata slot of the SingleCellExperiment object.

Usage

```
getSampleSummaryStatsTable(inSCE, statsName, ...)

setSampleSummaryStatsTable(inSCE, statsName, ...) <- value

## S4 method for signature 'SingleCellExperiment'
getSampleSummaryStatsTable(inSCE, statsName, ...)

## S4 replacement method for signature 'SingleCellExperiment'
setSampleSummaryStatsTable(inSCE, statsName, ...) <- value
```

Arguments

inSCE	Input SingleCellExperiment object with saved assay data and/or colData data. Required.
statsName	A character value indicating the slot that stores the stats table within the metadata of the SingleCellExperiment object. Required.
...	Other arguments passed to the function.
value	The summary table for QC statistics generated from SingleCellTK to be added to the SCE object.

Value

For getSampleSummaryStatsTable, A matrix/array object. Contains a summary table for QC statistics generated from SingleCellTK. For setSampleSummaryStatsTable<-, A SingleCellExperiment object where the summary table is updated in the metadata slot.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- sampleSummaryStats(sce, simple = TRUE, statsName = "qc_table")
getSampleSummaryStatsTable(sce, statsName = "qc_table")
```

---

getSceParams *Extract QC parameters from the SingleCellExperiment object*

Description

Extract QC parameters from the SingleCellExperiment object

Usage

```
getSceParams(
  inSCE,
  skip = c("runScrublet", "runDecontX", "runBarcodeRanksMetaOutput", "genesets",
    "runSoupX"),
  ignore = c("algorithms", "estimates", "contamination", "z", "sample", "rank",
    "BPPARAM", "batch", "geneSetCollection", "barcodeArgs"),
  directory = "./",
  samplename = "",
  writeYAML = TRUE
)
```

Arguments

inSCE	A SingleCellExperiment object.
skip	Skip extracting the parameters of the provided QC functions.
ignore	Skip extracting the content within QC functions.
directory	The output directory of the SCTK_runQC.R pipeline.
samplename	The sample name of the SingleCellExperiment objects.
writeYAML	Whether output yaml file to store parameters. Default if TRUE. If FALSE, return character object.

Value

If writeYAML TRUE, a yaml object will be generated. If FALSE, character object.

---

getSeuratVariableFeatures

Get variable feature names after running runSeuratFindHVG function

---

Description

Get variable feature names after running runSeuratFindHVG function

Usage

```
getSeuratVariableFeatures(inSCE)
```

Arguments

inSCE	Input SingleCellExperiment object.

Value

A list of variable feature names.

getSoupX<- *Get or Set SoupX Result*

Description

S4 method for getting and setting SoupX results that cannot be appended to either `rowData(inSCE)` or `colData(inSCE)`.

S4 method for getting and setting SoupX results that cannot be appended to either `rowData(inSCE)` or `colData(inSCE)`.

Usage

```
getSoupX(inSCE, sampleID, background = FALSE) <- value

getSoupX(inSCE, sampleID = NULL, background = FALSE)

## S4 method for signature 'SingleCellExperiment'
getSoupX(inSCE, sampleID = NULL, background = FALSE)

## S4 replacement method for signature 'SingleCellExperiment'
getSoupX(inSCE, sampleID, background = FALSE) <- value
```

Arguments

`inSCE`	A SingleCellExperiment object. For getter method, `runSoupX` must have been already applied.
`sampleID`	Character vector. For getter method, the samples that should be included in the returned list. Leave this `NULL` for all samples. Default `NULL`. For setter method, only one sample allowed.
`background`	Logical. Whether `background` was applied when running `runSoupX`. Default `FALSE`.
`value`	Dedicated list object of SoupX results.

Value

For getter method, a list with SoupX results for specified samples. For setter method, `inSCE` with SoupX results updated.

For getter method, a list with SoupX results for specified samples. For setter method, `inSCE` with SoupX results updated.

See Also

runSoupX, plotSoupXResults

Examples

```
## Not run:
sce <- importExampleData("pbmc3k")
sce <- runSoupX(sce, sample = "sample")
soupXResults <- getSoupX(sce)

## End(Not run)
```

`getTopHVG` *Get or set top HVG after calculation*

Description

Extracts or select the top variable genes from an input SingleCellExperiment object. Note that the variability metrics must be computed using the `runFeatureSelection` method before extracting the feature names of the top variable features. `getTopHVG` only returns a character vector of the HVG selection, while with `setTopHVG`, a logical vector of the selection will be saved in the `rowData`, and optionally, a subset object for the HVGs can be stored in the `altExps` slot at the same time.

Usage

```
getTopHVG(
  inSCE,
 method = c("vst", "dispersion", "mean.var.plot", "modelGeneVar", "seurat", "seurat_v3",
    "cell_ranger"),
  hvgNumber = 2000,
  useFeatureSubset = "hvf",
  featureDisplay = metadata(inSCE)$featureDisplay
)

setTopHVG(
  inSCE,
 method = c("vst", "dispersion", "mean.var.plot", "modelGeneVar", "seurat", "seurat_v3",
    "cell_ranger"),
  hvgNumber = 2000,
  featureSubsetName = "hvg2000",
  genes = NULL,
  genesBy = NULL,
  altExp = FALSE
)
```

Arguments

`inSCE`	Input SingleCellExperiment object
`method`	Specify which method to use for variable gene extraction from Seurat `"vst"`, `"mean.var.plot"`, `"dispersion"` or Scran `"modelGeneVar"` or Scanpy `"seurat"`, `"cell_ranger"`, `"seurat_v3"`. Default `"vst"`

hvgNumber Specify the number of top variable genes to extract.

useFeatureSubset
Get the feature names in the HVG list set by setTopHVG. method and hvgNumber will not be used if not this is not NULL. Default "hvf".

featureDisplay A character string for the rowData variable name to indicate what type of feature ID should be displayed. If set by setSCTKDisplayRow, will by default use it. If NULL, will use rownames(inSCE).

featureSubsetName
A character string for the rowData variable name to store a logical index of selected features. Default "hvg2000".

genes A customized character vector of gene list to be set as a rowData variable. Will ignore method and hvgNumber if set. Default NULL.

genesBy If setting customized genes, where should it be found in rowData? Leave NULL for matching rownames. Default NULL.

altExp TRUE for also creating a subset inSCE object with the selected HVGs and store this subset in the altExps slot, named by hvgListName. Default FALSE.

Value

getTopHVG A character vector of the top hvgNumber variable feature names

setTopHVG The input inSCE object with the logical vector of HVG selection updated in rowData, and related parameter updated in metadata. If altExp is TRUE, an altExp is also added

Author(s)

Irzam Sarfraz, Yichen Wang

See Also

runFeatureSelection, runSeuratFindHVG, runModelGeneVar, plotTopHVG

Examples

```
data("scExample", package = "singleCellTK")

# Create a "highy variable feature" subset using Seurat's vst method:
sce <- runSeuratFindHVG(sce,  method = "vst", hvgNumber = 2000,
       createFeatureSubset = "hvf")

# Get the list of genes for a feature subset:
hvgs <- getTopHVG(sce, useFeatureSubset = "hvf")

# Create a new feature subset on the fly without rerunning the algorithm:
sce <- setTopHVG(sce, method = "vst", hvgNumber = 100,
                 featureSubsetName = "hvf100")
hvgs <- getTopHVG(sce, useFeatureSubset = "hvf100")

# Get a list of variable features without creating a new feature subset:
```

```
hvgs <- getTopHVG(sce, useFeatureSubset = NULL,
                  method = "vst", hvgNumber = 10)
```

`getTSCANResults` *getTSCANResults accessor function*

Description

SCTK allows user to access all TSCAN related results with `"getTSCANResults"`. See details.

Usage

```
getTSCANResults(x, analysisName = NULL, pathName = NULL)

## S4 method for signature 'SingleCellExperiment'
getTSCANResults(x, analysisName = NULL, pathName = NULL)

getTSCANResults(x, analysisName, pathName = NULL) <- value

## S4 replacement method for signature 'SingleCellExperiment'
getTSCANResults(x, analysisName, pathName = NULL) <- value

listTSCANResults(x)

## S4 method for signature 'SingleCellExperiment'
listTSCANResults(x)

listTSCANTerminalNodes(x)

## S4 method for signature 'SingleCellExperiment'
listTSCANTerminalNodes(x)
```

Arguments

`x`	Input SingleCellExperiment object.
`analysisName`	Algorithm name implemented, should be one of `"Pseudotime"`, `"DEG"`, or `"ClusterDEAnalysis"`.
`pathName`	Sub folder name within the `analysisName`. See details.
`value`	Value to be stored within the `pathName` or `analysisName`

Details

When `analysisName = "Pseudotime"`, returns the list result from `runTSCAN`, including the MST structure.

When `analysisName = "DEG"`, returns the list result from `runTSCANDEG`, including `DataFrames` containing genes that increase/decrease along each the pseudotime paths. `pathName` indicates the path index, the available options of which can be listed by `listTSCANTerminalNodes`.

When analysisName = "ClusterDEAnalysis", returns the list result from runTSCANClusterDEAnalysis. Here pathName needs to match with the useCluster argument when running the algorithm.

Value

Get or set TSCAN results

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
results <- getTSCANResults(mouseBrainSubsetSCE, "Pseudotime")
```

---

importAlevin *Construct SCE object from Salmon-Alevin output*

Description

Construct SCE object from Salmon-Alevin output

Usage

```
importAlevin(
  alevinDir = NULL,
  sampleName = "sample",
  delayedArray = FALSE,
  class = c("Matrix", "matrix"),
  rowNamesDedup = TRUE
)
```

Arguments

alevinDir	Character. The output directory of salmon-Alevin pipeline. It should contain subfolder named 'alevin', which contains the count data which is stored in 'quants_mat.gz'. Default NULL.
sampleName	Character. A user-defined sample name for the sample to be imported. The 'sampleName' will be appended to the begining of cell barcodes. Default is 'sample'.
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
rowNamesDedup	Boolean. Whether to deduplicate rownames. Default TRUE.

Value

A SingleCellExperiment object containing the count matrix, the feature annotations, and the cell annotation (which includes QC metrics stored in 'featureDump.txt').

`importAnnData` *Create a SingleCellExperiment Object from Python AnnData .h5ad files*

Description

This function reads in one or more Python AnnData files in the .h5ad format and returns a single SingleCellExperiment object containing all the AnnData samples by concatenating their counts matrices and related information slots.

Usage

```
importAnnData(
  sampleDirs = NULL,
  sampleNames = NULL,
  delayedArray = FALSE,
  class = c("Matrix", "matrix"),
  rowNamesDedup = TRUE
)
```

Arguments

`sampleDirs`	Folder containing the .h5ad file. Can be one of - • Default `current working directory`. • Full path to the directory containing the .h5ad file. E.g `sampleDirs = '/path/to/sample'` • A vector of folder paths for the samples to import. E.g. `sampleDirs = c('/path/to/sample1', '/path/to/sample2','/path/to/sample3')` importAnnData will return a single SCE object containing all the samples with the sample name appended to each colname in colData
`sampleNames`	The prefix/name of the .h5ad file without the .h5ad extension e.g. if 'sample.h5ad' is the filename, pass `sampleNames = 'sample'`. Can be one of - • Default `sample`. • A vector of samples to import. Length of vector must be equal to length of sampleDirs vector E.g. `sampleDirs = c('sample1', 'sample2','sample3')` importAnnData will return a single SCE object containing all the samples with the sample name appended to each colname in colData
`delayedArray`	Boolean. Whether to read the expression matrix as DelayedArray object. Default `FALSE`.
`class`	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default `"Matrix"`.
`rowNamesDedup`	Boolean. Whether to deduplicate rownames. Default `TRUE`.

Details

importAnnData converts scRNA-seq data in the AnnData format to the SingleCellExperiment object. The .X slot in AnnData is transposed to the features x cells format and becomes the 'counts' matrix in the assay slot. The .vars AnnData slot becomes the SCE rowData and the .obs AnnData slot becomes the SCE colData. Multidimensional data in the .obsm AnnData slot is ported over to the SCE reducedDims slot. Additionally, unstructured data in the .uns AnnData slot is available through the SCE metadata slot. There are 2 currently known minor issues - Anndata python module depends on another python module h5pyto read hd5 format files. If there are errors reading the .h5ad files, such as "ValueError: invalid shape in fixed-type tuple." the user will need to do downgrade h5py by running pip3 install --user h5py==2.9.0 Additionally there might be errors in converting some python objects in the unstructured data slots. There are no known R solutions at present. Refer https://github.com/rstudio/reticulate/issues/209

Value

A SingleCellExperiment object.

Examples

```
file.path <- system.file("extdata/annData_pbmc_3k", package = "singleCellTK")
## Not run:
sce <- importAnnData(sampleDirs = file.path,
                     sampleNames = 'pbmc3k_20by20')

## End(Not run)
```

---

importBUStools *Construct SCE object from BUStools output*

Description

Read the barcodes, features (genes), and matrix from BUStools output. Import them as one SingleCellExperiment object. Note the cells in the output files for BUStools 0.39.4 are not filtered.

Usage

```
importBUStools(
  BUStoolsDirs,
  samples,
  matrixFileNames = "genes.mtx",
  featuresFileNames = "genes.genes.txt",
  barcodesFileNames = "genes.barcodes.txt",
  gzipped = "auto",
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)
```

Arguments

BUStoolsDirs	A vector of paths to BUStools output files. Each sample should have its own path. For example: ./genecount. Must have the same length as samples.
samples	A vector of user-defined sample names for the samples to be imported. Must have the same length as BUStoolsDirs.
matrixFileNames	Filenames for the Market Exchange Format (MEX) sparse matrix files (.mtx files). Must have length 1 or the same length as samples.
featuresFileNames	Filenames for the feature annotation files. Must have length 1 or the same length as samples.
barcodesFileNames	Filenames for the cell barcode list file. Must have length 1 or the same length as samples.
gzipped	Boolean. TRUE if the BUStools output files (barcodes.txt, genes.txt, and genes.mtx) were gzip compressed. FALSE otherwise. This is FALSE in BUStools 0.39.4. Default "auto" which automatically detects if the files are gzip compressed. Must have length 1 or the same length as samples.
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray-class object or not. Default FALSE.
rowNamesDedup	Boolean. Whether to deduplicate rownames. Default TRUE.

Value

A SingleCellExperiment object containing the count matrix, the gene annotation, and the cell annotation.

Examples

```
# Example #1
# FASTQ files were downloaded from
# https://support.10xgenomics.com/single-cell-gene-expression/datasets/3.0.0
# /pbmc_1k_v3
# They were concatenated as follows:
# cat pbmc_1k_v3_S1_L001_R1_001.fastq.gz pbmc_1k_v3_S1_L002_R1_001.fastq.gz >
# pbmc_1k_v3_R1.fastq.gz
# cat pbmc_1k_v3_S1_L001_R2_001.fastq.gz pbmc_1k_v3_S1_L002_R2_001.fastq.gz >
# pbmc_1k_v3_R2.fastq.gz
# The following BUStools command generates the gene, cell, and
# matrix files

# bustools correct -w ./3M-february-2018.txt -p output.bus | \
#   bustools sort -T tmp/ -t 4 -p - | \
#   bustools count -o genecount/genes \
#     -g ./transcripts_to_genes.txt \
```

```
#     -e matrix.ec \
#     -t transcripts.txt \
#     --genecounts -

# The top 20 genes and the first 20 cells are included in this example.
sce <- importBUStools(
  BUStoolsDirs = system.file("extdata/BUStools_PBMC_1k_v3_20x20/genecount/",
    package = "singleCellTK"),
  samples = "PBMC_1k_v3_20x20")
```

`importCellRanger` *Construct SCE object from Cell Ranger output*

Description

Read the filtered barcodes, features, and matrices for all samples from (preferably a single run of) Cell Ranger output. Import and combine them as one big SingleCellExperiment object.

Usage

```
importCellRanger(
  cellRangerDirs = NULL,
  sampleDirs = NULL,
  sampleNames = NULL,
  cellRangerOuts = NULL,
  dataType = c("filtered", "raw"),
  matrixFileNames = "matrix.mtx.gz",
  featuresFileNames = "features.tsv.gz",
  barcodesFileNames = "barcodes.tsv.gz",
  gzipped = "auto",
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)

importCellRangerV2(
  cellRangerDirs = NULL,
  sampleDirs = NULL,
  sampleNames = NULL,
  dataTypeV2 = c("filtered", "raw"),
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  reference = NULL,
  cellRangerOutsV2 = NULL,
  rowNamesDedup = TRUE
)

importCellRangerV3(
```

```
  cellRangerDirs = NULL,
  sampleDirs = NULL,
  sampleNames = NULL,
  dataType = c("filtered", "raw"),
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)
```

Arguments

cellRangerDirs	The root directories where Cell Ranger was run. These folders should contain sample specific folders. Default NULL, meaning the paths for each sample will be specified in *samples* argument.
sampleDirs	Default NULL. Can be one of • NULL. All samples within cellRangerDirs will be imported. The order of samples will be first determined by the order of cellRangerDirs and then by list.dirs. This is only for the case where cellRangerDirs is specified. • A list of vectors containing the folder names for samples to import. Each vector in the list corresponds to samples from one of cellRangerDirs. These names are the same as the folder names under cellRangerDirs. This is only for the case where cellRangerDirs is specified. • A vector of folder paths for the samples to import. This is only for the case where cellRangerDirs is NULL. The cells in the final SCE object will be ordered in the same order of sampleDirs.
sampleNames	A vector of user-defined sample names for the samples to be imported. Must have the same length as length(unlist(sampleDirs)) if sampleDirs is not NULL. Otherwise, make sure the length and order match the output of unlist(lapply(cellRangerDirs, list.dirs, recursive = FALSE)). Default NULL, in which case the folder names will be used as sample names.
cellRangerOuts	Character vector. The intermediate paths to filtered or raw cell barcode, feature, and matrix files for each sample. **Supercedes** dayaType. If NULL, dataType will be used to determine Cell Ranger output directory. If not NULL, dataType will be ingored and cellRangerOuts specifies the paths. Must have length 1 or the same length as length(unlist(sampleDirs)) if sampleDirs is not NULL. Otherwise, make sure the length and order match the output of unlist(lapply(cellRangerDirs, list.dirs, recursive = FALSE)). Reference genome names might need to be appended for CellRanger version below 3.0.0 if reads were mapped to multiple genomes when running Cell Ranger pipeline. Probable options include "outs/filtered_feature_bc_matrix/", "outs/raw_feature_bc_matrix/", "outs/filtered_gene_bc_matrix/", "outs/raw_gene_bc_matrix/".
dataType	Character. The type of data to import. Can be one of "filtered" (which is equivalent to cellRangerOuts = "outs/filtered_feature_bc_matrix/" or cellRangerOuts = "outs/filtered_gene_bc_matrix/") or "raw" (which is equivalent to cellRangerOuts = "outs/raw_feature_bc_matrix/" or cellRangerOuts = "outs/raw_gene_bc_matrix/"). Default "filtered" which imports the counts for filtered cell barcodes only.

matrixFileNames
: Character vector. Filenames for the Market Exchange Format (MEX) sparse matrix files (matrix.mtx or matrix.mtx.gz files). Must have length 1 or the same length as `length(unlist(sampleDirs))` if `sampleDirs` is not `NULL`. Otherwise, make sure the length and order match the output of `unlist(lapply(cellRangerDirs, list.dirs, recursive = FALSE))`.

featuresFileNames
: Character vector. Filenames for the feature annotation files. They are usually named *features.tsv.gz* or *genes.tsv*. Must have length 1 or the same length as `length(unlist(sampleDirs))` if `sampleDirs` is not `NULL`. Otherwise, make sure the length and order match the output of `unlist(lapply(cellRangerDirs, list.dirs, recursive = FALSE))`.

barcodesFileNames
: Character vector. Filename for the cell barcode list files. They are usually named *barcodes.tsv.gz* or *barcodes.tsv*. Must have length 1 or the same length as `length(unlist(sampleDirs))` if `sampleDirs` is not `NULL`. Otherwise, make sure the length and order match the output of `unlist(lapply(cellRangerDirs, list.dirs, recursive = FALSE))`.

gzipped
: `TRUE` if the Cell Ranger output files (barcodes.tsv, features.tsv, and matrix.mtx) were gzip compressed. `FALSE` otherwise. This is true after Cell Ranger 3.0.0 update. Default `"auto"` which automatically detects if the files are gzip compressed. If not `"auto"`, `gzipped` must have length 1 or the same length as `length(unlist(sampleDirs))` if `sampleDirs` is not `NULL`. Otherwise, make sure the length and order match the output of `unlist(lapply(cellRangerDirs, list.dirs, recursive = FALSE))`.

class
: Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default `"Matrix"`.

delayedArray
: Boolean. Whether to read the expression matrix as DelayedArray object or not. Default `FALSE`.

rowNamesDedup
: Boolean. Whether to deduplicate rownames. Default `TRUE`.

dataTypeV2
: Character. The type of output to import for Cellranger version below 3.0.0. Whether to import the filtered or the raw data. Can be one of 'filtered' or 'raw'. Default 'filtered'. When `cellRangerOuts` is specified, `dataTypeV2` and `reference` will be ignored.

reference
: Character vector. The reference genome names. Default `NULL`. If not `NULL`, it must gave the length and order as `length(unlist(sampleDirs))` if `sampleDirs` is not `NULL`. Otherwise, make sure the length and order match the output of `unlist(lapply(cellRangerDirs, list.dirs, recursive = FALSE))`. Only needed for Cellranger version below 3.0.0.

cellRangerOutsV2
: Character vector. The intermediate paths to filtered or raw cell barcode, feature, and matrix files for each sample for Cellranger version below 3.0.0. If `NULL`, `reference` and `dataTypeV2` will be used to determine Cell Ranger output directory. If it has length 1, it assumes that all samples use the same genome reference and the function will load only filtered or raw data.

Details

`importCellRangerV2` imports output from Cell Ranger V2. `importCellRangerV2Sample` imports output from one sample from Cell Ranger V2. `importCellRangerV3` imports output from Cell Ranger V3. `importCellRangerV3` imports output from one sample from Cell Ranger V3. Some implicit assumptions which match the output structure of Cell Ranger V2 & V3 are made in these 4 functions including `cellRangerOuts`, `matrixFileName`, `featuresFileName`, `barcodesFileName`, and `gzipped`. Alternatively, user can call `importCellRanger` to explicitly specify these arguments.

Value

A `SingleCellExperiment` object containing the combined count matrix, the feature annotations, and the cell annotation.

Examples

```
# Example #1
# The following filtered feature, cell, and matrix files were downloaded from
# https://support.10xgenomics.com/single-cell-gene-expression/datasets/
# 3.0.0/hgmm_1k_v3
# The top 10 hg19 & mm10 genes are included in this example.
# Only the first 20 cells are included.
sce <- importCellRanger(
    cellRangerDirs = system.file("extdata/", package = "singleCellTK"),
    sampleDirs = "hgmm_1k_v3_20x20",
    sampleNames = "hgmm1kv3",
    dataType = "filtered")
# The following filtered feature, cell, and matrix files were downloaded from
# https://support.10xgenomics.com/single-cell-gene-expression/datasets/
# 2.1.0/pbmc4k
# Top 20 genes are kept. 20 cell barcodes are extracted.
sce <- importCellRangerV2(
    cellRangerDirs = system.file("extdata/", package = "singleCellTK"),
    sampleDirs = "pbmc_4k_v2_20x20",
    sampleNames = "pbmc4k_20",
    reference = 'GRCh38',
    dataTypeV2 = "filtered")
sce <- importCellRangerV3(
    cellRangerDirs = system.file("extdata/", package = "singleCellTK"),
    sampleDirs = "hgmm_1k_v3_20x20",
    sampleNames = "hgmm1kv3",
    dataType = "filtered")
```

---

importCellRangerV2Sample

Construct SCE object from Cell Ranger V2 output for a single sample

---

Description

Read the filtered barcodes, features, and matrices for all samples from Cell Ranger V2 output. Files are assumed to be named "matrix.mtx", "genes.tsv", and "barcodes.tsv".

Usage

```
importCellRangerV2Sample(
  dataDir = NULL,
  sampleName = NULL,
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)
```

Arguments

dataDir	A path to the directory containing the data files. Default "./".
sampleName	A User-defined sample name. This will be prepended to all cell barcode IDs. Default "sample".
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.
rowNamesDedup	Boolean. Whether to deduplicate rownames. Default TRUE.

Value

A SingleCellExperiment object containing the count matrix, the feature annotations, and the cell annotation for the sample.

Examples

```
sce <- importCellRangerV2Sample(
    dataDir = system.file("extdata/pbmc_4k_v2_20x20/outs/",
        "filtered_gene_bc_matrices/GRCh38", package = "singleCellTK"),
    sampleName = "pbmc4k_20")
```

---

importCellRangerV3Sample

Construct SCE object from Cell Ranger V3 output for a single sample

---

Description

Read the filtered barcodes, features, and matrices for all samples from Cell Ranger V3 output. Files are assumed to be named "matrix.mtx.gz", "features.tsv.gz", and "barcodes.tsv.gz".

Usage

```
importCellRangerV3Sample(
  dataDir = "./",
  sampleName = "sample",
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)
```

Arguments

dataDir	A path to the directory containing the data files. Default "./".
sampleName	A User-defined sample name. This will be prepended to all cell barcode IDs. Default "sample".
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.
rowNamesDedup	Boolean. Whether to deduplicate rownames. Default TRUE.

Value

A SingleCellExperiment object containing the count matrix, the feature annotations, and the cell annotation for the sample.

Examples

```
sce <- importCellRangerV3Sample(
    dataDir = system.file("extdata/hgmm_1k_v3_20x20/outs/",
        "filtered_feature_bc_matrix", package = "singleCellTK"),
    sampleName = "hgmm1kv3")
```

---

importDropEst *Create a SingleCellExperiment Object from DropEst output*

Description

imports the RDS file created by DropEst (https://github.com/hms-dbmi/dropEst) and create a SingleCellExperiment object from either the raw or filtered counts matrix. Additionally parse through the RDS to obtain appropriate feature annotations as SCE coldata, in addition to any metadata.

Usage

```
importDropEst(
  sampleDirs = NULL,
  dataType = c("filtered", "raw"),
  rdsFileName = "cell.counts",
  sampleNames = NULL,
  delayedArray = FALSE,
  class = c("Matrix", "matrix"),
  rowNamesDedup = TRUE
)
```

Arguments

sampleDirs	A path to the directory containing the data files. Default "./".
dataType	can be "filtered" or "raw". Default "filtered".
rdsFileName	File name prefix of the DropEst RDS output. default is "cell.counts"
sampleNames	A User-defined sample name. This will be prepended to all cell barcode IDs. Default "sample".
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
rowNamesDedup	Boolean. Whether to deduplicate rownames. Default TRUE.

Details

importDropEst expects either raw counts matrix stored as "cm_raw" or filtered counts matrix stored as "cm" in the DropEst rds output. ColData is obtained from the DropEst corresponding to "mean_reads_per_umi","aligned_reads_per_cell", "aligned_umis_per_cell","requested_umis_per_cb","requested_reads_per
If using filtered counts matrix, the colData dataframe is subset to contain features from the filtered counts matrix alone. If any annotations of ("saturation_info","merge_targets","reads_per_umi_per_cell") are found in the DropEst rds, they will be added to the SCE metadata field

Value

A SingleCellExperiment object containing the count matrix, the feature annotations from DropEst as ColData, and any metadata from DropEst

Examples

```
# Example results were generated as per instructions from the developers of dropEst described in
# https://github.com/hms-dbmi/dropEst/blob/master/examples/EXAMPLES.md
sce <- importDropEst(sampleDirs = system.file("extdata/dropEst_scg71", package = "singleCellTK"),
                     sampleNames = 'scg71')
```

`importExampleData` *Retrieve example datasets*

Description

Retrieves published example datasets stored in SingleCellExperiment using the scRNAseq and TENxPBMCData packages. See 'Details' for a list of available datasets.

Usage

```
importExampleData(
  dataset,
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)
```

Arguments

`dataset`	Character. Name of the dataset to retrieve.
`class`	Character. The class of the expression matrix stored in the SCE object. Can be one of `"Matrix"` or `"matrix"`. `"Matrix"` will store the data as a sparse matrix from package Matrix while `"matrix"` will store the data in a standard matrix. Default `"Matrix"`.
`delayedArray`	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default `FALSE`.
`rowNamesDedup`	Boolean. Whether to deduplicate rownames. Default `TRUE`.

Details

See the list below for the available datasets and their descriptions.

"fluidigm_pollen" Retrieved with `ReprocessedFluidigmData`. Returns a dataset of 65 human neural cells from Pollen et al. (2014), each sequenced at high and low coverage (SRA accession SRP041736).

"allen_tasic" Retrieved with `ReprocessedAllenData`. Returns a dataset of 379 mouse brain cells from Tasic et al. (2016).

"NestorowaHSCData" Retrieved with `NestorowaHSCData`. Returns a dataset of 1920 mouse haematopoietic stem cells from Nestorowa et al. 2015

"pbmc3k" Retrieved with `TENxPBMCData`. 2,700 peripheral blood mononuclear cells (PBMCs) from 10X Genomics.

"pbmc4k" Retrieved with `TENxPBMCData`. 4,340 peripheral blood mononuclear cells (PBMCs) from 10X Genomics.

"pbmc6k" Retrieved with `TENxPBMCData`. 5,419 peripheral blood mononuclear cells (PBMCs) from 10X Genomics.

"pbmc8k" Retrieved with TENxPBMCData. 8,381 peripheral blood mononuclear cells (PBMCs) from 10X Genomics.

"pbmc33k" Retrieved with TENxPBMCData. 33,148 peripheral blood mononuclear cells (PBMCs) from 10X Genomics.

"pbmc68k" Retrieved with TENxPBMCData. 68,579 peripheral blood mononuclear cells (PBMCs) from 10X Genomics.

Value

The specified SingleCellExperiment object.

Author(s)

Joshua D. Campbell, David Jenkins

Examples

```
sce <- importExampleData("pbmc3k")
```

---

importFromFiles *Create a SingleCellExperiment object from files*

Description

Create a SingleCellExperiment object from files

Usage

```
importFromFiles(
  assayFile,
  annotFile = NULL,
  featureFile = NULL,
  assayName = "counts",
  inputDataFrames = FALSE,
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  annotFileHeader = FALSE,
  annotFileRowName = 1,
  annotFileSep = "\t",
  featureHeader = FALSE,
  featureRowName = 1,
  featureSep = "\t",
  gzipped = "auto",
  rowNamesDedup = TRUE
)
```

Arguments

assayFile	The path to a file in .mtx, .txt, .csv, .tab, or .tsv format.
annotFile	The path to a text file that contains columns of annotation information for each cell in the assayFile. This file should have the same number of rows as there are columns in the assayFile. If multiple samples are represented in the dataset, this should be denoted by a column called 'sample' within the annotFile.
featureFile	The path to a text file that contains columns of annotation information for each gene in the count matrix. This file should have the same genes in the same order as assayFile. This is optional.
assayName	The name of the assay that you are uploading. The default is "counts".
inputDataFrames	If TRUE, assayFile, annotFile and featureFile should be data.frames object (or its inheritance) instead of file paths. The default is FALSE.
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.
annotFileHeader	Whether there's a header (colnames) in the cell annotation file. Default is FALSE.
annotFileRowName	Which column is used as the rownames for the cell annotation file. This should match to the colnames of the assayFile. Default is 1 (first column).
annotFileSep	Separater used for the cell annotation file. Default is "\t".
featureHeader	Whether there's a header (colnames) in the feature annotation file. Default is FALSE.
featureRowName	Which column is used as the rownames for the feature annotation file. This should match to the rownames of the assayFile. Default is 1. (first column).
featureSep	Separater used for the feature annotation file. Default is "\t".
gzipped	Whether the input file is gzipped. Default is "auto" and it will automatically detect whether the file is gzipped. Other options are TRUE or FALSE.
rowNamesDedup	Boolean. Whether to deduplicate rownames. Default TRUE.

Details

Creates a SingleCellExperiment object from a counts file in various formats, and files of cell and feature annotation.

Value

a SingleCellExperiment object

`importGeneSetsFromCollection` *Imports gene sets from a GeneSetCollection object*

Description

Converts a list of gene sets stored in a GeneSetCollection object and stores it in the metadata of the SingleCellExperiment object. These gene sets can be used in downstream quality control and analysis functions in singleCellTK.

Usage

```
importGeneSetsFromCollection(
  inSCE,
  geneSetCollection,
  collectionName = "GeneSetCollection",
  by = "rownames",
  noMatchError = TRUE
)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`geneSetCollection`	A GeneSetCollection object. See GeneSetCollection for more details.
`collectionName`	Character. Name of collection to add gene sets to. If this collection already exists in `inSCE`, then these gene sets will be added to that collection. Any gene sets within the collection with the same name will be overwritten. Default `GeneSetCollection`.
`by`	Character, character vector, or NULL. Describes the location within `inSCE` where the gene identifiers in `geneSetCollection` should be mapped. If set to `"rownames"` then the features will be searched for among `rownames(inSCE)`. This can also be set to one of the column names of `rowData(inSCE)` in which case the gene identifies will be mapped to that column in the `rowData` of `inSCE`. `by` can be a vector the same length as the number of gene sets in the `GeneSetCollection` and the elements of the vector can point to different locations within `inSCE`. Finally, `by` can be `NULL`. In this case, the location of the gene identifiers in `inSCE` should be saved in the description slot for each gene set in the `GeneSetCollection`. See featureIndex for more information. Default `"rownames"`.
`noMatchError`	Boolean. Show an error if a collection does not have any matching features. Default `TRUE`.

Details

The gene identifiers in gene sets in the `GeneSetCollection` will be mapped to the rownames of `inSCE` using the `by` parameter and stored in a GeneSetCollection object from package GSEABase.

This object is stored in `metadata(inSCE)$sctk$genesets`, which can be accessed in downstream analysis functions such as runCellQC.

Value

A SingleCellExperiment object with gene set from `collectionName` output stored to the metadata slot.

Author(s)

Joshua D. Campbell

See Also

importGeneSetsFromList for importing from lists, importGeneSetsFromGMT for importing from GMT files, and importGeneSetsFromMSigDB for importing MSigDB gene sets.

Examples

```
data(scExample)
gs1 <- GSEABase::GeneSet(setName = "geneset1",
                         geneIds = rownames(sce)[seq(10)])
gs2 <- GSEABase::GeneSet(setName = "geneset2",
                         geneIds = rownames(sce)[seq(11,20)])
gsc <- GSEABase::GeneSetCollection(list(gs1, gs2))
sce <- importGeneSetsFromCollection(inSCE = sce,
                                    geneSetCollection = gsc,
                                    by = "rownames")
```

---

importGeneSetsFromGMT *Imports gene sets from a GMT file*

Description

Converts a list of gene sets stored in a GMT file into a GeneSetCollection and stores it in the metadata of the SingleCellExperiment object. These gene sets can be used in downstream quality control and analysis functions in singleCellTK.

Usage

```
importGeneSetsFromGMT(
  inSCE,
  file,
  collectionName = "GeneSetCollection",
  by = "rownames",
  sep = "\t",
  noMatchError = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
file	Character. Path to GMT file. See getGmt for more information on reading GMT files.
collectionName	Character. Name of collection to add gene sets to. If this collection already exists in inSCE, then these gene sets will be added to that collection. Any gene sets within the collection with the same name will be overwritten. Default GeneSetCollection.
by	Character, character vector, or NULL. Describes the location within inSCE where the gene identifiers in geneSetList should be mapped. If set to "rownames" then the features will be searched for among rownames(inSCE). This can also be set to one of the column names of rowData(inSCE) in which case the gene identifies will be mapped to that column in the rowData of inSCE. by can be a vector the same length as the number of gene sets in the GMT file and the elements of the vector can point to different locations within inSCE. Finally, by can be NULL. In this case, the location of the gene identifiers in inSCE should be saved in the description (2nd column) of the GMT file. See featureIndex for more information. Default "rownames".
sep	Character. Delimiter of the GMT file. Default "\t".
noMatchError	Boolean. Show an error if a collection does not have any matching features. Default TRUE.

Details

The gene identifiers in gene sets in the GMT file will be mapped to the rownames of inSCE using the by parameter and stored in a GeneSetCollection object from package GSEABase. This object is stored in metadata(inSCE)$sctk$genesets, which can be accessed in downstream analysis functions such as runCellQC.

Value

A SingleCellExperiment object with gene set from collectionName output stored to the metadata slot.

Author(s)

Joshua D. Campbell

See Also

importGeneSetsFromList for importing from lists, importGeneSetsFromCollection for importing from GeneSetCollection objects, and importGeneSetsFromMSigDB for importing MSigDB gene sets.

Examples

```
data(scExample)

# GMT file containing gene symbols for a subset of human mitochondrial genes
gmt <- system.file("extdata/mito_subset.gmt", package = "singleCellTK")

# "feature_name" is the second column in the GMT file, so the ids will
# be mapped using this column in the 'rowData' of 'sce'. This
# could also be accomplished by setting by = "feature_name" in the
# function call.
sce <- importGeneSetsFromGMT(inSCE = sce, file = gmt, by = NULL)
```

`importGeneSetsFromList` *Imports gene sets from a list*

Description

Converts a list of gene sets into a GeneSetCollection and stores it in the metadata of the SingleCellExperiment object. These gene sets can be used in downstream quality control and analysis functions in singleCellTK.

Usage

```
importGeneSetsFromList(
  inSCE,
  geneSetList,
  collectionName = "GeneSetCollection",
  by = "rownames",
  noMatchError = TRUE
)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`geneSetList`	Named List. A list containing one or more gene sets. Each element of the list should be a character vector of gene identifiers. The names of the list will be become the gene set names in the GeneSetCollection object.
`collectionName`	Character. Name of collection to add gene sets to. If this collection already exists in `inSCE`, then these gene sets will be added to that collection. Any gene sets within the collection with the same name will be overwritten. Default `GeneSetCollection`.
`by`	Character or character vector. Describes the location within `inSCE` where the gene identifiers in `geneSetList` should be mapped. If set to `"rownames"` then the features will be searched for among `rownames(inSCE)`. This can also be set to one of the column names of `rowData(inSCE)` in which case the gene

identifies will be mapped to that column in the rowData of inSCE. Finally, by can be a vector the same length as the number of gene sets in geneSetList and the elements of the vector can point to different locations within inSCE. See featureIndex for more information. Default "rownames".

noMatchError Boolean. Show an error if a collection does not have any matching features. Default TRUE.

Details

The gene identifiers in gene sets in geneSetList will be mapped to the rownames of inSCE using the by parameter and stored in a GeneSetCollection object from package GSEABase. This object is stored in metadata(inSCE)$sctk$genesets, which can be accessed in downstream analysis functions such as runCellQC.

Value

A SingleCellExperiment object with gene set from collectionName output stored to the metadata slot.

Author(s)

Joshua D. Campbell

See Also

importGeneSetsFromCollection for importing from GeneSetCollection objects, importGeneSetsFromGMT for importing from GMT files, and importGeneSetsFromMSigDB for importing MSigDB gene sets.

Examples

```
data(scExample)

# Generate gene sets from 'rownames'
gs1 <- rownames(sce)[seq(10)]
gs2 <- rownames(sce)[seq(11,20)]
gs <- list("geneset1" = gs1, "geneset2" = gs2)
sce <- importGeneSetsFromList(inSCE = sce,
                              geneSetList = gs,
                              by = "rownames")

# Generate a gene set for mitochondrial genes using
# Gene Symbols stored in 'rowData'
mito.ix <- grep("^MT-", rowData(sce)$feature_name)
mito <- list(mito = rowData(sce)$feature_name[mito.ix])
sce <- importGeneSetsFromList(inSCE = sce,
                              geneSetList = mito,
                              by = "feature_name")
```

`importGeneSetsFromMSigDB` *Imports gene sets from MSigDB*

Description

Gets a list of MSigDB gene sets stores it in the metadata of the SingleCellExperiment object. These gene sets can be used in downstream quality control and analysis functions in singleCellTK.

Usage

```
importGeneSetsFromMSigDB(
  inSCE,
  categoryIDs = "H",
  species = "Homo sapiens",
  mapping = c("gene_symbol", "human_gene_symbol", "entrez_gene"),
  by = "rownames",
  verbose = TRUE,
  noMatchError = TRUE
)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`categoryIDs`	Character vector containing the MSigDB gene set ids. The column `ID` in the table returned by `getMSigDBTable()` shows the list of possible gene set IDs that can be obtained. Default is `"H"`.
`species`	Character. Species available can be found using the function `msigdbr_show_species`. Default `"Homo sapiens"`.
`mapping`	Character. One of "gene_symbol", "human_gene_symbol", or "entrez_gene". Gene identifiers to be used for MSigDB gene sets. IDs denoted by the by parameter must be either in gene symbol or Entrez gene id format to match IDs from MSigDB.
`by`	Character. Describes the location within `inSCE` where the gene identifiers in the MSigDB gene sets should be mapped. If set to `"rownames"` then the features will be searched for among `rownames(inSCE)`. This can also be set to one of the column names of `rowData(inSCE)` in which case the gene identifies will be mapped to that column in the `rowData` of `inSCE`. See featureIndex for more information. Default `"rownames"`.
`verbose`	Boolean. Whether to display progress. Default `TRUE`.
`noMatchError`	Boolean. Show an error if a collection does not have any matching features. Default `TRUE`.

Details

The gene identifiers in gene sets from MSigDB will be retrieved using the msigdbr package. They will be mapped to the IDs in inSCE using the by parameter and stored in a GeneSetCollection object from package GSEABase. This object is stored in metadata(inSCE)$sctk$genesets, which can be accessed in downstream analysis functions such as runCellQC.

Value

A SingleCellExperiment object with gene set from collectionName output stored to the metadata slot.

Author(s)

Joshua D. Campbell

See Also

importGeneSetsFromList for importing from lists, importGeneSetsFromGMT for importing from GMT files, and GeneSetCollection objects.

Examples

```
data(scExample)
sce <- importGeneSetsFromMSigDB(inSCE = sce,
                                categoryIDs = "H",
                                species = "Homo sapiens",
                                mapping = "gene_symbol",
                                by = "feature_name")
```

---

importMitoGeneSet *Import mitochondrial gene sets*

Description

Imports mitochondrial gene sets and stores it in the metadata of the SingleCellExperiment object. These gene sets can be used in downstream quality control and analysis functions in singleCellTK.

Usage

```
importMitoGeneSet(
  inSCE,
  reference = "human",
  id = "ensembl",
  by = "rownames",
  collectionName = "mito",
  noMatchError = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
reference	Character. Species available are "human" and "mouse".
id	Types of gene id. Now it supports "symbol", "entrez", "ensembl" and "ensemblTranscriptID".
by	Character. Describes the location within inSCE where the gene identifiers in the mitochondrial gene sets should be mapped. If set to "rownames" then the features will be searched for among rownames(inSCE). This can also be set to one of the column names of rowData(inSCE) in which case the gene identifies will be mapped to that column in the rowData of inSCE. See featureIndex for more information. Default "rownames".
collectionName	Character. Name of collection to add gene sets to. If this collection already exists in inSCE, then these gene sets will be added to that collection. Any gene sets within the collection with the same name will be overwritten. Default "mito".
noMatchError	Boolean. Show an error if a collection does not have any matching features. Default TRUE.

Details

The gene identifiers of mitochondrial genes will be loaded with "data(AllMito)". Currently, it supports human and mouse references. Also, it supports entrez ID, gene symbol, ensemble ID and ensemble transcript ID. They will be mapped to the IDs in inSCE using the by parameter and stored in a GeneSetCollection object from package GSEABase. This object is stored in metadata(inSCE)$sctk$genesets, which can be accessed in downstream analysis functions such as runCellQC.

Value

A SingleCellExperiment object with gene set from collectionName output stored to the metadata slot.

Author(s)

Rui Hong

See Also

importGeneSetsFromList for importing from lists, importGeneSetsFromGMT for importing from GMT files, and GeneSetCollection objects.

Examples

```
data(scExample)
sce <- importMitoGeneSet(inSCE = sce,
                         reference = "human",
                         id = "ensembl",
                         collectionName = "human_mito",
                         by = "rownames")
```

importMultipleSources *Imports samples from different sources and compiles them into a list of SCE objects*

Description

Imports samples from different sources and compiles them into a list of SCE objects

Usage

```
importMultipleSources(allImportEntries, delayedArray = FALSE)
```

Arguments

allImportEntries	object containing the sources and parameters of all the samples being imported (from the UI)
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.

Value

A list of SingleCellExperiment object containing the droplet or cell data or both,depending on the dataType that users provided.

importOptimus *Construct SCE object from Optimus output*

Description

Read the barcodes, features (genes), and matrices from Optimus outputs. Import them as one SingleCellExperiment object.

Usage

```
importOptimus(
  OptimusDirs,
  samples,
  matrixLocation = "call-MergeCountFiles/sparse_counts.npz",
  colIndexLocation = "call-MergeCountFiles/sparse_counts_col_index.npy",
  rowIndexLocation = "call-MergeCountFiles/sparse_counts_row_index.npy",
  cellMetricsLocation = "call-MergeCellMetrics/merged-cell-metrics.csv.gz",
  geneMetricsLocation = "call-MergeGeneMetrics/merged-gene-metrics.csv.gz",
  emptyDropsLocation = "call-RunEmptyDrops/empty_drops_result.csv",
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)
```

Arguments

OptimusDirs	A vector of root directories of Optimus output files. The paths should be something like this: /PATH/TO/bb4a2a5e-ff34-41b6-97d2-0c0c0c534530. Each entry in OptimusDirs is considered a sample and should have its own path. Must have the same length as samples.
samples	A vector of user-defined sample names for the sample to be imported. Must have the same length as OptimusDirs.
matrixLocation	Character. It is the intermediate path to the filtered count maxtrix file saved in sparse matrix format (.npz). Default call-MergeCountFiles/sparse_counts.npz which works for optimus_v1.4.0.
colIndexLocation	Character. The intermediate path to the barcode index file. Default call-MergeCountFiles/sparse_cou
rowIndexLocation	Character. The intermediate path to the feature (gene) index file. Default call-MergeCountFiles/spars
cellMetricsLocation	Character. It is the intermediate path to the cell metrics file (merged-cell-metrics.csv.gz). Default call-MergeCellMetrics/merged-cell-metrics.csv.gz which works for optimus_v1.4.0.
geneMetricsLocation	Character. It is the intermediate path to the feature (gene) metrics file (merged-gene-metrics.csv.gz). Default call-MergeGeneMetrics/merged-gene-metrics.csv.gz which works for optimus_v1.4.0.
emptyDropsLocation	Character. It is the intermediate path to emptyDrops metrics file (empty_drops_result.csv). Default call-RunEmptyDrops/empty_drops_result.csv which works for optimus_v1.4.0.
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.
rowNamesDedup	Boolean. Whether to deduplicate rownames. Default TRUE.

Value

A SingleCellExperiment object containing the count matrix, the gene annotation, and the cell annotation.

Examples

```
file.path <- system.file("extdata/Optimus_20x1000",
  package = "singleCellTK")
## Not run:
sce <- importOptimus(OptimusDirs = file.path,
  samples = "Optimus_20x1000")

## End(Not run)
```

importSEQC *Construct SCE object from seqc output*

Description

Read the filtered barcodes, features, and matrices for all samples from (preferably a single run of) seqc output. Import and combine them as one big SingleCellExperiment object.

Usage

```
importSEQC(
  seqcDirs = NULL,
  samples = NULL,
  prefix = NULL,
  gzipped = FALSE,
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  cbNotFirstCol = TRUE,
  feNotFirstCol = TRUE,
  combinedSample = TRUE,
  rowNamesDedup = TRUE
)
```

Arguments

seqcDirs	A vector of paths to seqc output files. Each sample should have its own path. For example: "./pbmc_1k_50x50". Must have the same length as samples.
samples	A vector of user-defined sample names for the samples to be imported. Must have the same length as seqcDirs.
prefix	A vector containing the prefix of file names within each sample directory. It cannot be null and the vector should have the same length as *samples*.
gzipped	Boolean. TRUE if the seqc output files (sparse_counts_barcode.csv, sparse_counts_genes.csv, and sparse_molecule_counts.mtx) were gzip compressed. FALSE otherwise. Default seqc outputs are not gzipped. Default FALSE.
class	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
delayedArray	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default FALSE.
cbNotFirstCol	Boolean. TRUE if first column of sparse_counts_barcode.csv is row index and it will be removed. FALSE the first column will be kept.
feNotFirstCol	Boolean. TRUE if first column of sparse_counts_genes.csv is row index and it will be removed. FALSE the first column will be kept.

combinedSample Boolean. If TRUE, importSEQC returns a SingleCellExperiment object containing the combined count matrix, feature annotations and the cell annotations. If FALSE, importSEQC returns a list containing multiple SingleCellExperiment objects. Each SingleCellExperiment contains count matrix, feature annotations and cell annotations for each sample.

rowNamesDedup Boolean. Whether to deduplicate rownames. Only applied if combinedSample is TRUE or only one seqcDirs specified. Default TRUE.

Details

importSEQC imports output from seqc. The default sparse_counts_barcode.csv or sparse_counts_genes.csv from seqc output contains two columns. The first column is row index and the second column is cell-barcode or gene symbol. importSEQC will remove first column. Alternatively, user can call cbNotFirstCol or feNotFirstCol as FALSE to keep the first column of these files. When combinedSample is TRUE, importSEQC will combined count matrix with genes detected in at least one sample.

Value

A SingleCellExperiment object containing the combined count matrix, the feature annotations, and the cell annotation.

Examples

```
# Example #1
# The following filtered feature, cell, and matrix files were downloaded from
# https://support.10xgenomics.com/single-cell-gene-expression/datasets/
# 3.0.0/pbmc_1k_v3
# The top 50 hg38 genes are included in this example.
# Only the top 50 cells are included.
sce <- importSEQC(
    seqcDirs = system.file("extdata/pbmc_1k_50x50", package = "singleCellTK"),
    samples = "pbmc_1k_50x50",
    prefix = "pbmc_1k",
    combinedSample = FALSE)
```

---

importSTARsolo *Construct SCE object from STARsolo outputs*

Description

Read the barcodes, features (genes), and matrices from STARsolo outputs. Import them as one SingleCellExperiment object.

Usage

```
importSTARsolo(
  STARsoloDirs,
  samples,
  STARsoloOuts = c("Gene", "GeneFull"),
  matrixFileNames = "matrix.mtx",
  featuresFileNames = "features.tsv",
  barcodesFileNames = "barcodes.tsv",
  gzipped = "auto",
  class = c("Matrix", "matrix"),
  delayedArray = FALSE,
  rowNamesDedup = TRUE
)
```

Arguments

`STARsoloDirs`	A vector of root directories of STARsolo output files. The paths should be something like this: **/PATH/TO/*prefix*Solo.out**. For example: `./Solo.out`. Each sample should have its own path. Must have the same length as `samples`.
`samples`	A vector of user-defined sample names for the sample to be imported. Must have the same length as `STARsoloDirs`.
`STARsoloOuts`	Character. The intermediate folder to filtered or raw cell barcode, feature, and matrix files for each of `samples`. Default `"Gene"`. It can be either Gene or GeneFull as the main folder from which data needs to be imported.
`matrixFileNames`	Filenames for the Market Exchange Format (MEX) sparse matrix file (.mtx file). Must have length 1 or the same length as `samples`.
`featuresFileNames`	Filenames for the feature annotation file. Must have length 1 or the same length as `samples`.
`barcodesFileNames`	Filenames for the cell barcode list file. Must have length 1 or the same length as `samples`.
`gzipped`	Boolean. `TRUE` if the STARsolo output files (barcodes.tsv, features.tsv, and matrix.mtx) were gzip compressed. `FALSE` otherwise. This is `FALSE` in STAR 2.7.3a. Default `"auto"` which automatically detects if the files are gzip compressed. Must have length 1 or the same length as `samples`.
`class`	Character. The class of the expression matrix stored in the SCE object. Can be one of "Matrix" (as returned by readMM function), or "matrix" (as returned by matrix function). Default "Matrix".
`delayedArray`	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default `FALSE`.
`rowNamesDedup`	Boolean. Whether to deduplicate rownames. Default `TRUE`.

Value

A SingleCellExperiment object containing the count matrix, the gene annotation, and the cell annotation.

Examples

```
# Example #1
# FASTQ files were downloaded from
# https://support.10xgenomics.com/single-cell-gene-expression/datasets/3.0.0
# /pbmc_1k_v3
# They were concatenated as follows:
# cat pbmc_1k_v3_S1_L001_R1_001.fastq.gz pbmc_1k_v3_S1_L002_R1_001.fastq.gz >
# pbmc_1k_v3_R1.fastq.gz
# cat pbmc_1k_v3_S1_L001_R2_001.fastq.gz pbmc_1k_v3_S1_L002_R2_001.fastq.gz >
# pbmc_1k_v3_R2.fastq.gz
# The following STARsolo command generates the filtered feature, cell, and
# matrix files
# STAR \
#   --genomeDir ./index \
#   --readFilesIn ./pbmc_1k_v3_R2.fastq.gz \
#                 ./pbmc_1k_v3_R1.fastq.gz \
#   --readFilesCommand zcat \
#   --outSAMtype BAM Unsorted \
#   --outBAMcompression -1 \
#   --soloType CB_UMI_Simple \
#   --soloCBwhitelist ./737K-august-2016.txt \
#   --soloUMIlen 12

# The top 20 genes and the first 20 cells are included in this example.
sce <- importSTARsolo(
  STARsoloDirs = system.file("extdata/STARsolo_PBMC_1k_v3_20x20",
    package = "singleCellTK"),
  samples = "PBMC_1k_v3_20x20")
```

---

iterateSimulations *Returns significance data from a snapshot.*

Description

Returns significance data from a snapshot.

Usage

```
iterateSimulations(
  originalData,
  useAssay = "counts",
  realLabels,
  totalReads,
  cells,
```

```
  iterations
)
```

Arguments

originalData	The SingleCellExperiment object storing all assay data from the shiny app.
useAssay	Character. The name of the assay to be used for subsampling.
realLabels	Character. The name of the condition of interest. Must match a name from sample data.
totalReads	Numeric. The total number of reads in the simulated dataset, to be split between all simulated cells.
cells	Numeric. The number of virtual cells to simulate.
iterations	Numeric. How many times should each experimental design be simulated.

Value

A matrix of significance information from a snapshot

Examples

```
data("mouseBrainSubsetSCE")
res <- iterateSimulations(mouseBrainSubsetSCE, realLabels = "level1class",
                          totalReads = 1000, cells = 10, iterations = 2)
```

---

listSampleSummaryStatsTables

Lists the table of SCTK QC outputs stored within the metadata.

Description

Returns a character vector of the tables within the metadata slot of the SingleCellExperiment object.

Usage

```
listSampleSummaryStatsTables(inSCE, ...)

## S4 method for signature 'SingleCellExperiment'
listSampleSummaryStatsTables(inSCE, ...)
```

Arguments

inSCE	Input SingleCellExperiment object with saved table within the metadata data. Required.
...	Other arguments passed to the function.

Value

A character vector. Contains a list of summary tables within the SingleCellExperiment object.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- sampleSummaryStats(sce, simple = TRUE, statsName = "qc_table")
listSampleSummaryStatsTables(sce)
```

---

mergeSCEColData *Merging colData from two singleCellExperiment objects*

Description

Merges colData of the singleCellExperiment objects obtained from the same dataset which contain differing colData. (i.e. raw data and filtered data)

Usage

```
mergeSCEColData(inSCE1, inSCE2, id1 = "column_name", id2 = "column_name")
```

Arguments

inSCE1	Input SingleCellExperiment object. The function will output this singleCellExperiment object with a combined colData from inSCE1 and inSCE2.
inSCE2	Input SingleCellExperiment object. colData from this object will be merged with colData from inSCE1 and loaded into inSCE1.
id1	Character vector. Column in colData of inSCE1 that will be used to combine inSCE1 and inSCE2. Default "column_name"
id2	Character vector. Column in colData of inSCE2 that will be used to combine inSCE1 and inSCE2. Default "column_name"

Value

SingleCellExperiment object containing combined colData from both singleCellExperiment for samples in inSCE1.

Examples

```
sce1 <- importCellRanger(
    cellRangerDirs = system.file("extdata/", package = "singleCellTK"),
    sampleDirs = "hgmm_1k_v3_20x20",
    sampleNames = "hgmm1kv3",
    dataType = "filtered")
data(scExample)
sce2 <- sce
sce <- mergeSCEColData(inSCE1 = sce1, inSCE2 = sce2, id1 = "column_name", id2 = "column_name")
```

MitoGenes *List of mitochondrial genes of multiple reference*

Description

A list of gene set that contains mitochondrial genes of multiple reference (hg38, hg19, mm10 and mm9). It contains multiple types of gene identifier: gene symbol, entrez ID, ensemble ID and ensemble transcript ID. It's used for the function 'importMitoGeneSet'.

Usage

```
data("MitoGenes")
```

Format

A list

Value

List of mitochondrial genes of multiple reference

Examples

```
data("MitoGenes")
```

mouseBrainSubsetSCE *Example Single Cell RNA-Seq data in SingleCellExperiment Object, GSE60361 subset*

Description

A subset of 30 cells from a single cell RNA-Seq experiment from Zeisel, et al. Science 2015. The data was produced from cells from the mouse somatosensory cortex (S1) and hippocampus (CA1). 15 of the cells were identified as oligodendrocytes and 15 of the cell were identified as microglia.

Usage

```
data("mouseBrainSubsetSCE")
```

Format

SingleCellExperiment

Value

A subset of 30 cells from a single cell RNA-Seq experiment

Source

DOI: 10.1126/science.aaa1934

Examples

```
data("mouseBrainSubsetSCE")
```

`msigdb_table` *MSigDB gene get Category table*

Description

A table of gene set categories that can be download from MSigDB. The categories and descriptions can be found here: https://www.gsea-msigdb.org/gsea/msigdb/collections.jsp. The IDs in the first column can be used to retrieve the gene sets for these categories using the importGeneSetsFromMSigDB function.

Usage

```
data("msigdb_table")
```

Format

A data.frame.

Value

A table of gene set categories

Examples

```
data("msigdb_table")
```

`plotBarcodeRankDropsResults`
Plots for runBarcodeRankDrops outputs.

Description

A wrapper function which visualizes outputs from the `runBarcodeRankDrops` function stored in the `metadata` slot of the SingleCellExperiment object.

Usage

```
plotBarcodeRankDropsResults(
  inSCE,
  sample = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  titleSize = 18,
  axisSize = 15,
  axisLabelSize = 18,
  legendSize = 15
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runBarcodeRankDrops. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
titleSize	Size of title of plot. Default 18.
axisSize	Size of x/y-axis ticks. Default 15.
axisLabelSize	Size of x/y-axis labels. Default 18.
legendSize	size of legend. Default 15.

Value

list of .ggplot objects

Examples

```
data(scExample, package = "singleCellTK")
sce <- runBarcodeRankDrops(inSCE = sce)
plotBarcodeRankDropsResults(inSCE = sce)
```

---

plotBarcodeRankScatter

Plots for runBarcodeRankDrops outputs.

Description

A plotting function which visualizes outputs from the runBarcodeRankDrops function stored in the colData slot of the SingleCellExperiment object via scatterplot.

Usage

```
plotBarcodeRankScatter(
  inSCE,
  sample = NULL,
  defaultTheme = TRUE,
  dotSize = 0.1,
  title = NULL,
  titleSize = 18,
  xlab = NULL,
  ylab = NULL,
  axisSize = 12,
  axisLabelSize = 15,
  legendSize = 10,
  combinePlot = "none",
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runBarcodeRankDrops. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.1.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 18.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 12.
axisLabelSize	Size of x/y-axis labels. Default 15.
legendSize	size of legend. Default 10.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
sampleRelHeights	If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.
sampleRelWidths	If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

a ggplot object of the scatter plot.

See Also

plotBarcodeRankDropsResults, runBarcodeRankDrops

Examples

```
data(scExample, package = "singleCellTK")
sce <- runBarcodeRankDrops(inSCE = sce)
plotBarcodeRankScatter(inSCE = sce)
```

---

plotBatchCorrCompare *Plot comparison of batch corrected result against original assay*

Description

Plot comparison of batch corrected result against original assay

Usage

```
plotBatchCorrCompare(
  inSCE,
  corrMat,
  batch = NULL,
  condition = NULL,
  origAssay = NULL,
  origLogged = NULL,
  method = NULL,
  matType = NULL
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
corrMat	A single character indicating the name of the corrected matrix.
batch	A single character. The name of batch annotation column in colData(inSCE).
condition	A single character. The name of an additional covariate annotation column in colData(inSCE).
origAssay	A single character indicating what the original assay used for batch correction is.
origLogged	Logical scalar indicating whether origAssay is log-normalized.
method	A single character indicating the name of the batch correction method. Only used for the titles of plots.
matType	A single character indicating the type of the batch correction result matrix, choose from "assay", "altExp", "reducedDim".

Details

Four plots will be combined. Two of them are violin/box-plots for percent variance explained by the batch variation, and optionally the covariate, for original and corrected. The other two are UMAPs of the original assay and the correction result matrix. If SCTK batch correction methods are performed in advance, this function will automatically detect necessary input. Otherwise, users can also customize the input. Future improvement might include solution to reduce redundant UMAP calculation.

Value

An object of class "gtable", combining four ggplots.

Author(s)

Yichen Wang

Examples

```
data("sceBatches")
logcounts(sceBatches) <- log1p(counts(sceBatches))
sceBatches <- runLimmaBC(sceBatches)
plotBatchCorrCompare(sceBatches, "LIMMA", condition = "cell_type")
```

---

plotBatchVariance *Plot the percent of the variation that is explained by batch and condition in the data*

Description

Visualize the percent variation in the data that is explained by batch and condition, individually, and that explained by combining both annotations. Plotting only the variation explained by batch is supported but not recommended, because this can be confounded by potential condition.

Usage

```
plotBatchVariance(
  inSCE,
  useAssay = NULL,
  useReddim = NULL,
  useAltExp = NULL,
  batch = "batch",
  condition = NULL,
  title = NULL
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
useAssay	A single character. The name of the assay that stores the value to plot. For useReddim and useAltExp also. Default NULL.
useReddim	A single character. The name of the dimension reduced matrix that stores the value to plot. Default NULL.
useAltExp	A single character. The name of the alternative experiment that stores an assay of the value to plot. Default NULL.
batch	A single character. The name of batch annotation column in colData(inSCE). Default "batch".
condition	A single character. The name of an additional condition annotation column in colData(inSCE). Default NULL.
title	A single character. The title text on the top. Default NULL.

Details

When condition and batch both are causing some variation, if the difference between full variation and condition variation is close to batch variation, this might imply that batches are causing some effect; if the difference is much less than batch variation, then the batches are likely to be confounded by the conditions.

Value

A ggplot object of a boxplot of variation explained by batch, condition, and batch+condition.

Examples

```
data('sceBatches', package = 'singleCellTK')
plotBatchVariance(sceBatches,
                  useAssay="counts",
                  batch="batch",
                  condition = "cell_type")
```

---

plotBcdsResults *Plots for runBcds outputs.*

Description

A wrapper function which visualizes outputs from the runBcds function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotBcdsResults(
  inSCE,
  sample = NULL,
  shape = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
  dots = TRUE,
  reducedDimName = "UMAP",
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  summary = "median",
  summaryTextSize = 3,
  transparency = 1,
  baseSize = 15,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL,
  relHeights = 1,
  relWidths = c(1, 1, 1),
  plotNCols = NULL,
  plotNRows = NULL,
  labelSamples = TRUE,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runBcds. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
shape	If provided, add shapes based on the value. Default NULL.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in inSCE, or can be retrieved from the colData slot. Default NULL.

combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
reducedDimName	Saved dimension reduction name in inSCE. Default "UMAP".
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Similar to dim1. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into bin groups. If more than one value, will bin numeric values using values as a cut point. Default NULL.
binLabel	Character vector. Labels for the bins created by bin. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
transparency	Transparency of the dots, values will be 0-1. Default 1.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
titleSize	Size of title of plot. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
axisSize	Size of x/y-axis ticks. Default NULL.
legendSize	size of legend. Default NULL.
legendTitleSize	size of legend title. Default NULL.
relHeights	Relative heights of plots when combine is set. Default 1.
relWidths	Relative widths of plots when combine is set. Default c(1, 1, 1).
plotNCols	Number of columns when plots are combined in a grid. Default NULL.
plotNRows	Number of rows when plots are combined in a grid. Default NULL.
labelSamples	Will label sample name in title of plot if TRUE. Default TRUE.
samplePerColumn	If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.

sampleRelHeights
: If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.

sampleRelWidths
: If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

list of .ggplot objects

See Also

runBcds

Examples

```
data(scExample, package="singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runQuickUMAP(sce)
sce <- runBcds(sce)
plotBcdsResults(inSCE=sce, reducedDimName="UMAP")
```

---

plotBubble *Plot Bubble plot*

Description

Plot a bubble plot with the color of the plot being the mean expression and the size of the dot being the percent of cells in the cluster expressing the gene.

Usage

```
plotBubble(
  inSCE,
  useAssay = "logcounts",
  featureNames,
  displayName = NULL,
  groupNames = "cluster",
  title = "",
  xlab = NULL,
  ylab = NULL,
  colorLow = "white",
  colorHigh = "blue",
  scale = FALSE
)
```

Arguments

inSCE	The single cell experiment to use.
useAssay	The assay to use.
featureNames	A string or vector of strings with each gene to aggregate.
displayName	A string that is the name of the column used for genes.
groupNames	The name of a colData entry that can be used as groupNames.
title	The title of the bubble plot
xlab	The x-axis label
ylab	The y-axis label
colorLow	The color to be used for lowest value of mean expression
colorHigh	The color to be used for highest value of mean expression
scale	Option to scale the data. Default: /codeFALSE. Selected assay will not be scaled.

Value

A ggplot of the bubble plot.

Examples

```
data("scExample")
plotBubble(inSCE=sce, useAssay="counts", featureNames=c("B2M", "MALAT1"),
displayName="feature_name", groupNames="type", title="cell type test",
xlab="gene", ylab="cluster", colorLow="white", colorHigh="blue")
```

---

plotClusterAbundance *Plot the differential Abundance*

Description

Plot the differential Abundance

Usage

```
plotClusterAbundance(inSCE, cluster, variable, combinePlot = c("all", "none"))
```

Arguments

inSCE	A SingleCellExperiment object.
cluster	A single character, specifying the name to store the cluster label in colData.
variable	A single character, specifying the name to store the phenotype labels in colData.
combinePlot	Must be either "all" or "none". "all" will combine all plots into a single ggplot object. Default "all".

Details

This function will visualize the differential abundance in two given variables, by making bar plots that presents the cell counting and fraction in different cases.

Value

When `combinePlot = "none"`, a `list` with 4 `ggplot` objects; when `combinePlot = "all"`, a single `ggplot` object with for subplots.

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
plotClusterAbundance(inSCE = mouseBrainSubsetSCE,
                     cluster = "tissue",
                     variable = "level1class")
```

`plotCxdsResults` *Plots for runCxds outputs.*

Description

A wrapper function which visualizes outputs from the `runCxds` function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotCxdsResults(
  inSCE,
  sample = NULL,
  shape = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
  dots = TRUE,
  reducedDimName = "UMAP",
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  summary = "median",
  summaryTextSize = 3,
  transparency = 1,
```

```
  baseSize = 15,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL,
  relHeights = 1,
  relWidths = c(1, 1, 1),
  plotNCols = NULL,
  plotNRows = NULL,
  labelSamples = TRUE,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runCxds. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
shape	If provided, add shapes based on the value. Default NULL.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in inSCE, or can be retrieved from the colData slot. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
reducedDimName	Saved dimension reduction name in inSCE. Default "UMAP".
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Similar to dim1. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into bin groups. If more than one value, will bin numeric values using values as a cut point. Default NULL.
binLabel	Character vector. Labels for the bins created by bin. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.

dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
transparency	Transparency of the dots, values will be 0-1. Default 1.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
titleSize	Size of title of plot. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
axisSize	Size of x/y-axis ticks. Default NULL.
legendSize	size of legend. Default NULL.
legendTitleSize	size of legend title. Default NULL.
relHeights	Relative heights of plots when combine is set. Default 1.
relWidths	Relative widths of plots when combine is set. Default c(1, 1, 1).
plotNCols	Number of columns when plots are combined in a grid. Default NULL.
plotNRows	Number of rows when plots are combined in a grid. Default NULL.
labelSamples	Will label sample name in title of plot if TRUE. Default TRUE.
samplePerColumn	If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.
sampleRelHeights	If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.
sampleRelWidths	If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

list of .ggplot objects

See Also

runCxds

Examples

```
data(scExample, package="singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runQuickUMAP(sce)
sce <- runCxds(sce)
plotCxdsResults(inSCE=sce, reducedDimName="UMAP")
```

plotDecontXResults *Plots for runDecontX outputs.*

Description

A wrapper function which visualizes outputs from the runDecontX function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotDecontXResults(
  inSCE,
  sample = NULL,
  bgResult = FALSE,
  shape = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
  dots = TRUE,
  reducedDimName = "UMAP",
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  summary = "median",
  summaryTextSize = 3,
  transparency = 1,
  baseSize = 15,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL,
  relHeights = 1,
  relWidths = c(1, 1, 1),
  plotNCols = NULL,
  plotNRows = NULL,
  labelSamples = TRUE,
  labelClusters = TRUE,
  clusterLabelSize = 3.5,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
```

```
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runDecontX. Required.
sample	Character vector. Indicates which sample each cell belongs to. Default NULL.
bgResult	Boolean. If TRUE, will plot decontX results generated with raw/droplet matrix Default FALSE.
shape	If provided, add shapes based on the value.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
reducedDimName	Saved dimension reduction name in the SingleCellExperiment object. Required. Default = "UMAP"
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into the 'bin' groups. If more than one value, will bin numeric values using values as a cut point.
binLabel	Character vector. Labels for the bins created by the 'bin' parameter. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
transparency	Transparency of the dots, values will be 0-1. Default 1.

baseSize The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.

titleSize Size of title of plot. Default NULL.

axisLabelSize Size of x/y-axis labels. Default NULL.

axisSize Size of x/y-axis ticks. Default NULL.

legendSize size of legend. Default NULL.

legendTitleSize
size of legend title. Default NULL.

relHeights Relative heights of plots when combine is set.

relWidths Relative widths of plots when combine is set.

plotNCols Number of columns when plots are combined in a grid.

plotNRows Number of rows when plots are combined in a grid.

labelSamples Will label sample name in title of plot if TRUE. Default TRUE.

labelClusters Logical. Whether the cluster labels are plotted. Default FALSE.

clusterLabelSize
Numeric. Determines the size of cluster label when ‘labelClusters‘ is set to TRUE. Default 3.5.

samplePerColumn
If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.

sampleRelHeights
If there are multiple samples and combining by "all", the relative heights for each plot.

sampleRelWidths
If there are multiple samples and combining by "all", the relative widths for each plot.

Value

list of .ggplot objects

Examples

```
data(scExample, package="singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runDecontX(sce)
plotDecontXResults(inSCE=sce, reducedDimName="decontX_UMAP")
```

plotDEGHeatmap *Heatmap visualization of DEG result*

Description

Heatmap visualization of DEG result

Usage

```
plotDEGHeatmap(
  inSCE,
  useResult,
  onlyPos = FALSE,
  log2fcThreshold = 0.25,
  fdrThreshold = 0.05,
  minGroup1MeanExp = NULL,
  maxGroup2MeanExp = NULL,
  minGroup1ExprPerc = NULL,
  maxGroup2ExprPerc = NULL,
  useAssay = NULL,
  doLog = FALSE,
  featureAnnotations = NULL,
  cellAnnotations = NULL,
  featureAnnotationColor = NULL,
  cellAnnotationColor = NULL,
  rowDataName = NULL,
  colDataName = NULL,
  colSplitBy = "condition",
  rowSplitBy = "regulation",
  rowLabel = S4Vectors::metadata(inSCE)$featureDisplay,
  title = paste0("DE Analysis: ", useResult),
  ...
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
useResult	character. A string specifying the analysisName used when running a differential expression analysis function.
onlyPos	logical. Whether to only plot DEG with positive log2_FC value. Default FALSE.
log2fcThreshold	numeric. Only plot DEGs with the absolute values of log2FC larger than this value. Default 0.25.
fdrThreshold	numeric. Only plot DEGs with FDR value smaller than this value. Default 0.05.

minGroup1MeanExp
: numeric. Only plot DEGs with mean expression in group1 greater then this value. Default NULL.

maxGroup2MeanExp
: numeric. Only plot DEGs with mean expression in group2 less then this value. Default NULL.

minGroup1ExprPerc
: numeric. Only plot DEGs expressed in greater then this fraction of cells in group1. Default NULL.

maxGroup2ExprPerc
: numeric. Only plot DEGs expressed in less then this fraction of cells in group2. Default NULL.

useAssay
: character. A string specifying an assay of expression value to plot. By default the assay used for runMAST() will be used. Default NULL.

doLog
: Logical scalar. Whether to do log(assay + 1) transformation on the assay used for the analysis. Default FALSE.

featureAnnotations
: data.frame, with rownames containing all the features going to be plotted. Character columns should be factors. Default NULL.

cellAnnotations
: data.frame, with rownames containing all the cells going to be plotted. Character columns should be factors. Default NULL.

featureAnnotationColor
: A named list. Customized color settings for feature labeling. Should match the entries in the featureAnnotations or rowDataName. For each entry, there should be a list/vector of colors named with categories. Default NULL.

cellAnnotationColor
: A named list. Customized color settings for cell labeling. Should match the entries in the cellAnnotations or colDataName. For each entry, there should be a list/vector of colors named with categories. Default NULL.

rowDataName
: character. The column name(s) in rowData that need to be added to the annotation. Default NULL.

colDataName
: character. The column name(s) in colData that need to be added to the annotation. Default NULL.

colSplitBy
: character. Do semi-heatmap based on the grouping of this(these) annotation(s). Should exist in either colDataName or names(cellAnnotations). Default "condition".

rowSplitBy
: character. Do semi-heatmap based on the grouping of this(these) annotation(s). Should exist in either rowDataName or names(featureAnnotations). Default "regulation".

rowLabel
: FALSE for not displaying; a variable in rowData to display feature identifiers stored there; if have run setSCTKDisplayRow, display the specified feature name; TRUE for the rownames of inSCE; NULL for auto-display rownames when the number of filtered feature is less than 60. Default looks for setSCTKDisplayRow information.

title
: character. Main title of the heatmap. Default "DE Analysis: <useResult>".

...
: Other arguments passed to plotSCEHeatmap

Details

A differential expression analysis function has to be run in advance so that information is stored in the metadata of the input SCE object. This function wraps plotSCEHeatmap. A feature annotation basing on the log2FC level called "regulation" will be automatically added. A cell annotation basing on the condition selection while running the analysis called "condition", and the annotations used from colData(inSCE) while setting the condition and covariates will also be added.

Value

A ggplot object

Author(s)

Yichen Wang

Examples

```
data("sceBatches")
logcounts(sceBatches) <- log1p(counts(sceBatches))
sce.w <- subsetSCECols(sceBatches, colData = "batch == 'w'")
sce.w <- runWilcox(sce.w, class = "cell_type",
                   classGroup1 = "alpha", classGroup2 = "beta",
                   groupName1 = "w.alpha", groupName2 = "w.beta",
                   analysisName = "w.aVSb")
plotDEGHeatmap(sce.w, "w.aVSb")
```

---

plotDEGRegression *Create linear regression plot to show the expression the of top DEGs*

Description

Create linear regression plot to show the expression the of top DEGs

Usage

```
plotDEGRegression(
  inSCE,
  useResult,
  threshP = FALSE,
  labelBy = NULL,
  nrow = 6,
  ncol = 6,
  defaultTheme = TRUE,
  isLogged = TRUE,
  check_sanity = TRUE
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
useResult	character. A string specifying the analysisName used when running a differential expression analysis function.
threshP	logical. Whether to plot threshold values from adaptive thresholding, instead of using the assay used by when performing DE analysis. Default FALSE.
labelBy	A single character for a column of rowData(inSCE) as where to search for the labeling text. Default NULL.
nrow	Integer. Number of rows in the plot grid. Default 6.
ncol	Integer. Number of columns in the plot grid. Default 6.
defaultTheme	Logical scalar. Whether to use default SCTK theme in ggplot. Default TRUE.
isLogged	Logical scalar. Whether the assay used for the analysis is logged. If not, will do a log(assay + 1) transformation. Default TRUE.
check_sanity	Logical scalar. Whether to perform MAST's sanity check to see if the counts are logged. Default TRUE

Details

Any of the differential expression analysis method from SCTK should be performed prior to using this function

Value

A ggplot object of linear regression

Examples

```
data("sceBatches")
logcounts(sceBatches) <- log1p(counts(sceBatches))
sce.w <- subsetSCECols(sceBatches, colData = "batch == 'w'")
sce.w <- runWilcox(sce.w, class = "cell_type",
                   classGroup1 = "alpha", classGroup2 = "beta",
                   groupName1 = "w.alpha", groupName2 = "w.beta",
                   analysisName = "w.aVSb")
plotDEGRegression(sce.w, "w.aVSb")
```

---

plotDEGViolin *Generate violin plot to show the expression of top DEGs*

Description

Generate violin plot to show the expression of top DEGs

Usage

```
plotDEGViolin(
  inSCE,
  useResult,
  threshP = FALSE,
  labelBy = NULL,
  nrow = 6,
  ncol = 6,
  defaultTheme = TRUE,
  isLogged = TRUE,
  check_sanity = TRUE
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
useResult	character. A string specifying the analysisName used when running a differential expression analysis function.
threshP	logical. Whether to plot threshold values from adaptive thresholding, instead of using the assay used by runMAST(). Default FALSE.
labelBy	A single character for a column of rowData(inSCE) as where to search for the labeling text. Default NULL.
nrow	Integer. Number of rows in the plot grid. Default 6.
ncol	Integer. Number of columns in the plot grid. Default 6.
defaultTheme	Logical scalar. Whether to use default SCTK theme in ggplot. Default TRUE.
isLogged	Logical scalar. Whether the assay used for the analysis is logged. If not, will do a log(assay + 1) transformation. Default TRUE.
check_sanity	Logical scalar. Whether to perform MAST's sanity check to see if the counts are logged. Default TRUE

Details

Any of the differential expression analysis method from SCTK should be performed prior to using this function

Value

A ggplot object of violin plot

Examples

```
data("sceBatches")
logcounts(sceBatches) <- log1p(counts(sceBatches))
sce.w <- subsetSCECols(sceBatches, colData = "batch == 'w'")
sce.w <- runWilcox(sce.w, class = "cell_type",
                   classGroup1 = "alpha", classGroup2 = "beta",
                   groupName1 = "w.alpha", groupName2 = "w.beta",
```

```
                    analysisName = "w.aVSb")
plotDEGViolin(sce.w, "w.aVSb")
```

`plotDEGVolcano` *Generate volcano plot for DEGs*

Description

Generate volcano plot for DEGs

Usage

```
plotDEGVolcano(
  inSCE,
  useResult,
  labelTopN = 10,
  log2fcThreshold = 0.25,
  fdrThreshold = 0.05,
  featureDisplay = S4Vectors::metadata(inSCE)$featureDisplay
)
```

Arguments

`inSCE`	SingleCellExperiment inherited object.
`useResult`	character. A string specifying the `analysisName` used when running a differential expression analysis function.
`labelTopN`	Integer, label this number of top DEGs that pass the filters. `FALSE` for not labeling. Default `10`.
`log2fcThreshold`	numeric. Label genes with the absolute values of log2FC greater than this value as regulated. Default `0.25`.
`fdrThreshold`	numeric. Label genes with FDR value less than this value as regulated. Default `0.05`.
`featureDisplay`	A character string to indicate a variable in `rowData(inSCE)` for feature labeling. `NULL` for using `rownames`. Default `metadata(inSCE)$featureDisplay` (see `setSCTKDisplayRow`)

Details

Any of the differential expression analysis method from SCTK should be performed prior to using this function to generate volcano plots.

Value

A `ggplot` object of volcano plot

See Also

runDEAnalysis, plotDEGHeatmap

Examples

```
data("sceBatches")
sceBatches <- scaterlogNormCounts(sceBatches, "logcounts")
sce.w <- subsetSCECols(sceBatches, colData = "batch == 'w'")
sce.w <- runWilcox(sce.w, class = "cell_type",
                   classGroup1 = "alpha", classGroup2 = "beta",
                   groupName1 = "w.alpha", groupName2 = "w.beta",
                   analysisName = "w.aVSb")
plotDEGVolcano(sce.w, "w.aVSb")
```

---

plotDimRed *Plot dimensionality reduction from computed metrics including PCA, ICA, tSNE and UMAP*

Description

Plot dimensionality reduction from computed metrics including PCA, ICA, tSNE and UMAP

Usage

```
plotDimRed(
  inSCE,
  useReduction = "PCA",
  showLegend = FALSE,
  xDim = 1,
  yDim = 2,
  xAxisLabel = NULL,
  yAxisLabel = NULL
)
```

Arguments

inSCE	Input SCE object
useReduction	Reduction to plot. Default is "PCA".
showLegend	If legends should be plotted or not
xDim	Numeric value indicating the dimension to use for X-axis. Default is 1 (refers to PC1).
yDim	Numeric value indicating the dimension to use for Y-axis. Default is 2 (refers to PC2).
xAxisLabel	Specify the label for x-axis. Default is NULL which will specify the label as 'x'.
yAxisLabel	Specify the label for y-axis. Default is NULL which will specify the label as 'y'.

Value

plot object

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
plotDimRed(mouseBrainSubsetSCE, "PCA_logcounts")
```

`plotDoubletFinderResults` *Plots for runDoubletFinder outputs.*

Description

A wrapper function which visualizes outputs from the runDoubletFinder function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotDoubletFinderResults(
  inSCE,
  sample = NULL,
  shape = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
  dots = TRUE,
  reducedDimName = "UMAP",
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  summary = "median",
  summaryTextSize = 3,
  transparency = 1,
  baseSize = 15,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL,
  relHeights = 1,
```

```
  relWidths = c(1, 1, 1),
  plotNCols = NULL,
  plotNRows = NULL,
  labelSamples = TRUE,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runDoubletFinder. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
shape	If provided, add shapes based on the value. Default NULL.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in inSCE, or can be retrieved from the colData slot. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
reducedDimName	Saved dimension reduction name in inSCE. Default "UMAP".
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Similar to dim1. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into bin groups. If more than one value, will bin numeric values using values as a cut point. Default NULL.
binLabel	Character vector. Labels for the bins created by bin. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
transparency	Transparency of the dots, values will be 0-1. Default 1.

baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
titleSize	Size of title of plot. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
axisSize	Size of x/y-axis ticks. Default NULL.
legendSize	size of legend. Default NULL.
legendTitleSize	size of legend title. Default NULL.
relHeights	Relative heights of plots when combine is set. Default 1.
relWidths	Relative widths of plots when combine is set. Default c(1, 1, 1).
plotNCols	Number of columns when plots are combined in a grid. Default NULL.
plotNRows	Number of rows when plots are combined in a grid. Default NULL.
labelSamples	Will label sample name in title of plot if TRUE. Default TRUE.
samplePerColumn	If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.
sampleRelHeights	If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.
sampleRelWidths	If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

list of .ggplot objects

See Also

runDoubletFinder

Examples

```
data(scExample, package="singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runQuickUMAP(sce)
sce <- runDoubletFinder(sce)
plotDoubletFinderResults(inSCE = sce, reducedDimName = "UMAP")
```

plotEmptyDropsResults *Plots for runEmptyDrops outputs.*

Description

A wrapper function which visualizes outputs from the runEmptyDrops function stored in the colData slot of the SingleCellExperiment object.

Usage

```
plotEmptyDropsResults(
  inSCE,
  sample = NULL,
  combinePlot = "all",
  fdrCutoff = 0.01,
  defaultTheme = TRUE,
  dotSize = 0.5,
  titleSize = 18,
  axisLabelSize = 18,
  axisSize = 15,
  legendSize = 15,
  legendTitleSize = 16,
  relHeights = 1,
  relWidths = 1,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runEmptyDrops. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
combinePlot	Must be either "all", "sample", or object, "none". "all" will combine all plots into a single .ggplot while "sample" will output a list of plots separated by sample. Default "all".
fdrCutoff	Numeric. Thresholds barcodes based on the FDR values from runEmptyDrops as "Empty Droplet" or "Putative Cell". Default 0.01.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
titleSize	Size of title of plot. Default 18.
axisLabelSize	Size of x/y-axis labels. Default 18.
axisSize	Size of x/y-axis ticks. Default 15.

legendSize size of legend. Default 15.

legendTitleSize
size of legend title. Default 16.

relHeights Relative heights of plots when combine is set. Default 1.

relWidths Relative widths of plots when combine is set. Default 1.

samplePerColumn
If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.

sampleRelHeights
If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.

sampleRelWidths
If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

list of .ggplot objects

See Also

runEmptyDrops, plotEmptyDropsScatter

Examples

```
data(scExample, package = "singleCellTK")
sce <- runEmptyDrops(inSCE = sce)
plotEmptyDropsResults(inSCE = sce)
```

---

plotEmptyDropsScatter *Plots for runEmptyDrops outputs.*

Description

A plotting function which visualizes outputs from the runEmptyDrops function stored in the colData slot of the SingleCellExperiment object via scatter plots.

Usage

```
plotEmptyDropsScatter(
  inSCE,
  sample = NULL,
  fdrCutoff = 0.01,
  defaultTheme = TRUE,
  dotSize = 0.1,
  title = NULL,
```

```
  titleSize = 18,
  xlab = NULL,
  ylab = NULL,
  axisSize = 12,
  axisLabelSize = 15,
  legendTitle = NULL,
  legendTitleSize = 12,
  legendSize = 10,
  combinePlot = "none",
  relHeights = 1,
  relWidths = 1,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runEmptyDrops. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
fdrCutoff	Numeric. Thresholds barcodes based on the FDR values from runEmptyDrops as "Empty Droplet" or "Putative Cell". Default 0.01.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.1.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 18.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 12.
axisLabelSize	Size of x/y-axis labels. Default 15.
legendTitle	Title of legend. Default NULL.
legendTitleSize	size of legend title. Default 12.
legendSize	size of legend. Default 10.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
relHeights	Relative heights of plots when combine is set. Default 1.
relWidths	Relative widths of plots when combine is set. Default 1.
samplePerColumn	If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.

sampleRelHeights
: If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.

sampleRelWidths
: If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

a ggplot object of the scatter plot.

See Also

runEmptyDrops, plotEmptyDropsResults

Examples

```
data(scExample, package = "singleCellTK")
sce <- runEmptyDrops(inSCE = sce)
plotEmptyDropsScatter(inSCE = sce)
```

---

plotFindMarkerHeatmap *Plot a heatmap to visualize the result of* runFindMarker

---

Description

This function will first reads the result saved in metadata slot, named by "findMarker" and generated by runFindMarker. Then it do the filtering on the statistics based on the input parameters and get unique genes to plot. We choose the genes that are identified as up-regulated only. As for the genes identified as up-regulated for multiple clusters, we only keep the belonging towards the one they have the highest Log2FC value. In the heatmap, there will always be a cell annotation for the cluster labeling used when finding the markers, and a feature annotation for which cluster each gene belongs to. And by default we split the heatmap by these two annotations. Additional legends can be added and the splitting can be canceled.

Usage

```
plotFindMarkerHeatmap(
  inSCE,
  orderBy = "size",
  log2fcThreshold = 1,
  fdrThreshold = 0.05,
  minClustExprPerc = 0.7,
  maxCtrlExprPerc = 0.4,
  minMeanExpr = 1,
  topN = 10,
  decreasing = TRUE,
  rowLabel = TRUE,
```

```
  rowDataName = NULL,
  colDataName = NULL,
  featureAnnotations = NULL,
  cellAnnotations = NULL,
  featureAnnotationColor = NULL,
  cellAnnotationColor = NULL,
  colSplitBy = NULL,
  rowSplitBy = "marker",
  rowDend = FALSE,
  colDend = FALSE,
  title = "Top Marker Heatmap",
  ...
)

plotMarkerDiffExp(
  inSCE,
  orderBy = "size",
  log2fcThreshold = 1,
  fdrThreshold = 0.05,
  minClustExprPerc = 0.7,
  maxCtrlExprPerc = 0.4,
  minMeanExpr = 1,
  topN = 10,
  decreasing = TRUE,
  rowDataName = NULL,
  colDataName = NULL,
  featureAnnotations = NULL,
  cellAnnotations = NULL,
  featureAnnotationColor = NULL,
  cellAnnotationColor = NULL,
  colSplitBy = NULL,
  rowSplitBy = "marker",
  rowDend = FALSE,
  colDend = FALSE,
  title = "Top Marker Heatmap",
  ...
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
orderBy	The ordering method of the clusters on the splitted heatmap. Can be chosen from "size" or "name", specified with vector of ordered unique cluster labels, or set as NULL for unsplitted heatmap. Default "size".
log2fcThreshold	Only use DEGs with the absolute values of log2FC larger than this value. Default 1
fdrThreshold	Only use DEGs with FDR value smaller than this value. Default 0.05

`minClustExprPerc`
: A numeric scalar. The minimum cutoff of the percentage of cells in the cluster of interests that expressed the marker gene. Default `0.7`.

`maxCtrlExprPerc`
: A numeric scalar. The maximum cutoff of the percentage of cells out of the cluster (control group) that expressed the marker gene. Default `0.4`.

`minMeanExpr`
: A numeric scalar. The minimum cutoff of the mean expression value of the marker in the cluster of interests. Default `1`.

`topN`
: An integer. Only to plot this number of top markers for each cluster in maximum, in terms of log2FC value. Use `NULL` to cancel the top N subscription. Default `10`.

`decreasing`
: Order the cluster decreasingly. Default `TRUE`.

`rowLabel`
: `TRUE` for displaying `rownames` of `inSCE`, a `rowData` variable to use other feature identifiers, or a vector for customized row labels. Use `FALSE` for not displaying. Default `TRUE`.

`rowDataName`
: character. The column name(s) in `rowData` that need to be added to the annotation. Default `NULL`.

`colDataName`
: character. The column name(s) in `colData` that need to be added to the annotation. Default `NULL`.

`featureAnnotations`
: `data.frame`, with `rownames` containing all the features going to be plotted. Character columns should be factors. Default `NULL`.

`cellAnnotations`
: `data.frame`, with `rownames` containing all the cells going to be plotted. Character columns should be factors. Default `NULL`.

`featureAnnotationColor`
: A named list. Customized color settings for feature labeling. Should match the entries in the `featureAnnotations` or `rowDataName`. For each entry, there should be a list/vector of colors named with categories. Default `NULL`.

`cellAnnotationColor`
: A named list. Customized color settings for cell labeling. Should match the entries in the `cellAnnotations` or `colDataName`. For each entry, there should be a list/vector of colors named with categories. Default `NULL`.

`colSplitBy`
: character vector. Do semi-heatmap based on the grouping of this(these) annotation(s). Should exist in either `colDataName` or `names(cellAnnotations)`. Default is the value of `cluster` in `runFindMarker` when `orderBy` is not `NULL`, or `NULL` otherwise.

`rowSplitBy`
: character vector. Do semi-heatmap based on the grouping of this(these) annotation(s). Should exist in either `rowDataName` or `names(featureAnnotations)`. Default `"marker"`, which indicates an auto generated annotation for this plot.

`rowDend`
: Whether to display row dendrogram. Default `FALSE`.

`colDend`
: Whether to display column dendrogram. Default `FALSE`.

`title`
: Text of the title, at the top of the heatmap. Default `"Top Marker Heatmap"`.

`...`
: Other arguments passed to `plotSCEHeatmap`.

Value

A Heatmap object

Author(s)

Yichen Wang

See Also

runFindMarker, getFindMarkerTopTable

Examples

```
data("sceBatches")
logcounts(sceBatches) <- log1p(counts(sceBatches))
sce.w <- subsetSCECols(sceBatches, colData = "batch == 'w'")
sce.w <- runFindMarker(sce.w, method = "wilcox", cluster = "cell_type")
plotFindMarkerHeatmap(sce.w)
```

---

plotMASTThresholdGenes

MAST Identify adaptive thresholds

Description

Calculate and produce a list of thresholded counts (on natural scale), thresholds, bins, densities estimated on each bin, and the original data from thresholdSCRNACountMatrix

Usage

```
plotMASTThresholdGenes(
  inSCE,
  useAssay = "logcounts",
  doPlot = TRUE,
  isLogged = TRUE,
  check_sanity = TRUE
)
```

Arguments

inSCE	SingleCellExperiment object
useAssay	character, default "logcounts"
doPlot	Logical scalar. Whether to directly plot in the plotting area. If FALSE, will return a graphical object which can be visualized with grid.draw(). Default TRUE.
isLogged	Logical scalar. Whether the assay used for the analysis is logged. If not, will do a log(assay + 1) transformation. Default TRUE.
check_sanity	Logical scalar. Whether to perform MAST's sanity check to see if the counts are logged. Default TRUE

Value

Plot the thresholding onto the plotting region if `plot == TRUE` or a graphical object if `plot == FALSE`.

Examples

```
data("mouseBrainSubsetSCE")
plotMASTThresholdGenes(mouseBrainSubsetSCE)
```

`plotPathway` *Generate violin plots for pathway analysis results*

Description

Generate violin plots for pathway analysis results

Usage

```
plotPathway(
  inSCE,
  resultName,
  geneset,
  groupBy = NULL,
  boxplot = FALSE,
  violin = TRUE,
  dots = TRUE,
  summary = "median",
  axisSize = 10,
  axisLabelSize = 10,
  dotSize = 0.5,
  transparency = 1,
  defaultTheme = TRUE,
  gridLine = FALSE,
  title = geneset,
  titleSize = NULL
)
```

Arguments

`inSCE`	Input SingleCellExperiment object. With `runGSVA()` or `runVAM()` applied in advance.
`resultName`	A single character of the name of a score matrix, which should be found in `getPathwayResultNames(inSCE)`.
`geneset`	A single character specifying the geneset of interest. Should be found in the geneSetCollection used for performing the analysis.

groupBy	Either a single character specifying a column of colData(inSCE) or a vector of equal length as the number of cells. Default NULL.
boxplot	Boolean, Whether to add a boxplot. Default FALSE.
violin	Boolean, Whether to add a violin plot. Default TRUE.
dots	Boolean, If TRUE, will plot dots for each violin plot. Default TRUE.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median", and NULL for not adding. Default "median".
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
dotSize	Size of dots. Default 0.5.
transparency	Transparency of the dots, values will be 0-1. Default 1.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
gridLine	Adds a horizontal grid line if TRUE. Will still be drawn even if defaultTheme is TRUE. Default FALSE.
title	Title of plot. Default using geneset.
titleSize	Size of the title of the plot. Default 15.

Details

runGSVA() or runVAM() should be applied in advance of using this function. Users can group the data by specifying groupby.

Value

A ggplot object for the violin plot

Examples

```
data("scExample", package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- scaterlogNormCounts(sce, assayName = "logcounts")
gs1 <- rownames(sce)[seq(10)]
gs2 <- rownames(sce)[seq(11,20)]
gs <- list("geneset1" = gs1, "geneset2" = gs2)
sce <- importGeneSetsFromList(inSCE = sce, geneSetList = gs,
                              by = "rownames")
sce <- runVAM(inSCE = sce, geneSetCollectionName = "GeneSetCollection",
              useAssay = "logcounts")
plotPathway(sce, "VAM_GeneSetCollection_CDF", "geneset1")
```

plotPCA *Plot PCA run data from its components.*

Description

Plot PCA run data from its components.

Usage

```
plotPCA(
  inSCE,
  colorBy = NULL,
  shape = NULL,
  pcX = "PC1",
  pcY = "PC2",
  reducedDimName = "PCA",
  runPCA = FALSE,
  useAssay = "logcounts"
)
```

Arguments

inSCE	Input SingleCellExperiment object.
colorBy	The variable to color clusters by
shape	Shape of the points
pcX	User choice for the first principal component
pcY	User choice for the second principal component
reducedDimName	a name to store the results of the dimension reduction coordinates obtained from this method. This is stored in the SingleCellExperiment object in the reducedDims slot. Required.
runPCA	Run PCA if the reducedDimName does not exist. the Default is FALSE.
useAssay	Indicate which assay to use. The default is "logcounts".

Value

A PCA plot

Examples

```
data("mouseBrainSubsetSCE")
plotPCA(mouseBrainSubsetSCE, colorBy = "level1class",
        reducedDimName = "PCA_counts")
```

plotRunPerCellQCResults *Plots for runPerCellQC outputs.*

Description

A wrapper function which visualizes outputs from the runPerCellQC function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotRunPerCellQCResults(
  inSCE,
  sample = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
  dots = TRUE,
  dotSize = 0.5,
  summary = "median",
  summaryTextSize = 3,
  baseSize = 15,
  axisSize = NULL,
  axisLabelSize = NULL,
  transparency = 1,
  defaultTheme = TRUE,
  titleSize = NULL,
  relHeights = 1,
  relWidths = 1,
  labelSamples = TRUE,
  plotNCols = NULL,
  plotNRows = NULL,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runPerCellQC. Required.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
groupBy	Groupings for each numeric value. Users may input a vector equal length to the number of the samples in inSCE, or can be retrieved from the colData slot. Default NULL.

combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default FALSE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default "median".
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
baseSize	The base font size for all text. Default 15. Can be overwritten by titleSize, axisSize, and axisLabelSize.
axisSize	Size of x/y-axis ticks. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
transparency	Transparency of the dots, values will be 0-1. Default 1.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
titleSize	Size of title of plot. Default NULL.
relHeights	Relative heights of plots when combine is set. Default 1.
relWidths	Relative widths of plots when combine is set. Default 1.
labelSamples	Will label sample name in title of plot if TRUE. Default TRUE.
plotNCols	Number of columns when plots are combined in a grid. Default NULL.
plotNRows	Number of rows when plots are combined in a grid. Default NULL.
samplePerColumn	If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.
sampleRelHeights	If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.
sampleRelWidths	If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

list of .ggplot objects

See Also

runPerCellQC

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runPerCellQC(sce)
plotRunPerCellQCResults(inSCE = sce)
```

---

plotScanpyDotPlot *plotScanpyDotPlot*

Description

plotScanpyDotPlot

Usage

```
plotScanpyDotPlot(
  inSCE,
  useAssay = NULL,
  features,
  groupBy,
  standardScale = NULL,
  title = "",
  vmin = NULL,
  vmax = NULL,
  colorBarTitle = "Mean expression in group"
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Assay to use for plotting. By default it will use counts assay.
features	Genes to plot. Sometimes is useful to pass a specific list of var names (e.g. genes). The var_names could be a dictionary or a list.
groupBy	The key of the observation grouping to consider.
standardScale	Whether or not to standardize the given dimension between 0 and 1, meaning for each variable or group, subtract the minimum and divide each by its maximum. Default NULL means that it doesn't perform any scaling.
title	Provide title for the figure.
vmin	The value representing the lower limit of the color scale. Values smaller than vmin are plotted with the same color as vmin. Default NULL
vmax	The value representing the upper limit of the color scale. Values larger than vmax are plotted with the same color as vmax. Default NULL
colorBarTitle	Title for the color bar.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyUMAP(sce, useReducedDim = "scanpyPCA")
markers <- c("MALAT1" ,"RPS27" ,"CST3")
plotScanpyDotPlot(sce, features = markers, groupBy = 'Scanpy_louvain_1')

## End(Not run)
```

---

plotScanpyEmbedding *plotScanpyEmbedding*

Description

plotScanpyEmbedding

Usage

```
plotScanpyEmbedding(
  inSCE,
  reducedDimName,
  useAssay = NULL,
  color = NULL,
  legend = "right margin",
  title = ""
)
```

Arguments

inSCE	Input SingleCellExperiment object.
reducedDimName	Name of reducedDims object containing embeddings. Eg. scanpyUMAP.
useAssay	Specify name of assay to use. Default is NULL, which will use scaled assay by default.
color	Keys for annotations of observations/cells or variables/genes.
legend	Location of legend, either 'on data', 'right margin' or a valid keyword for the loc parameter of Legend.
title	Provide title for panels either as string or list of strings

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyUMAP(sce, useReducedDim = "scanpyPCA")
plotScanpyEmbedding(sce, reducedDimName = "scanpyUMAP", color = 'Scanpy_louvain_1')

## End(Not run)
```

---

plotScanpyHeatmap *plotScanpyHeatmap*

Description

plotScanpyHeatmap

Usage

```
plotScanpyHeatmap(
  inSCE,
  useAssay = NULL,
  features,
  groupBy,
  standardScale = "var",
  vmin = NULL,
  vmax = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Assay to use for plotting. By default it will use counts assay.
features	Genes to plot. Sometimes is useful to pass a specific list of var names (e.g. genes). The var_names could be a dictionary or a list.
groupBy	The key of the observation grouping to consider.
standardScale	Whether or not to standardize the given dimension between 0 and 1, meaning for each variable or group, subtract the minimum and divide each by its maximum. Default NULL means that it doesn't perform any scaling.

vmin	The value representing the lower limit of the color scale. Values smaller than vmin are plotted with the same color as vmin. Default NULL
vmax	The value representing the upper limit of the color scale. Values larger than vmax are plotted with the same color as vmax. Default NULL

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyUMAP(sce, useReducedDim = "scanpyPCA")
markers <- c("MALAT1" ,"RPS27" ,"CST3")
plotScanpyHeatmap(sce, features = markers, groupBy = 'Scanpy_louvain_1')

## End(Not run)
```

---

plotScanpyHVG *plotScanpyHVG*

Description

plotScanpyHVG

Usage

```
plotScanpyHVG(inSCE, log = FALSE)
```

Arguments

inSCE	Input SingleCellExperiment object.
log	Plot on logarithmic axes. Default FALSE.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
plotScanpyHVG(sce)

## End(Not run)
```

`plotScanpyMarkerGenes` *plotScanpyMarkerGenes*

Description

plotScanpyMarkerGenes

Usage

```
plotScanpyMarkerGenes(
  inSCE,
  groups = NULL,
  nGenes = 10,
  nCols = 4,
  sharey = FALSE
)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`groups`	The groups for which to show the gene ranking. Default `NULL` means that all groups will be considered.
`nGenes`	Number of genes to show. Default `10`
`nCols`	Number of panels shown per row. Default `4`
`sharey`	Controls if the y-axis of each panels should be shared. Default `FALSE` allows each panel to have its own y-axis range.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
```

```
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyFindMarkers(sce, colDataName = "Scanpy_louvain_1" )
plotScanpyMarkerGenes(sce, groups = '0')

## End(Not run)
```

---

plotScanpyMarkerGenesDotPlot *plotScanpyMarkerGenesDotPlot*

Description

plotScanpyMarkerGenesDotPlot

Usage

```
plotScanpyMarkerGenesDotPlot(
  inSCE,
  groups = NULL,
  nGenes = 10,
  groupBy,
  log2fcThreshold = NULL,
  parameters = "logfoldchanges",
  standardScale = NULL,
  features = NULL,
  title = "",
  vmin = NULL,
  vmax = NULL,
  colorBarTitle = "log fold change"
)
```

Arguments

inSCE	Input `SingleCellExperiment` object.
groups	The groups for which to show the gene ranking. Default `NULL` means that all groups will be considered.
nGenes	Number of genes to show. Default `10`
groupBy	The key of the observation grouping to consider. By default, the groupby is chosen from the rank genes groups parameter.
log2fcThreshold	Only output DEGs with the absolute values of log2FC larger than this value. Default `NULL`.
parameters	The options for marker genes results to plot are: 'scores', 'logfoldchanges', 'pvals', 'pvals_adj', 'log10_pvals', 'log10_pvals_adj'. If NULL provided then it uses mean gene value to plot.

standardScale	Whether or not to standardize the given dimension between 0 and 1, meaning for each variable or group, subtract the minimum and divide each by its maximum. Default NULL means that it doesn't perform any scaling.
features	Genes to plot. Sometimes is useful to pass a specific list of var names (e.g. genes) to check their fold changes or p-values, instead of the top/bottom genes. The gene names could be a dictionary or a list. Default NULL
title	Provide title for the figure.
vmin	The value representing the lower limit of the color scale. Values smaller than vmin are plotted with the same color as vmin. Default NULL
vmax	The value representing the upper limit of the color scale. Values larger than vmax are plotted with the same color as vmax. Default NULL
colorBarTitle	Title for the color bar.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyFindMarkers(sce, colDataName = "Scanpy_louvain_1" )
plotScanpyMarkerGenesDotPlot(sce, groupBy = 'Scanpy_louvain_1')

## End(Not run)
```

---

plotScanpyMarkerGenesHeatmap

plotScanpyMarkerGenesHeatmap

Description

plotScanpyMarkerGenesHeatmap

Usage

```
plotScanpyMarkerGenesHeatmap(
  inSCE,
  groups = NULL,
  groupBy,
  nGenes = 10,
  features = NULL,
  log2fcThreshold = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object.
groups	The groups for which to show the gene ranking. Default NULL means that all groups will be considered.
groupBy	The key of the observation grouping to consider. By default, the groupby is chosen from the rank genes groups parameter.
nGenes	Number of genes to show. Default 10
features	Genes to plot. Sometimes is useful to pass a specific list of var names (e.g. genes). The var_names could be a dictionary or a list.
log2fcThreshold	Only output DEGs with the absolute values of log2FC larger than this value. Default NULL.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyFindMarkers(sce, colDataName = "Scanpy_louvain_1" )
plotScanpyMarkerGenesHeatmap(sce, groupBy = 'Scanpy_louvain_1')

## End(Not run)
```

---

plotScanpyMarkerGenesMatrixPlot

plotScanpyMarkerGenesMatrixPlot

Description

plotScanpyMarkerGenesMatrixPlot

Usage

```
plotScanpyMarkerGenesMatrixPlot(
  inSCE,
  groups = NULL,
  nGenes = 10,
  groupBy,
```

```
  log2fcThreshold = NULL,
  parameters = "logfoldchanges",
  standardScale = "var",
  features = NULL,
  title = "",
  vmin = NULL,
  vmax = NULL,
  colorBarTitle = "log fold change"
)
```

Arguments

inSCE	Input SingleCellExperiment object.
groups	The groups for which to show the gene ranking. Default NULL means that all groups will be considered.
nGenes	Number of genes to show. Default 10
groupBy	The key of the observation grouping to consider. By default, the groupby is chosen from the rank genes groups parameter.
log2fcThreshold	Only output DEGs with the absolute values of log2FC larger than this value. Default NULL.
parameters	The options for marker genes results to plot are: 'scores', 'logfoldchanges', 'pvals', 'pvals_adj', 'log10_pvals', 'log10_pvals_adj'. If NULL provided then it uses mean gene value to plot.
standardScale	Whether or not to standardize the given dimension between 0 and 1, meaning for each variable or group, subtract the minimum and divide each by its maximum. Default NULL means that it doesn't perform any scaling.
features	Genes to plot. Sometimes is useful to pass a specific list of var names (e.g. genes) to check their fold changes or p-values, instead of the top/bottom genes. The var_names could be a dictionary or a list. Default NULL
title	Provide title for the figure.
vmin	The value representing the lower limit of the color scale. Values smaller than vmin are plotted with the same color as vmin. Default NULL
vmax	The value representing the upper limit of the color scale. Values larger than vmax are plotted with the same color as vmax. Default NULL
colorBarTitle	Title for the color bar.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
```

```
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyFindMarkers(sce, colDataName = "Scanpy_louvain_1" )
plotScanpyMarkerGenesMatrixPlot(sce, groupBy = 'Scanpy_louvain_1')

## End(Not run)
```

`plotScanpyMarkerGenesViolin` *plotScanpyMarkerGenesViolin*

Description

plotScanpyMarkerGenesViolin

Usage

```
plotScanpyMarkerGenesViolin(inSCE, groups = NULL, features = NULL, nGenes = 10)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`groups`	The groups for which to show the gene ranking. Default `NULL` means that all groups will be considered.
`features`	List of genes to plot. Is only useful if interested in a custom gene list
`nGenes`	Number of genes to show. Default `10`

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyFindMarkers(sce, colDataName = "Scanpy_louvain_1" )
plotScanpyMarkerGenesViolin(sce, groups = '0')

## End(Not run)
```

plotScanpyMatrixPlot *plotScanpyMatrixPlot*

Description

plotScanpyMatrixPlot

Usage

```
plotScanpyMatrixPlot(
  inSCE,
  useAssay = NULL,
  features,
  groupBy,
  standardScale = NULL,
  title = "",
  vmin = NULL,
  vmax = NULL,
  colorBarTitle = "Mean expression in group"
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Assay to use for plotting. By default it will use counts assay.
features	Genes to plot. Sometimes is useful to pass a specific list of var names (e.g. genes). The var_names could be a dictionary or a list.
groupBy	The key of the observation grouping to consider.
standardScale	Whether or not to standardize the given dimension between 0 and 1, meaning for each variable or group, subtract the minimum and divide each by its maximum. Default NULL means that it doesn't perform any scaling.
title	Provide title for the figure.
vmin	The value representing the lower limit of the color scale. Values smaller than vmin are plotted with the same color as vmin. Default NULL
vmax	The value representing the upper limit of the color scale. Values larger than vmax are plotted with the same color as vmax. Default NULL
colorBarTitle	Title for the color bar.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyUMAP(sce, useReducedDim = "scanpyPCA")
markers <- c("MALAT1" ,"RPS27" ,"CST3")
plotScanpyMatrixPlot(sce, features = markers, groupBy = 'Scanpy_louvain_1')

## End(Not run)
```

`plotScanpyPCA` *plotScanpyPCA*

Description

plotScanpyPCA

Usage

```
plotScanpyPCA(
  inSCE,
  reducedDimName = "scanpyPCA",
  color = NULL,
  title = "",
  legend = "right margin"
)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`reducedDimName`	Name of new reducedDims object containing Scanpy PCA.
`color`	Keys for annotations of observations/cells or variables/genes.
`title`	Provide title for panels either as string or list of strings
`legend`	Location of legend, either 'on data', 'right margin' or a valid keyword for the loc parameter of Legend.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
plotScanpyPCA(sce)

## End(Not run)
```

---

plotScanpyPCAGeneRanking

plotScanpyPCAGeneRanking

Description

plotScanpyPCAGeneRanking

Usage

```
plotScanpyPCAGeneRanking(inSCE, PC_comp = "1,2,3", includeLowest = TRUE)
```

Arguments

inSCE	Input SingleCellExperiment object.
PC_comp	For example, '1,2,3' means [1, 2, 3], first, second, third principal component.
includeLowest	Whether to show the variables with both highest and lowest loadings. Default TRUE

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
plotScanpyPCAGeneRanking(sce)

## End(Not run)
```

plotScanpyPCAVariance *plotScanpyPCAVariance*

Description

plotScanpyPCAVariance

Usage

```
plotScanpyPCAVariance(inSCE, nPCs = 50, log = FALSE)
```

Arguments

inSCE	Input SingleCellExperiment object.
nPCs	Number of PCs to show. Default 50.
log	Plot on logarithmic scale. Default FALSE

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
plotScanpyPCAVariance(sce)

## End(Not run)
```

plotScanpyViolin *plotScanpyViolin*

Description

plotScanpyViolin

Usage

```
plotScanpyViolin(
  inSCE,
  useAssay = NULL,
  features,
  groupBy,
  xlabel = "",
  ylabel = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Assay to use for plotting. By default it will use counts assay.
features	Genes to plot. Sometimes is useful to pass a specific list of var names (e.g. genes). The var_names could be a dictionary or a list.
groupBy	The key of the observation grouping to consider.
xlabel	Label of the x axis. Defaults to groupBy.
ylabel	Label of the y axis. If NULL and groupBy is NULL, defaults to 'value'. If NULL and groupBy is not NULL, defaults to features.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyUMAP(sce, useReducedDim = "scanpyPCA")
markers <- c("MALAT1" ,"RPS27" ,"CST3")
plotScanpyViolin(sce, features = markers, groupBy = "Scanpy_louvain_1")

## End(Not run)
```

---

plotScDblFinderResults

Plots for runScDblFinder outputs.

Description

A wrapper function which visualizes outputs from the runScDblFinder function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotScDblFinderResults(
  inSCE,
  sample = NULL,
  shape = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
  dots = TRUE,
  reducedDimName = "UMAP",
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  summary = "median",
  summaryTextSize = 3,
  transparency = 1,
  baseSize = 15,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL,
  relHeights = 1,
  relWidths = c(1, 1, 1),
  plotNCols = NULL,
  plotNRows = NULL,
  labelSamples = TRUE,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runScDblFinder. Required.

sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
shape	If provided, add shapes based on the value. Default NULL.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in inSCE, or can be retrieved from the colData slot. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
reducedDimName	Saved dimension reduction name in inSCE. Default "UMAP".
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Similar to dim1. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into bin groups. If more than one value, will bin numeric values using values as a cut point. Default NULL.
binLabel	Character vector. Labels for the bins created by bin. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
transparency	Transparency of the dots, values will be 0-1. Default 1.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
titleSize	Size of title of plot. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
axisSize	Size of x/y-axis ticks. Default NULL.
legendSize	size of legend. Default NULL.
legendTitleSize	size of legend title. Default NULL.
relHeights	Relative heights of plots when combine is set. Default 1.
relWidths	Relative widths of plots when combine is set. Default c(1, 1, 1).

plotNCols Number of columns when plots are combined in a grid. Default NULL.

plotNRows Number of rows when plots are combined in a grid. Default NULL.

labelSamples Will label sample name in title of plot if TRUE. Default TRUE.

samplePerColumn
If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.

sampleRelHeights
If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.

sampleRelWidths
If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

list of .ggplot objects

See Also

runScDblFinder

Examples

```
data(scExample, package="singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runQuickUMAP(sce)
sce <- runScDblFinder(sce)
plotScDblFinderResults(inSCE = sce, reducedDimName = "UMAP")
```

---

plotScdsHybridResults *Plots for runCxdsBcdsHybrid outputs.*

Description

A wrapper function which visualizes outputs from the runCxdsBcdsHybrid function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotScdsHybridResults(
  inSCE,
  sample = NULL,
  shape = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
```

```
  dots = TRUE,
  reducedDimName = "UMAP",
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  summary = "median",
  summaryTextSize = 3,
  transparency = 1,
  baseSize = 15,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL,
  relHeights = 1,
  relWidths = c(1, 1, 1),
  plotNCols = NULL,
  plotNRows = NULL,
  labelSamples = TRUE,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runCxdsBcdsHybrid. Required.
sample	Character vector. Indicates which sample each cell belongs to. Default NULL.
shape	If provided, add shapes based on the value.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
reducedDimName	Saved dimension reduction name in the SingleCellExperiment object. Required.
xlab	Character vector. Label for x-axis. Default NULL.

ylab	Character vector. Label for y-axis. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into the 'bin' groups. If more than one value, will bin numeric values using values as a cut point.
binLabel	Character vector. Labels for the bins created by the 'bin' parameter. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
transparency	Transparency of the dots, values will be 0-1. Default 1.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
titleSize	Size of title of plot. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
axisSize	Size of x/y-axis ticks. Default NULL.
legendSize	size of legend. Default NULL.
legendTitleSize	size of legend title. Default NULL.
relHeights	Relative heights of plots when combine is set.
relWidths	Relative widths of plots when combine is set.
plotNCols	Number of columns when plots are combined in a grid.
plotNRows	Number of rows when plots are combined in a grid.
labelSamples	Will label sample name in title of plot if TRUE. Default TRUE.
samplePerColumn	If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.
sampleRelHeights	If there are multiple samples and combining by "all", the relative heights for each plot.
sampleRelWidths	If there are multiple samples and combining by "all", the relative widths for each plot.

Value

list of .ggplot objects

Examples

```
data(scExample, package="singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runQuickUMAP(sce)
sce <- runCxdsBcdsHybrid(sce)
plotScdsHybridResults(inSCE=sce, reducedDimName="UMAP")
```

---

plotSCEBarAssayData *Bar plot of assay data.*

Description

Visualizes values stored in the assay slot of a SingleCellExperiment object via a bar plot.

Usage

```
plotSCEBarAssayData(
  inSCE,
  feature,
  sample = NULL,
  useAssay = "counts",
  featureLocation = NULL,
  featureDisplay = NULL,
  groupBy = NULL,
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
  axisLabelSize = 10,
  dotSize = 0.1,
  transparency = 1,
  defaultTheme = TRUE,
  gridLine = FALSE,
  summary = NULL,
  title = NULL,
  titleSize = NULL,
  combinePlot = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
feature	Name of feature stored in assay of SingleCellExperiment object.

sample	Character vector. Indicates which sample each cell belongs to.
useAssay	Indicate which assay to use. Default "counts".
featureLocation	Indicates which column name of rowData to query gene.
featureDisplay	Indicates which column name of rowData to use to display feature for visualization.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
dotSize	Size of dots. Default 0.1.
transparency	Transparency of the dots, values will be 0-1. Default 1.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
gridLine	Adds a horizontal grid line if TRUE. Will still be drawn even if defaultTheme is TRUE. Default FALSE.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
combinePlot	Boolean. If multiple plots are generated (multiple samples, etc.), will combined plots using ‘cowplot::plot_grid‘. Default TRUE.

Value

a ggplot of the barplot of assay data.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEBarAssayData(
  inSCE = mouseBrainSubsetSCE,
  feature = "Apoe", groupBy = "sex"
)
```

plotSCEBarColData *Bar plot of colData.*

Description

Visualizes values stored in the colData slot of a SingleCellExperiment object via a bar plot.

Usage

```
plotSCEBarColData(
  inSCE,
  coldata,
  sample = NULL,
  groupBy = NULL,
  dots = TRUE,
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
  axisLabelSize = 10,
  dotSize = 0.1,
  transparency = 1,
  defaultTheme = TRUE,
  gridLine = FALSE,
  summary = NULL,
  title = NULL,
  titleSize = NULL,
  combinePlot = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
coldata	colData value that will be plotted.
sample	Character vector. Indicates which sample each cell belongs to.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
dotSize	Size of dots. Default 0.1.

transparency	Transparency of the dots, values will be 0-1. Default 1.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
gridLine	Adds a horizontal grid line if TRUE. Will still be drawn even if defaultTheme is TRUE. Default FALSE.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
combinePlot	Boolean. If multiple plots are generated (multiple samples, etc.), will combined plots using ‘cowplot::plot_grid‘. Default TRUE.

Value

a ggplot of the barplot of coldata.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEBarColData(
  inSCE = mouseBrainSubsetSCE,
  coldata = "age", groupBy = "sex"
)
```

---

plotSCEBatchFeatureMean

Plot mean feature value in each batch of a SingleCellExperiment object

---

Description

Plot mean feature value in each batch of a SingleCellExperiment object

Usage

```
plotSCEBatchFeatureMean(
  inSCE,
  useAssay = NULL,
  useReddim = NULL,
  useAltExp = NULL,
  batch = "batch",
  xlab = "batch",
  ylab = "Feature Mean",
  ...
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
useAssay	A single character. The name of the assay that stores the value to plot. For useReddim and useAltExp also. Default NULL.
useReddim	A single character. The name of the dimension reduced matrix that stores the value to plot. Default NULL.
useAltExp	A single character. The name of the alternative experiment that stores an assay of the value to plot. Default NULL.
batch	A single character. The name of batch annotation column in colData(inSCE). Default "batch".
xlab	label for x-axis. Default "batch".
ylab	label for y-axis. Default "Feature Mean".
...	Additional arguments passed to .ggViolin.

Value

ggplot

Examples

```
data('sceBatches', package = 'singleCellTK')
plotSCEBatchFeatureMean(sceBatches, useAssay = "counts")
```

---

plotSCEDensity *Density plot of any data stored in the SingleCellExperiment object.*

Description

Visualizes values stored in any slot of a SingleCellExperiment object via a densityn plot.

Usage

```
plotSCEDensity(
  inSCE,
  slotName,
  itemName,
  sample = NULL,
  feature = NULL,
  dimension = NULL,
  groupBy = NULL,
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
  axisLabelSize = 10,
  defaultTheme = TRUE,
```

```
  title = NULL,
  titleSize = 18,
  cutoff = NULL,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
slotName	Desired slot of SingleCellExperiment used for plotting. Possible options: "assays", "colData", "metadata", "reducedDims". Required.
itemName	Desired vector within the slot used for plotting. Required.
sample	Character vector. Indicates which sample each cell belongs to.
feature	Desired name of feature stored in assay of SingleCellExperiment object. Only used when "assays" slotName is selected. Default NULL.
dimension	Desired dimension stored in the specified reducedDims. Either an integer which indicates the column or a character vector specifies column name. By default, the 1st dimension/column will be used. Only used when "reducedDims" slotName is selected. Default NULL.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
cutoff	Numeric value. The plot will be annotated with a vertical line if set. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot object of the density plot.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEDensity(
  inSCE = mouseBrainSubsetSCE, slotName = "assays",
  itemName = "counts", feature = "Apoe", groupBy = "sex"
)
```

`plotSCEDensityAssayData` *Density plot of assay data.*

Description

Visualizes values stored in the assay slot of a SingleCellExperiment object via a density plot.

Usage

```
plotSCEDensityAssayData(
  inSCE,
  feature,
  sample = NULL,
  useAssay = "counts",
  featureLocation = NULL,
  featureDisplay = NULL,
  groupBy = NULL,
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
  axisLabelSize = 10,
  defaultTheme = TRUE,
  cutoff = NULL,
  title = NULL,
  titleSize = 18,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

`inSCE`	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
`feature`	Name of feature stored in assay of SingleCellExperiment object.
`sample`	Character vector. Indicates which sample each cell belongs to.
`useAssay`	Indicate which assay to use. Default "counts".
`featureLocation`	Indicates which column name of rowData to query gene.

featureDisplay	Indicates which column name of rowData to use to display feature for visualization.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
cutoff	Numeric value. The plot will be annotated with a vertical line if set. Default NULL.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the density plot of assay data.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEDensityAssayData(
  inSCE = mouseBrainSubsetSCE,
  feature = "Apoe"
)
```

---

plotSCEDensityColData *Density plot of colData.*

Description

Visualizes values stored in the colData slot of a SingleCellExperiment object via a density plot.

Usage

```
plotSCEDensityColData(
  inSCE,
  coldata,
  sample = NULL,
  groupBy = NULL,
  xlab = NULL,
  ylab = NULL,
  baseSize = 12,
  axisSize = NULL,
  axisLabelSize = NULL,
  defaultTheme = TRUE,
  title = NULL,
  titleSize = 18,
  cutoff = NULL,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
coldata	colData value that will be plotted.
sample	Character vector. Indicates which sample each cell belongs to.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
axisSize	Size of x/y-axis ticks. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
cutoff	Numeric value. The plot will be annotated with a vertical line if set. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the density plot of colData.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEDensityColData(
  inSCE = mouseBrainSubsetSCE,
  coldata = "age", groupBy = "sex"
)
```

`plotSCEDimReduceColData` *Dimension reduction plot tool for colData*

Description

Plot results of reduced dimensions data and colors by annotation data stored in the colData slot.

Usage

```
plotSCEDimReduceColData(
  inSCE,
  colorBy,
  reducedDimName,
  sample = NULL,
  groupBy = NULL,
  conditionClass = NULL,
  shape = NULL,
  xlab = NULL,
  ylab = NULL,
  baseSize = 12,
  axisSize = NULL,
  axisLabelSize = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  dotSize = 0.1,
  transparency = 1,
  colorScale = NULL,
  colorLow = "white",
  colorMid = "gray",
  colorHigh = "blue",
  defaultTheme = TRUE,
  title = NULL,
  titleSize = 15,
```

```
  labelClusters = TRUE,
  clusterLabelSize = 3.5,
  legendTitle = NULL,
  legendTitleSize = NULL,
  legendSize = NULL,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
colorBy	Color by a condition(any column of the annotation data). Required.
reducedDimName	Saved dimension reduction matrix name in the SingleCellExperiment object. Required.
sample	Character vector. Indicates which sample each cell belongs to.
groupBy	Group by a condition(any column of the annotation data). Default NULL.
conditionClass	Class of the annotation data used in colorBy. Options are NULL, "factor" or "numeric". If NULL, class will default to the original class. Default NULL.
shape	Add shapes to each condition.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
axisSize	Size of x/y-axis ticks. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into the 'bin' groups. If more than one value, will bin numeric values using values as a cut point.
binLabel	Character vector. Labels for the bins created by the 'bin' parameter. Default NULL.
dotSize	Size of dots. Default 0.1.
transparency	Transparency of the dots, values will be 0-1. Default 1.
colorScale	Vector. Needs to be same length as the number of unique levels of colorBy. Will be used only if conditionClass = "factor" or "character". Default NULL.

colorLow	Character. A color available from 'colors()'. The color will be used to signify the lowest values on the scale. Default 'white'.
colorMid	Character. A color available from 'colors()'. The color will be used to signify the midpoint on the scale. Default 'gray'.
colorHigh	Character. A color available from 'colors()'. The color will be used to signify the highest values on the scale. Default 'blue'.
defaultTheme	adds grid to plot when TRUE. Default TRUE.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
labelClusters	Logical. Whether the cluster labels are plotted.
clusterLabelSize	Numeric. Determines the size of cluster label when 'labelClusters' is set to TRUE. Default 3.5.
legendTitle	title of legend. Default NULL.
legendTitleSize	size of legend title. Default 12.
legendSize	size of legend. Default NULL. Default FALSE.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the reduced dimension plot of coldata.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEDimReduceColData(
  inSCE = mouseBrainSubsetSCE, colorBy = "tissue",
  shape = NULL, conditionClass = "factor",
  reducedDimName = "TSNE_counts",
  xlab = "tSNE1", ylab = "tSNE2", labelClusters = TRUE
)

plotSCEDimReduceColData(
  inSCE = mouseBrainSubsetSCE, colorBy = "age",
  shape = NULL, conditionClass = "numeric",
  reducedDimName = "TSNE_counts", bin = c(-Inf, 20, 25, +Inf),
  xlab = "tSNE1", ylab = "tSNE2", labelClusters = FALSE
)
```

plotSCEDimReduceFeatures

Dimension reduction plot tool for assay data

Description

Plot results of reduced dimensions data and colors by feature data stored in the assays slot.

Usage

```
plotSCEDimReduceFeatures(
  inSCE,
  feature,
  reducedDimName,
  sample = NULL,
  featureLocation = NULL,
  featureDisplay = NULL,
  shape = NULL,
  useAssay = "logcounts",
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
  axisLabelSize = 10,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  dotSize = 0.1,
  transparency = 1,
  colorLow = "white",
  colorMid = "gray",
  colorHigh = "blue",
  defaultTheme = TRUE,
  title = NULL,
  titleSize = 15,
  legendTitle = NULL,
  legendSize = 10,
  legendTitleSize = 12,
  groupBy = NULL,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.

feature	Name of feature stored in assay of SingleCellExperiment object.
reducedDimName	saved dimension reduction name in the SingleCellExperiment object. Required.
sample	Character vector. Indicates which sample each cell belongs to.
featureLocation	Indicates which column name of rowData to query gene.
featureDisplay	Indicates which column name of rowData to use to display feature for visualization.
shape	add shapes to each condition. Default NULL.
useAssay	Indicate which assay to use. The default is "logcounts"
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into the 'bin' groups. If more than one value, will bin numeric values using values as a cut point.
binLabel	Character vector. Labels for the bins created by the 'bin' parameter. Default NULL.
dotSize	Size of dots. Default 0.1.
transparency	Transparency of the dots, values will be 0-1. Default 1.
colorLow	Character. A color available from 'colors()'. The color will be used to signify the lowest values on the scale. Default 'white'.
colorMid	Character. A color available from 'colors()'. The color will be used to signify the midpoint on the scale. Default 'gray'.
colorHigh	Character. A color available from 'colors()'. The color will be used to signify the highest values on the scale. Default 'blue'.
defaultTheme	adds grid to plot when TRUE. Default TRUE.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
legendTitle	title of legend. Default NULL.
legendSize	size of legend. Default 10.
legendTitleSize	size of legend title. Default 12.
groupBy	Facet wrap the scatterplot based on value. Default NULL.

combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the reduced dimension plot of feature data.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEDimReduceFeatures(
  inSCE = mouseBrainSubsetSCE, feature = "Apoe",
  shape = NULL, reducedDimName = "TSNE_counts",
  useAssay = "counts", xlab = "tSNE1", ylab = "tSNE2"
)
```

---

plotSCEHeatmap *Plot heatmap of using data stored in SingleCellExperiment Object*

Description

Plot heatmap of using data stored in SingleCellExperiment Object

Usage

```
plotSCEHeatmap(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  doLog = FALSE,
  featureIndex = NULL,
  cellIndex = NULL,
  scale = TRUE,
  trim = c(-2, 2),
  featureIndexBy = "rownames",
  cellIndexBy = "rownames",
  rowDataName = NULL,
  colDataName = NULL,
  aggregateRow = NULL,
  aggregateCol = NULL,
  featureAnnotations = NULL,
  cellAnnotations = NULL,
  featureAnnotationColor = NULL,
  cellAnnotationColor = NULL,
  palette = c("ggplot", "celda", "random"),
```

```
  rowSplitBy = NULL,
  colSplitBy = NULL,
  rowLabel = FALSE,
  colLabel = FALSE,
  rowLabelSize = 6,
  colLabelSize = 6,
  rowDend = TRUE,
  colDend = TRUE,
  title = NULL,
  rowTitle = "Features",
  colTitle = "Cells",
  rowGap = grid::unit(0, "mm"),
  colGap = grid::unit(0, "mm"),
  border = FALSE,
  colorScheme = NULL,
  ...
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
useAssay	character. A string indicating the assay name that provides the expression level to plot. Only for plotSCEHeatmap.
useReducedDim	character. A string indicating the reducedDim name that provides the expression level to plot. Only for plotSCEDimReduceHeatmap.
doLog	Logical scalar. Whether to do log(assay + 1) transformation on the assay indicated by useAssay. Default FALSE.
featureIndex	A vector that can subset the input SCE object by rows (features). Alternatively, it can be a vector identifying features in another feature list indicated by featureIndexBy. Default NULL.
cellIndex	A vector that can subset the input SCE object by columns (cells). Alternatively, it can be a vector identifying cells in another cell list indicated by featureIndexBy. Default NULL.
scale	Whether to perform z-score scaling on each row. Default TRUE.
trim	A 2-element numeric vector. Values outside of this range will be trimmed to their nearst bound. Default c(-2, 2)
featureIndexBy	A single character specifying a column name of rowData(inSCE), or a vector of the same length as nrow(inSCE), where we search for the non-rowname feature indices. Not applicable for plotSCEDimReduceHeatmap. Default "rownames".
cellIndexBy	A single character specifying a column name of colData(inSCE), or a vector of the same length as ncol(inSCE), where we search for the non-rowname cell indices. Default "rownames".
rowDataName	character. The column name(s) in rowData that need to be added to the annotation. Not applicable for plotSCEDimReduceHeatmap. Default NULL.
colDataName	character. The column name(s) in colData that need to be added to the annotation. Default NULL.

aggregateRow Feature variable for aggregating the heatmap by row. Can be a vector or a rowData column name for feature variable. Multiple variables are allowed. Default NULL.

aggregateCol Cell variable for aggregating the heatmap by column. Can be a vector or a colData column name for cell variable. Multiple variables are allowed. Default NULL.

featureAnnotations
data.frame, with rownames containing all the features going to be plotted. Character columns should be factors. Default NULL.

cellAnnotations
data.frame, with rownames containing all the cells going to be plotted. Character columns should be factors. Default NULL.

featureAnnotationColor
A named list. Customized color settings for feature labeling. Should match the entries in the featureAnnotations or rowDataName. For each entry, there should be a list/vector of colors named with categories. Default NULL.

cellAnnotationColor
A named list. Customized color settings for cell labeling. Should match the entries in the cellAnnotations or colDataName. For each entry, there should be a list/vector of colors named with categories. Default NULL.

palette Choose from "ggplot", "celda" or "random" to generate unique category colors.

rowSplitBy character. Do semi-heatmap based on the grouping of this(these) annotation(s). Should exist in either rowDataName or names(featureAnnotations). Default NULL.

colSplitBy character. Do semi-heatmap based on the grouping of this(these) annotation(s). Should exist in either colDataName or names(cellAnnotations). Default NULL.

rowLabel Use a logical for whether to display all the feature names, a single character to display a column of rowData(inSCE) annotation, a vector of the same length as full/subset nrow(inSCE) to display customized info. Default FALSE.

colLabel Use a logical for whether to display all the cell names, a single character to display a column of colData(inSCE) annotation, a vector of the same length as full/subset ncol(inSCE) to display customized info. Default FALSE.

rowLabelSize A number for the font size of feature names. Default 8

colLabelSize A number for the font size of cell names. Default 8

rowDend Whether to display row dendrogram. Default TRUE.

colDend Whether to display column dendrogram. Default TRUE.

title The main title of the whole plot. Default NULL.

rowTitle The subtitle for the rows. Default "Genes".

colTitle The subtitle for the columns. Default "Cells".

rowGap A numeric value or a unit object. For the gap size between rows of the splitted heatmap. Default grid::unit(0, 'mm').

colGap	A numeric value or a unit object. For the gap size between columns of the splitted heatmap. Default grid::unit(0, 'mm').
border	A logical scalar. Whether to show the border of the heatmap or splitted heatmaps. Default TRUE.
colorScheme	function. A function that generates color code by giving a value. Can be generated by colorRamp2. Default NULL.
...	Other arguments passed to Heatmap.

Value

A ggplot object.

Author(s)

Yichen Wang

Examples

```
data(scExample, package = "singleCellTK")
plotSCEHeatmap(sce[1:3,1:3], useAssay = "counts")
```

---

plotSCEScatter *Dimension reduction plot tool for all types of data*

Description

Plot results of reduced dimensions data of counts stored in any slot in the SingleCellExperiment object.

Usage

```
plotSCEScatter(
  inSCE,
  annotation,
  reducedDimName = NULL,
  slot = NULL,
  sample = NULL,
  feature = NULL,
  groupBy = NULL,
  shape = NULL,
  conditionClass = NULL,
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
  axisLabelSize = 10,
  dim1 = NULL,
  dim2 = NULL,
```

```
  bin = NULL,
  binLabel = NULL,
  dotSize = 0.1,
  transparency = 1,
  colorLow = "white",
  colorMid = "gray",
  colorHigh = "blue",
  defaultTheme = TRUE,
  title = NULL,
  titleSize = 15,
  labelClusters = TRUE,
  legendTitle = NULL,
  legendTitleSize = 12,
  legendSize = 10,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
annotation	Desired vector within the slot used for plotting. Default NULL.
reducedDimName	saved dimension reduction name in the SingleCellExperiment object.
slot	Desired slot of SingleCellExperiment used for plotting. Possible options: "assays", "colData", "metadata", "reducedDims". Default NULL.
sample	Character vector. Indicates which sample each cell belongs to.
feature	name of feature stored in assay of SingleCellExperiment object. Will be used only if "assays" slot is chosen. Default NULL.
groupBy	Group by a condition(any column of the annotation data). Default NULL.
shape	add shapes to each condition.
conditionClass	class of the annotation data used in colorBy. Options are NULL, "factor" or "numeric". If NULL, class will default to the original class. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.

bin	Numeric vector. If single value, will divide the numeric values into the 'bin' groups. If more than one value, will bin numeric values using values as a cut point.
binLabel	Character vector. Labels for the bins created by the 'bin' parameter. Default NULL.
dotSize	Size of dots. Default 0.1.
transparency	Transparency of the dots, values will be 0-1. Default 1.
colorLow	Character. A color available from 'colors()'. The color will be used to signify the lowest values on the scale. Default 'white'.
colorMid	Character. A color available from 'colors()'. The color will be used to signify the midpoint on the scale. Default 'gray'.
colorHigh	Character. A color available from 'colors()'. The color will be used to signify the highest values on the scale. Default 'blue'.
defaultTheme	adds grid to plot when TRUE. Default TRUE.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
labelClusters	Logical. Whether the cluster labels are plotted.
legendTitle	title of legend. Default NULL.
legendTitleSize	size of legend title. Default 12.
legendSize	size of legend. Default 10.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the reduced dimensions.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEScatter(
  inSCE = mouseBrainSubsetSCE, legendTitle = NULL,
  slot = "assays", annotation = "counts", feature = "Apoe",
  reducedDimName = "TSNE_counts", labelClusters = FALSE
)
```

plotSCEViolin *Violin plot of any data stored in the SingleCellExperiment object.*

Description

Visualizes values stored in any slot of a SingleCellExperiment object via a violin plot.

Usage

```
plotSCEViolin(
  inSCE,
  slotName,
  itemName,
  feature = NULL,
  sample = NULL,
  dimension = NULL,
  groupBy = NULL,
  violin = TRUE,
  boxplot = TRUE,
  dots = TRUE,
  plotOrder = NULL,
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
  axisLabelSize = 10,
  dotSize = 0.1,
  transparency = 1,
  defaultTheme = TRUE,
  gridLine = FALSE,
  summary = NULL,
  title = NULL,
  titleSize = NULL,
  hcutoff = NULL,
  hcolor = "red",
  hsize = 1,
  hlinetype = 1,
  vcutoff = NULL,
  vcolor = "red",
  vsize = 1,
  vlinetype = 1,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.

slotName	Desired slot of SingleCellExperiment used for plotting. Possible options: "assays", "colData", "metadata", "reducedDims". Required.
itemName	Desired vector within the slot used for plotting. Required.
feature	Desired name of feature stored in assay of SingleCellExperiment object. Only used when "assays" slotName is selected. Default NULL.
sample	Character vector. Indicates which sample each cell belongs to.
dimension	Desired dimension stored in the specified reducedDims. Either an integer which indicates the column or a character vector specifies column name. By default, the 1st dimension/column will be used. Only used when "reducedDims" slotName is selected. Default NULL.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
plotOrder	Character vector. If set, reorders the violin plots in the order of the character vector when ‘groupBy‘ is set. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
dotSize	Size of dots. Default 0.1.
transparency	Transparency of the dots, values will be 0-1. Default 1.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
gridLine	Adds a horizontal grid line if TRUE. Will still be drawn even if defaultTheme is TRUE. Default FALSE.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
hcutoff	Adds a horizontal line with the y-intercept at given value. Default NULL.
hcolor	Character. A color available from ‘colors()‘. Controls the color of the horizontal cutoff line, if drawn. Default ’black’.
hsize	Size of horizontal line, if drawn. Default 0.5.
hlinetype	Type of horizontal line, if drawn. can be specified with either an integer or a name (0 = blank, 1 = solid, 2 = dashed, 3 = dotted, 4 = dotdash, 5 = longdash, 6 = twodash). Default 1.
vcutoff	Adds a vertical line with the x-intercept at given value. Default NULL.
vcolor	Character. A color available from ‘colors()‘. Controls the color of the vertical cutoff line, if drawn. Default ’black’.

vsize	Size of vertical line, if drawn. Default 0.5.
vlinetype	Type of vertical line, if drawn. can be specified with either an integer or a name (0 = blank, 1 = solid, 2 = dashed, 3 = dotted, 4 = dotdash, 5 = longdash, 6 = twodash). Default 1.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the violin plot.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEViolin(
  inSCE = mouseBrainSubsetSCE, slotName = "assays",
  itemName = "counts", feature = "Apoe", groupBy = "sex"
)
```

---

plotSCEViolinAssayData

Violin plot of assay data.

Description

Visualizes values stored in the assay slot of a SingleCellExperiment object via a violin plot.

Usage

```
plotSCEViolinAssayData(
  inSCE,
  feature,
  sample = NULL,
  useAssay = "counts",
  featureLocation = NULL,
  featureDisplay = NULL,
  groupBy = NULL,
  violin = TRUE,
  boxplot = TRUE,
  dots = TRUE,
  plotOrder = NULL,
  xlab = NULL,
  ylab = NULL,
  axisSize = 10,
```

```
  axisLabelSize = 10,
  dotSize = 0.1,
  transparency = 1,
  defaultTheme = TRUE,
  gridLine = FALSE,
  summary = NULL,
  title = NULL,
  titleSize = NULL,
  hcutoff = NULL,
  hcolor = "red",
  hsize = 1,
  hlinetype = 1,
  vcutoff = NULL,
  vcolor = "red",
  vsize = 1,
  vlinetype = 1,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
feature	Name of feature stored in assay of SingleCellExperiment object.
sample	Character vector. Indicates which sample each cell belongs to.
useAssay	Indicate which assay to use. Default "counts".
featureLocation	Indicates which column name of rowData to query gene.
featureDisplay	Indicates which column name of rowData to use to display feature for visualization.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
plotOrder	Character vector. If set, reorders the violin plots in the order of the character vector when ‘groupBy‘ is set. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
axisSize	Size of x/y-axis ticks. Default 10.
axisLabelSize	Size of x/y-axis labels. Default 10.
dotSize	Size of dots. Default 0.1.

transparency	Transparency of the dots, values will be 0-1. Default 1.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
gridLine	Adds a horizontal grid line if TRUE. Will still be drawn even if defaultTheme is TRUE. Default FALSE.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
hcutoff	Adds a horizontal line with the y-intercept at given value. Default NULL.
hcolor	Character. A color available from 'colors()'. Controls the color of the horizontal cutoff line, if drawn. Default 'black'.
hsize	Size of horizontal line, if drawn. Default 0.5.
hlinetype	Type of horizontal line, if drawn. can be specified with either an integer or a name (0 = blank, 1 = solid, 2 = dashed, 3 = dotted, 4 = dotdash, 5 = longdash, 6 = twodash). Default 1.
vcutoff	Adds a vertical line with the x-intercept at given value. Default NULL.
vcolor	Character. A color available from 'colors()'. Controls the color of the vertical cutoff line, if drawn. Default 'black'.
vsize	Size of vertical line, if drawn. Default 0.5.
vlinetype	Type of vertical line, if drawn. can be specified with either an integer or a name (0 = blank, 1 = solid, 2 = dashed, 3 = dotted, 4 = dotdash, 5 = longdash, 6 = twodash). Default 1.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the violin plot of assay data.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEViolinAssayData(
  inSCE = mouseBrainSubsetSCE,
  feature = "Apoe", groupBy = "sex"
)
```

plotSCEViolinColData *Violin plot of colData.*

Description

Visualizes values stored in the colData slot of a SingleCellExperiment object via a violin plot.

Usage

```
plotSCEViolinColData(
  inSCE,
  coldata,
  sample = NULL,
  groupBy = NULL,
  violin = TRUE,
  boxplot = TRUE,
  dots = TRUE,
  plotOrder = NULL,
  xlab = NULL,
  ylab = NULL,
  baseSize = 12,
  axisSize = NULL,
  axisLabelSize = NULL,
  dotSize = 0.1,
  transparency = 1,
  defaultTheme = TRUE,
  gridLine = FALSE,
  summary = NULL,
  summaryTextSize = 3,
  title = NULL,
  titleSize = NULL,
  hcutoff = NULL,
  hcolor = "red",
  hsize = 1,
  hlinetype = 1,
  vcutoff = NULL,
  vcolor = "red",
  vsize = 1,
  vlinetype = 1,
  combinePlot = "none",
  plotLabels = NULL
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.

coldata	colData value that will be plotted.
sample	Character vector. Indicates which sample each cell belongs to.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in the SingleCellExperiment object, or can be retrieved from the colData slot. Default NULL.
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
plotOrder	Character vector. If set, reorders the violin plots in the order of the character vector when ‘groupBy‘ is set. Default NULL.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize.
axisSize	Size of x/y-axis ticks. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
dotSize	Size of dots. Default 0.1.
transparency	Transparency of the dots, values will be 0-1. Default 1.
defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
gridLine	Adds a horizontal grid line if TRUE. Will still be drawn even if defaultTheme is TRUE. Default FALSE.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
title	Title of plot. Default NULL.
titleSize	Size of title of plot. Default 15.
hcutoff	Adds a horizontal line with the y-intercept at given value. Default NULL.
hcolor	Character. A color available from ‘colors()‘. Controls the color of the horizontal cutoff line, if drawn. Default ’black’.
hsize	Size of horizontal line, if drawn. Default 0.5.
hlinetype	Type of horizontal line, if drawn. can be specified with either an integer or a name (0 = blank, 1 = solid, 2 = dashed, 3 = dotted, 4 = dotdash, 5 = longdash, 6 = twodash). Default 1.
vcutoff	Adds a vertical line with the x-intercept at given value. Default NULL.
vcolor	Character. A color available from ‘colors()‘. Controls the color of the vertical cutoff line, if drawn. Default ’black’.
vsize	Size of vertical line, if drawn. Default 0.5.
vlinetype	Type of vertical line, if drawn. can be specified with either an integer or a name (0 = blank, 1 = solid, 2 = dashed, 3 = dotted, 4 = dotdash, 5 = longdash, 6 = twodash). Default 1.

combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "none".
plotLabels	labels to each plot. If set to "default", will use the name of the samples as the labels. If set to "none", no label will be plotted.

Value

a ggplot of the violin plot of coldata.

Examples

```
data("mouseBrainSubsetSCE")
plotSCEViolinColData(
  inSCE = mouseBrainSubsetSCE,
  coldata = "age", groupBy = "sex"
)
```

---

plotScrubletResults *Plots for runScrublet outputs.*

Description

A wrapper function which visualizes outputs from the runScrublet function stored in the colData slot of the SingleCellExperiment object via various plots.

Usage

```
plotScrubletResults(
  inSCE,
  reducedDimName,
  sample = NULL,
  shape = NULL,
  groupBy = NULL,
  combinePlot = "all",
  violin = TRUE,
  boxplot = FALSE,
  dots = TRUE,
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  bin = NULL,
  binLabel = NULL,
  defaultTheme = TRUE,
  dotSize = 0.5,
  summary = "median",
```

```
  summaryTextSize = 3,
  transparency = 1,
  baseSize = 15,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL,
  relHeights = 1,
  relWidths = c(1, 1, 1),
  plotNCols = NULL,
  plotNRows = NULL,
  labelSamples = TRUE,
  samplePerColumn = TRUE,
  sampleRelHeights = 1,
  sampleRelWidths = 1
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results from runCxds. Required.
reducedDimName	Saved dimension reduction name in inSCE. Default "UMAP".
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
shape	If provided, add shapes based on the value. Default NULL.
groupBy	Groupings for each numeric value. A user may input a vector equal length to the number of the samples in inSCE, or can be retrieved from the colData slot. Default NULL.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
violin	Boolean. If TRUE, will plot the violin plot. Default TRUE.
boxplot	Boolean. If TRUE, will plot boxplots for each violin plot. Default TRUE.
dots	Boolean. If TRUE, will plot dots for each violin plot. Default TRUE.
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.
dim1	1st dimension to be used for plotting. Can either be a string which specifies the name of the dimension to be plotted from reducedDims, or a numeric value which specifies the index of the dimension to be plotted. Default is NULL.
dim2	2nd dimension to be used for plotting. Similar to dim1. Default is NULL.
bin	Numeric vector. If single value, will divide the numeric values into bin groups. If more than one value, will bin numeric values using values as a cut point. Default NULL.
binLabel	Character vector. Labels for the bins created by bin. Default NULL.

defaultTheme	Removes grid in plot and sets axis title size to 10 when TRUE. Default TRUE.
dotSize	Size of dots. Default 0.5.
summary	Adds a summary statistic, as well as a crossbar to the violin plot. Options are "mean" or "median". Default NULL.
summaryTextSize	The text size of the summary statistic displayed above the violin plot. Default 3.
transparency	Transparency of the dots, values will be 0-1. Default 1.
baseSize	The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize.
titleSize	Size of title of plot. Default NULL.
axisLabelSize	Size of x/y-axis labels. Default NULL.
axisSize	Size of x/y-axis ticks. Default NULL.
legendSize	size of legend. Default NULL.
legendTitleSize	size of legend title. Default NULL.
relHeights	Relative heights of plots when combine is set. Default 1.
relWidths	Relative widths of plots when combine is set. Default c(1, 1, 1).
plotNCols	Number of columns when plots are combined in a grid. Default NULL.
plotNRows	Number of rows when plots are combined in a grid. Default NULL.
labelSamples	Will label sample name in title of plot if TRUE. Default TRUE.
samplePerColumn	If TRUE, when there are multiple samples and combining by "all", the output .ggplot will have plots from each sample on a single column. Default TRUE.
sampleRelHeights	If there are multiple samples and combining by "all", the relative heights for each plot. Default 1.
sampleRelWidths	If there are multiple samples and combining by "all", the relative widths for each plot. Default 1.

Value

list of .ggplot objects

See Also

runScrublet

Examples

```
data(scExample, package="singleCellTK")
## Not run:
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runQuickUMAP(sce)
sce <- runScrublet(sce)
```

```
plotScrubletResults(inSCE=sce, reducedDimName="UMAP")

## End(Not run)
```

`plotSeuratElbow` *plotSeuratElbow Computes the plot object for elbow plot from the pca slot in the input sce object*

Description

plotSeuratElbow Computes the plot object for elbow plot from the pca slot in the input sce object

Usage

```
plotSeuratElbow(
  inSCE,
  significantPC = NULL,
  reduction = "pca",
  ndims = 20,
  externalReduction = NULL,
  interactive = TRUE
)
```

Arguments

`inSCE`	(sce) object from which to compute the elbow plot (pca should be computed)
`significantPC`	Number of significant principal components to plot. This is used to alter the color of the points for the corresponding PCs. If `NULL`, all points will be the same color. Default `NULL`.
`reduction`	Reduction to use for elbow plot generation. Either `"pca"` or `"ica"`. Default `"pca"`.
`ndims`	Number of components to use. Default `20`.
`externalReduction`	Pass DimReduc object if PCA/ICA computed through other libraries. Default `NULL`.
`interactive`	Logical value indicating if the returned object should be an interactive plotly object if `TRUE` or a ggplot object if set to `FALSE`. Default is `TRUE`.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")
plotSeuratElbow(sce)

## End(Not run)
```

---

plotSeuratGenes *Compute and plot visualizations for marker genes*

Description

Compute and plot visualizations for marker genes

Usage

```
plotSeuratGenes(
  inSCE,
  useAssay = "seuratNormData",
  plotType,
  features,
  groupVariable,
  reducedDimName = "seuratUMAP",
  splitBy = NULL,
  cols = c("lightgrey", "blue"),
  ncol = 1,
  combine = FALSE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Specify the name of the assay that will be scaled by this function.
plotType	Specify the type of the plot to compute. Options are limited to "ridge", "violin", "feature", "dot" and "heatmap".
features	Specify the features to compute the plot against.
groupVariable	Specify the column name from the colData slot that should be used as grouping variable.
reducedDimName	saved dimension reduction name in the SingleCellExperiment object. Default seuratUMAP.
splitBy	Specify the column name from the colData slot that should be used to split samples. Default is NULL.

cols	Specify two colors to form a gradient between. Default is c("lightgrey", "blue").
ncol	Visualizations will be adjusted in "ncol" number of columns. Default is 1.
combine	A logical value that indicates if the plots should be combined together into a single plot if TRUE, else if FALSE returns separate ggplot objects for each feature. Only works when plotType parameter is "feature", "violin" or "ridge". For "heatmap" and "dot", plots for all features are always combined into a single plot. Default FALSE.

Value

Plot object

---

plotSeuratHeatmap *plotSeuratHeatmap Modifies the heatmap plot object so it contains specified number of heatmaps in a single plot*

Description

plotSeuratHeatmap Modifies the heatmap plot object so it contains specified number of heatmaps in a single plot

Usage

```
plotSeuratHeatmap(plotObject, dims, ncol, labels)
```

Arguments

plotObject	plot object computed from runSeuratHeatmap() function
dims	numerical value of how many heatmaps to draw (default is 0)
ncol	numerical value indicating that in how many columns should the heatmaps be distrbuted (default is 2)
labels	list() of labels to draw on heatmaps

Value

modified plot object

`plotSeuratHVG` *plotSeuratHVG Plot highly variable genes from input sce object (must have highly variable genes computations stored)*

Description

plotSeuratHVG Plot highly variable genes from input sce object (must have highly variable genes computations stored)

Usage

```
plotSeuratHVG(inSCE, labelPoints = 0)
```

Arguments

`inSCE`	(sce) object that contains the highly variable genes computations
`labelPoints`	Numeric value indicating the number of top genes that should be labeled. Default is `0`, which will not label any point.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
plotSeuratHVG(sce)

## End(Not run)
```

`plotSeuratJackStraw` *plotSeuratJackStraw Computes the plot object for jackstraw plot from the pca slot in the input sce object*

Description

plotSeuratJackStraw Computes the plot object for jackstraw plot from the pca slot in the input sce object

Usage

```
plotSeuratJackStraw(
  inSCE,
  dims = NULL,
  xmax = 0.1,
  ymax = 0.3,
  externalReduction = NULL
)
```

Arguments

inSCE	(sce) object from which to compute the jackstraw plot (pca should be computed)
dims	Number of components to plot in Jackstraw. If NULL, then all components are plotted Default NULL.
xmax	X-axis maximum on each QQ plot. Default 0.1.
ymax	Y-axis maximum on each QQ plot. Default 0.3.
externalReduction	Pass DimReduc object if PCA/ICA computed through other libraries. Default NULL.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")
sce <- runSeuratJackStraw(sce, useAssay = "counts")
plotSeuratJackStraw(sce)

## End(Not run)
```

---

plotSeuratReduction *plotSeuratReduction Plots the selected dimensionality reduction method*

Description

plotSeuratReduction Plots the selected dimensionality reduction method

Usage

```
plotSeuratReduction(
  inSCE,
  useReduction = c("pca", "ica", "tsne", "umap"),
  showLegend = FALSE,
  groupBy = NULL,
  splitBy = NULL
)
```

Arguments

inSCE	(sce) object which has the selected dimensionality reduction algorithm already computed and stored
useReduction	Dimentionality reduction to plot. One of "pca", "ica", "tsne", or "umap". Default "umap".
showLegend	Select if legends and labels should be shown on the output plot or not. Either "TRUE" or "FALSE". Default FALSE.
groupBy	Specify a colData column name that be used for grouping. Default is NULL.
splitBy	Specify a colData column name that be used for splitting the output plot. Default is NULL.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")
plotSeuratReduction(sce, useReduction = "pca")

## End(Not run)
```

---

plotSoupXResults *Plot SoupX Result*

Description

This function will generate a combination of plots basing on the correction done by SoupX. For each sample, there will be a UMAP with cluster labeling, followed by a number of UMAPs showing the change in selected top markers. The cluster labeling is what should be used for SoupX to estimate the contamination. The Soup Fraction is calculated by subtracting the gene expression value of the output corrected matrix from that of the original input matrix, and then devided by the input.

Usage

```
plotSoupXResults(
  inSCE,
  sample = NULL,
  background = FALSE,
  reducedDimName = NULL,
  plotNCols = 3,
  plotNRows = 2,
  baseSize = 8,
  combinePlot = c("all", "sample", "none"),
  xlab = NULL,
  ylab = NULL,
  dim1 = NULL,
  dim2 = NULL,
  labelClusters = FALSE,
  clusterLabelSize = 3.5,
  defaultTheme = TRUE,
  dotSize = 0.5,
  transparency = 1,
  titleSize = NULL,
  axisLabelSize = NULL,
  axisSize = NULL,
  legendSize = NULL,
  legendTitleSize = NULL
)
```

Arguments

inSCE	A SingleCellExperiment object. With runSoupX already applied.
sample	Character vector. Indicates which sample each cell belongs to. Default NULL.
background	Logical. Whether background was applied when running runSoupX. Default FALSE.
reducedDimName	Character. The embedding to use for plotting. Leave it NULL for using the sample-specific UMAPs generated when running runSoupX. Default NULL.
plotNCols	Integer. Number of columns for the plot grid per sample. Will determine the number of top markers to show together with plotNRows. Default 3.
plotNRows	Integer. Number of rows for the plot grid per sample. Will determine the number of top markers to show together with plotNCols. Default 2.
baseSize	Numeric. The base font size for all text. Default 12. Can be overwritten by titleSize, axisSize, and axisLabelSize, legendSize, legendTitleSize. Default 8.
combinePlot	Must be either "all", "sample", or "none". "all" will combine all plots into a single .ggplot object, while "sample" will output a list of plots separated by sample. Default "all".
xlab	Character vector. Label for x-axis. Default NULL.
ylab	Character vector. Label for y-axis. Default NULL.

dim1	See plotSCEDimReduceColData. Default NULL.
dim2	See plotSCEDimReduceColData. Default NULL.
labelClusters	Logical. Whether the cluster labels are plotted. Default FALSE.
clusterLabelSize	Numeric. Determines the size of cluster label when labelClusters is set to TRUE. Default 3.5.
defaultTheme	Logical. Adds grid to plot when TRUE. Default TRUE.
dotSize	Numeric. Size of dots. Default 0.5.
transparency	Numeric. Transparency of the dots, values will be from 0 to 1. Default 1.
titleSize	Numeric. Size of title of plot. Default 15.
axisLabelSize	Numeric. Size of x/y-axis labels. Default NULL.
axisSize	Numeric. Size of x/y-axis ticks. Default NULL.
legendSize	Numeric. Size of legend. Default NULL.
legendTitleSize	Numeric. Size of legend title. Default NULL.

Value

ggplot object of the combination of UMAPs. See description.

See Also

runSoupX

Examples

```
## Not run:
sce <- importExampleData("pbmc3k")
sce <- runSoupX(sce, sample = "sample")
plotSoupXResults(sce, sample = "sample")

## End(Not run)
```

---

plotTopHVG *Plot highly variable genes*

Description

Plot highly variable genes

Usage

```
plotTopHVG(
  inSCE,
  method = "modelGeneVar",
  hvgNumber = 2000,
  useFeatureSubset = NULL,
  labelsCount = 10,
  featureDisplay = metadata(inSCE)$featureDisplay,
  labelSize = 2,
  dotSize = 2,
  textSize = 12
)
```

Arguments

inSCE	Input SingleCellExperiment object containing the computations.
method	Select either "vst", "mean.var.plot", "dispersion" or "modelGeneVar".
hvgNumber	Specify the number of top genes to highlight in red. Default 2000. See details.
useFeatureSubset	A character string for the rowData variable name to store a logical index of selected features. Default NULL. See details.
labelsCount	Specify the number of data points/genes to label. Should be less than hvgNumber. Default 10. See details.
featureDisplay	A character string for the rowData variable name to indicate what type of feature ID should be displayed. If set by setSCTKDisplayRow, will by default use it. If NULL, will use rownames(inSCE).
labelSize	Numeric, size of the text label on top HVGs. Default 2.
dotSize	Numeric, size of the dots of the features. Default 2.
textSize	Numeric, size of the text of axis title, axis label, etc. Default 12.

Details

When hvgNumber = NULL and useFeature = NULL, only plot the mean VS variance/dispersion scatter plot. When only hvgNumber set, label the top hvgNumber HVGs ranked by the metrics calculated by method. When useFeatureSubset set, label the features in the subset on the scatter plot created with method and ignore hvgNumber.

Value

ggplot of HVG metrics and top HVG labels

See Also

runFeatureSelection, runSeuratFindHVG, runModelGeneVar, getTopHVG

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runModelGeneVar(mouseBrainSubsetSCE)
plotTopHVG(mouseBrainSubsetSCE, method = "modelGeneVar")
```

`plotTSCANClusterDEG` *Plot features identified by* `runTSCANClusterDEAnalysis` *on cell 2D embedding with MST overlaid*

Description

A wrapper function which plot the top features expression identified by `runTSCANClusterDEAnalysis` on the 2D embedding of the cells cluster used in the analysis. The related MST edges are overlaid.

Usage

```
plotTSCANClusterDEG(
  inSCE,
  useCluster,
  pathIndex = NULL,
  useReducedDim = "UMAP",
  topN = 9,
  useAssay = NULL,
  featureDisplay = metadata(inSCE)$featureDisplay,
  combinePlot = c("all", "none")
)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`useCluster`	Choose a cluster used for identifying DEG with `runTSCANClusterDEAnalysis`. Required.
`pathIndex`	Specifies one of the branching paths from `useCluster` and plot the top DEGs on this path. Ususally presented by the terminal cluster of a path. By default `NULL` plot top DEGs of all paths.
`useReducedDim`	A single character for the matrix of 2D embedding. Should exist in `reducedDims` slot. Default `"UMAP"`.
`topN`	Integer. Use top N genes identified. Default `9`.
`useAssay`	A single character for the feature expression matrix. Should exist in `assayNames(inSCE)`. Default `NULL` for using the one used in `runTSCANClusterDEAnalysis`.
`featureDisplay`	Specify the feature ID type to display. Users can set default value with `setSCTKDisplayRow`. `NULL` or `"rownames"` specifies the rownames of `inSCE`. Other character values indicates `rowData` variable.
`combinePlot`	Must be either `"all"` or `"none"`. `"all"` will combine plots of each feature into a single `.ggplot` object, while `"none"` will output a list of plots. Default `"all"`.

Value

A .ggplot object of cell scatter plot, colored by the expression of a gene identified by runTSCANClusterDEAnalysis, with the layer of trajectory.

Author(s)

Yichen Wang

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
mouseBrainSubsetSCE <- runTSCANClusterDEAnalysis(inSCE = mouseBrainSubsetSCE,
                                                 useCluster = 1)
plotTSCANClusterDEG(mouseBrainSubsetSCE, useCluster = 1,
                    useReducedDim = "TSNE_logcounts")
```

---

plotTSCANClusterPseudo

Plot TSCAN pseudotime rooted from given cluster

Description

This function finds all paths that root from a given cluster useCluster. For each path, this function plots the recomputed pseudotime starting from the root on a scatter plot which contains cells only in this cluster. MST has to be pre-calculated with runTSCAN.

Usage

```
plotTSCANClusterPseudo(
  inSCE,
  useCluster,
  useReducedDim = "UMAP",
  combinePlot = c("all", "none")
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useCluster	The cluster to be regarded as the root, has to existing in colData(inSCE)$TSCAN_clusters.
useReducedDim	Saved dimension reduction name in the SingleCellExperiment object. Required.
combinePlot	Must be either "all" or "none". "all" will combine plots of pseudotime along each path into a single .ggplot object, while "none" will output a list of plots. Default "all".

Value

`combinePlot = "all"`
: A `.ggplot` object

`combinePlot = "none"`
: A list of `.ggplot`

Author(s)

Nida Pervaiz

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
plotTSCANClusterPseudo(mouseBrainSubsetSCE, useCluster = 1,
                       useReducedDim = "TSNE_logcounts")
```

---

plotTSCANDimReduceFeatures

Plot feature expression on cell 2D embedding with MST overlaid

---

Description

A wrapper function which plots all cells or cells in chosen cluster. Each point is a cell colored by the expression of a feature of interest, the relevant edges of the MST are overlaid on top.

Usage

```
plotTSCANDimReduceFeatures(
  inSCE,
  features,
  useReducedDim = "UMAP",
  useAssay = "logcounts",
  by = "rownames",
  useCluster = NULL,
  featureDisplay = metadata(inSCE)$featureDisplay,
  combinePlot = c("all", "none")
)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`features`	Choose the feature of interest to explore the expression level on the trajectory. Required.
`useReducedDim`	A single character for the matrix of 2D embedding. Should exist in `reducedDims` slot. Default `"UMAP"`.

useAssay	A single character for the feature expression matrix. Should exist in assayNames(inSCE). Default "logcounts".
by	Where should features be found? NULL, "rownames" for rownames(inSCE), otherwise will be regarded as rowData variable.
useCluster	Choose specific clusters where gene expression needs to be visualized. By default NULL, all clusters are chosen.
featureDisplay	Specify the feature ID type to display. Users can set default value with setSCTKDisplayRow. NULL or "rownames" specifies the rownames of inSCE. Other character values indicates rowData variable.
combinePlot	Must be either "all" or "none". "all" will combine plots of each feature into a single .ggplot object, while "none" will output a list of plots. Default "all".

Value

A .ggplot object of cell scatter plot, colored by the expression of a gene of interest, with the layer of trajectory.

Author(s)

Yichen Wang

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
plotTSCANDimReduceFeatures(inSCE = mouseBrainSubsetSCE,
                           features = "Tshz1",
                           useReducedDim = "TSNE_logcounts")
```

---

plotTSCANPseudotimeGenes

Plot expression changes of top features along a TSCAN pseudotime path

---

Description

A wrapper function which visualizes outputs from the runTSCANDEG function. Plots the genes that increase or decrease in expression with increasing pseudotime along the path in the MST. runTSCANDEG has to be run in advance with using the same pathIndex of interest.

Usage

```
plotTSCANPseudotimeGenes(
  inSCE,
  pathIndex,
  direction = c("increasing", "decreasing"),
  topN = 10,
  useAssay = NULL,
  featureDisplay = metadata(inSCE)$featureDisplay
)
```

Arguments

inSCE	Input SingleCellExperiment object.
pathIndex	Path index for which the pseudotime values should be used. Should have being used in runTSCANDEG.
direction	Should we show features with expression increasing or decreeasing along the increase in TSCAN pseudotime? Choices are "increasing" or "decreasing".
topN	An integer. Only to plot this number of top genes that are increasing/decreasing in expression with increasing pseudotime along the path in the MST. Default 10
useAssay	A single character to specify a feature expression matrix in assays slot. The expression of top features from here will be visualized. Default NULL use the one used for runTSCANDEG.
featureDisplay	Specify the feature ID type to display. Users can set default value with setSCTKDisplayRow. NULL or "rownames" specifies the rownames of inSCE. Other character values indicates rowData variable.

Value

A .ggplot object with the facets of the top genes. Expression on y-axis, pseudotime on x-axis.

Author(s)

Nida Pervaiz

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
terminalNodes <- listTSCANTerminalNodes(mouseBrainSubsetSCE)
mouseBrainSubsetSCE <- runTSCANDEG(inSCE = mouseBrainSubsetSCE,
                                   pathIndex = terminalNodes[1])
plotTSCANPseudotimeGenes(mouseBrainSubsetSCE,
                         pathIndex = terminalNodes[1],
                         useAssay = "logcounts")
```

plotTSCANPseudotimeHeatmap *Plot heatmap of genes with expression change along TSCAN pseudotime*

Description

A wrapper function which visualizes outputs from the runTSCANDEG function. Plots the top genes that change in expression with increasing pseudotime along the path in the MST. runTSCANDEG has to be run in advance with using the same pathIndex of interest.

Usage

```
plotTSCANPseudotimeHeatmap(
  inSCE,
  pathIndex,
  direction = c("both", "increasing", "decreasing"),
  topN = 50,
  log2fcThreshold = NULL,
  useAssay = NULL,
  featureDisplay = metadata(inSCE)$featureDisplay
)
```

Arguments

inSCE	Input SingleCellExperiment object.
pathIndex	Path index for which the pseudotime values should be used. Should have being used in runTSCANDEG.
direction	Should we show features with expression increasing or decreeasing along the increase in TSCAN pseudotime? Choices are "both", "increasing" or "decreasing".
topN	An integer. Only to plot this number of top genes along the path in the MST, in terms of FDR value. Use NULL to cancel the top N subscription. Default 30.
log2fcThreshold	Only output DEGs with the absolute values of log2FC larger than this value. Default NULL.
useAssay	A single character to specify a feature expression matrix in assays slot. The expression of top features from here will be visualized. Default NULL use the one used for runTSCANDEG.
featureDisplay	Whether to display feature ID and what ID type to display. Users can set default ID type by setSCTKDisplayRow. NULL will display when number of features to display is less than 60. FALSE for no display. Variable name in rowData to indicate ID type. "rownames" or TRUE for using rownames(inSCE).

Value

A ComplexHeatmap in .ggplot class

Author(s)

Nida Pervaiz

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
terminalNodes <- listTSCANTerminalNodes(mouseBrainSubsetSCE)
mouseBrainSubsetSCE <- runTSCANDEG(inSCE = mouseBrainSubsetSCE,
                                   pathIndex = terminalNodes[1])
plotTSCANPseudotimeHeatmap(mouseBrainSubsetSCE,
                           pathIndex = terminalNodes[1])
```

---

plotTSCANResults *Plot MST pseudotime values on cell 2D embedding*

Description

A wrapper function which visualizes outputs from the runTSCAN function. Plots the pseudotime ordering of the cells and project them onto the MST.

Usage

```
plotTSCANResults(inSCE, useReducedDim = "UMAP")
```

Arguments

inSCE	Input SingleCellExperiment object.
useReducedDim	Saved dimension reduction name in inSCE object. Required.

Value

A .ggplot object with the pseudotime ordering of the cells colored on a cell 2D embedding, and the MST path drawn on it.

Author(s)

Nida Pervaiz

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
plotTSCANResults(inSCE = mouseBrainSubsetSCE,
                 useReducedDim = "TSNE_logcounts")
```

plotTSNE *Plot t-SNE plot on dimensionality reduction data run from t-SNE method.*

Description

Plot t-SNE plot on dimensionality reduction data run from t-SNE method.

Usage

```
plotTSNE(
  inSCE,
  colorBy = NULL,
  shape = NULL,
  reducedDimName = "TSNE",
  runTSNE = FALSE,
  useAssay = "counts"
)
```

Arguments

inSCE	Input SingleCellExperiment object.
colorBy	color by condition.
shape	add shape to each distinct label.
reducedDimName	a name to store the results of the dimension reduction coordinates obtained from this method. This is stored in the SingleCellExperiment object in the reducedDims slot. Required.
runTSNE	Run t-SNE if the reducedDimName does not exist. the Default is FALSE.
useAssay	Indicate which assay to use. The default is "logcounts".

Value

A t-SNE plot

Examples

```
data("mouseBrainSubsetSCE")
plotTSNE(mouseBrainSubsetSCE, colorBy = "level1class",
         reducedDimName = "TSNE_counts")
```

plotUMAP *Plot UMAP results either on already run results or run first and then plot.*

Description

Plot UMAP results either on already run results or run first and then plot.

Usage

```
plotUMAP(
  inSCE,
  colorBy = NULL,
  shape = NULL,
  reducedDimName = "UMAP",
  runUMAP = FALSE,
  useAssay = "counts"
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components. Required
colorBy	color by a condition(any column of the annotation data).
shape	add shapes to each condition.
reducedDimName	saved dimension reduction name in the SingleCellExperiment object. Required.
runUMAP	If the dimension reduction components are already available set this to FALSE, otherwise set to TRUE. Default is False.
useAssay	Indicate which assay to use. The default is "logcounts"

Value

a UMAP plot of the reduced dimensions.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runQuickUMAP(sce)
plotUMAP(sce)
```

qcInputProcess *Create SingleCellExperiment object from command line input arguments*

Description

Create SingleCellExperiment object from command line input arguments

Usage

```
qcInputProcess(
  preproc,
  samplename,
  path,
  raw,
  fil,
  ref,
  rawFile,
  filFile,
  flatFiles,
  dataType
)
```

Arguments

preproc	Method used to preprocess the data. It's one of the path provided in –preproc argument.
samplename	The sample name of the data. It's one of the path provided in –sample argument.
path	Base path of the dataset. It's one of the path provided in –bash_path argument.
raw	The directory contains droplet matrix, gene and cell barcodes information. It's one of the path provided in –raw_data_path argument.
fil	The directory contains cell matrix, gene and cell barcodes information. It's one of the path provided in –cell_data_path argument.
ref	The name of reference used by cellranger. Only need for CellrangerV2 data.
rawFile	The full path of the RDS file or Matrix file of the raw gene count matrix. It's one of the path provided in –raw_data argument.
filFile	The full path of the RDS file or Matrix file of the cell count matrix. It's one of the path provided in –cell_data argument.
flatFiles	The full paths of the matrix, barcode, and features (in that order) files used to construct an SCE object.
dataType	Type of the input. It can be "Both", "Droplet" or "Cell". It's one of the path provided in –genome argument.

Value

A list of SingleCellExperiment object containing the droplet or cell data or both,depending on the dataType that users provided.

`readSingleCellMatrix` *Read single cell expression matrix*

Description

Automatically detact the format of the input file and read the file.

Usage

```
readSingleCellMatrix(
  file,
  class = c("Matrix", "matrix"),
  delayedArray = TRUE,
  colIndexLocation = NULL,
  rowIndexLocation = NULL
)
```

Arguments

`file`	Path to input file. Supported file endings include .mtx, .txt, .csv, .tab, .tsv, .npz, and their corresponding `gzip`, `bzip2`, or `xz` compressed extensions (`*.gz`, `*.bz2`, or `*.xz`).
`class`	Character. Class of matrix. One of "Matrix" or "matrix". Specifying "Matrix" will convert to a sparse format which should be used for datasets with large numbers of cells. Default "Matrix".
`delayedArray`	Boolean. Whether to read the expression matrix as DelayedArray object or not. Default `TRUE`.
`colIndexLocation`	Character. For Optimus output, the path to the barcode index .npy file. Used only if `file` has .npz extension. Default `NULL`.
`rowIndexLocation`	Character. For Optimus output, The path to the feature (gene) index .npy file. Used only if `file` has .npz extension. Default `NULL`.

Value

A DelayedArray object or matrix.

Examples

```
mat <- readSingleCellMatrix(system.file("extdata/hgmm_1k_v3_20x20/outs/",
    "filtered_feature_bc_matrix/matrix.mtx.gz", package = "singleCellTK"))
```

`reportCellQC` *Get runCellQC .html report*

Description

A function to generate .html Rmarkdown report containing the visualizations of the runCellQC function output

Usage

```
reportCellQC(
  inSCE,
  output_file = NULL,
  output_dir = NULL,
  subTitle = NULL,
  studyDesign = NULL,
  useReducedDim = NULL
)
```

Arguments

`inSCE`	A SingleCellExperiment object containing the filtered count matrix with the output from runCellQC function
`output_file`	Character. The name of the generated file. If NULL/default then the output file name will be based on the name of the Rmarkdown template.
`output_dir`	Character. The name of the output directory to save the rendered file. If NULL/default the file is stored to the current working directory
`subTitle`	subtitle of the QC HTML report. Default is NULL.
`studyDesign`	Character. The description of the data set and experiment design. It would be shown at the top of QC HTML report. Default is NULL.
`useReducedDim`	Character. The name of the saved dimension reduction slot including cells from all samples in thenSingleCellExperiment object, Default is NULL

Value

.html file

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
## Not run:
sce <- runCellQC(sce)
reportCellQC(inSCE = sce)

## End(Not run)
```

reportClusterAbundance *Get plotClusterAbundance .html report*

Description

A function to generate .html Rmarkdown report containing the visualizations of the plotClusterAbundance function output

Usage

```
reportClusterAbundance(
  inSCE,
  cluster,
  variable,
  output_dir = ".",
  output_file = "plotClusterAbundance_Report",
  pdf = FALSE,
  showSession = TRUE
)
```

Arguments

inSCE	A SingleCellExperiment object.
cluster	A single character, specifying the name to store the cluster label in colData.
variable	A single character, specifying the name to store the phenotype labels in colData.
output_dir	name of the output directory to save the rendered file. If NULL the file is stored to the current working directory. Default NULL.
output_file	name of the generated file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is TRUE.
showSession	A logical value indicating if session information should be displayed or not. Default is TRUE.

Value

An HTML file of the report will be generated at the path specified in the arguments.

reportDiffAbundanceFET *Get diffAbundanceFET .html report*

Description

A function to generate .html Rmarkdown report containing the visualizations of the diffAbundanceFET function output

Usage

```
reportDiffAbundanceFET(
  inSCE,
  cluster,
  variable,
  control,
  case,
  analysisName,
  output_dir = ".",
  output_file = "DifferentialAbundanceFET_Report",
  pdf = FALSE,
  showSession = TRUE
)
```

Arguments

inSCE	A SingleCellExperiment object.
cluster	A single character, specifying the name to store the cluster label in colData.
variable	A single character, specifying the name to store the phenotype labels in colData.
control	character. Specifying one or more categories that can be found in the vector specified by variable.
case	character. Specifying one or more categories that can be found in the vector specified by variable.
analysisName	A single character. Will be used for naming the result table, which will be saved in metadata slot.
output_dir	name of the output directory to save the rendered file. If NULL the file is stored to the current working directory. Default NULL.
output_file	name of the generated file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is TRUE.
showSession	A logical value indicating if session information should be displayed or not. Default is TRUE.

Value

An HTML file of the report will be generated at the path specified in the arguments.

`reportDiffExp` *Get runDEAnalysis .html report*

Description

A function to generate .html Rmarkdown report containing the visualizations of the `runDEAnalysis` function output

Usage

```
reportDiffExp(
  inSCE,
  study,
  useReducedDim,
  featureDisplay = NULL,
  output_file = NULL,
  output_dir = NULL
)
```

Arguments

`inSCE`	A `SingleCellExperiment` object containing the output from `runDEAnalysis` function
`study`	The specific analysis to visualize, used as `analysisName` argument when running differential expression.
`useReducedDim`	Specify an embedding for visualizing the relation ship between the conditions.
`featureDisplay`	The feature ID type to use for displaying. Should exists as a variable name of `rowData`. Default `NULL` use rownames of `inSCE`.
`output_file`	name of the generated file. If `NULL` then the output file name will be based on the name of the Rmarkdown template. Default `NULL`.
`output_dir`	name of the output directory to save the rendered file. If `NULL` the file is stored to the current working directory. Default `NULL`.

Value

Saves the HTML report in the specified output directory.

`reportDropletQC` *Get runDropletQC .html report*

Description

A function to generate .html Rmarkdown report containing the visualizations of the runDropletQC function output

Usage

```
reportDropletQC(
  inSCE,
  output_file = NULL,
  output_dir = NULL,
  subTitle = NULL,
  studyDesign = NULL
)
```

Arguments

`inSCE`	A SingleCellExperiment object containing the full droplet count matrix with the output from runDropletQC function
`output_file`	name of the generated file. If NULL/default then the output file name will be based on the name of the Rmarkdown template
`output_dir`	name of the output directory to save the rendered file. If NULL/default the file is stored to the current working directory
`subTitle`	subtitle of the QC HTML report. Default is NULL.
`studyDesign`	description of the data set and experiment design. It would be shown at the top of QC HTML report. Default is NULL.

Value

.html file

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runDropletQC(sce)
reportDropletQC(inSCE = sce)

## End(Not run)
```

reportFindMarker *Get runFindMarker .html report*

Description

A function to generate .html Rmarkdown report containing the visualizations of the runFindMarker function output

Usage

```
reportFindMarker(inSCE, output_file = NULL, output_dir = NULL)
```

Arguments

inSCE	A SingleCellExperiment object containing the output from runFindMarker function
output_file	name of the generated file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
output_dir	name of the output directory to save the rendered file. If NULL the file is stored to the current working directory. Default NULL.

Value

An HTML file of the report will be generated at the path specified in the arguments.

reportQCTool *Get .html report of the output of the selected QC algorithm*

Description

A function to generate .html Rmarkdown report for the specified QC algorithm output

Usage

```
reportQCTool(
  inSCE,
 algorithm = c("BarcodeRankDrops", "EmptyDrops", "QCMetrics", "Scrublet", "ScDblFinder",
    "Cxds", "Bcds", "CxdsBcdsHybrid", "DoubletFinder", "DecontX", "SoupX"),
  output_file = NULL,
  output_dir = NULL
)
```

Arguments

inSCE	A SingleCellExperiment object containing the count matrix (full droplets or filtered matrix, depends on the selected QC algorithm) with the output from at least one of these functions: runQCMetrics, runScrublet, runScDblFinder, runCxds, runBcds, runCxdsBcdsHybrid, runDecontX, runBarcodeRankDrops, runEmptyDrops
algorithm	Character. Specifies which QC algorithm report to generate. Available options are "BarcodeRankDrops", "EmptyDrops", "QCMetrics", "Scrublet", "ScDblFinder", "Cxds", "Bcds", "CxdsBcdsHybrid", "DoubletFinder", "DecontX" and "SoupX".
output_file	name of the generated file. If NULL/default then the output file name will be based on the name of the selected QC algorithm name .
output_dir	name of the output directory to save the rendered file. If NULL/default the file is stored to the current working directory

Value

.html file

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
## Not run:
sce <- runDecontX(sce)
sce <- runQuickUMAP(sce)
reportQCTool(inSCE = sce, algorithm = "DecontX")

## End(Not run)
```

---

reportSeurat *Generates an HTML report for the complete Seurat workflow and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for the complete Seurat workflow and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeurat(
  inSCE,
  biological.group = NULL,
  phenotype.groups = NULL,
  selected.markers = NULL,
  clustering.resolution = 0.8,
```

```
  variable.features = 2000,
  pc.count = 50,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  runHVG = TRUE,
  plotHVG = TRUE,
  runDimRed = TRUE,
  plotJackStraw = FALSE,
  plotElbowPlot = TRUE,
  plotHeatmaps = TRUE,
  runClustering = TRUE,
  plotTSNE = TRUE,
  plotUMAP = TRUE,
  minResolution = 0.3,
  maxResolution = 1.5,
  runMSClusters = TRUE,
  runMSBioGroup = TRUE,
  numTopFeatures = 10,
  forceRun = TRUE
)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`biological.group`	A character value that specifies the name of the `colData()` column to use as the main biological group in the Seurat report for marker selection and grouping.
`phenotype.groups`	A character vector that specifies the names of the `colData()` columns to use for differential expression in addition to the `biological.group` parameter.
`selected.markers`	A character vector containing the user-specified gene symbols or feature names of marker genes that be used to generate gene plots in addition to the gene markers computed from differential expression.
`clustering.resolution`	A numeric value indicating the user-specified final resolution to use with clustering. Default is `0.8`.
`variable.features`	A numeric value indicating the number of top variable features to identify. Default `2000`.
`pc.count`	A numeric value indicating the number of principal components to use in the analysis workflow. Default is `50`.
`outputFile`	Specify the name of the generated output HTML file. If `NULL` then the output file name will be based on the name of the Rmarkdown template. Default `NULL`.

outputPath	Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.
subtitle	A character value specifying the subtitle to use in the report. Default NULL.
authors	A character value specifying the names of the authors to use in the report. Default NULL.
showSession	A logical value indicating if session information should be displayed or not. Default is FALSE.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.
runHVG	A logical value indicating if the feature selection computation should be run or not. Default is TRUE.
plotHVG	A logical value indicating if the plot for the top most variable genes should be visualized in a mean-to-variance plot. Default is TRUE.
runDimRed	A logical value indicating if PCA should be computed. Default is TRUE.
plotJackStraw	A logical value indicating if JackStraw plot be visualized for the principal components. Default is FALSE.
plotElbowPlot	A logical value indicating if the ElbowPlot be visualized for the principal components. Default is TRUE.
plotHeatmaps	A logical value indicating if heatmaps should be plotted for the principal components. Default is TRUE.
runClustering	A logical value indicating if clustering section should be run in the report. Default is TRUE.
plotTSNE	A logical value indicating if TSNE plots should be visualized for clustering results. Default is TRUE.
plotUMAP	A logical value indicating if the UMAP plots should be visualized for the clustering results. Default is TRUE.
minResolution	A numeric value indicating the minimum resolution to use for clustering. Default is 0.3.
maxResolution	A numeric value indicating the maximum resolution to use for clustering. Default is 1.5.
runMSClusters	A logical value indicating if marker selection should be run between clusters. Default is TRUE.
runMSBioGroup	A logical value indicating if marker selection should be run between the biological.group parameter. Default is TRUE.
numTopFeatures	A numeric value indicating the number of top features to visualize in each group. Default 10.
forceRun	A logical value indicating if all algorithms should be re-run regardless if they have been computed previously in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

reportSeuratClustering *Generates an HTML report for Seurat Clustering and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for Seurat Clustering and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratClustering(
  inSCE,
  biological.group = NULL,
  phenotype.groups = NULL,
  runClustering = TRUE,
  plotTSNE = TRUE,
  plotUMAP = TRUE,
  minResolution = 0.3,
  maxResolution = 1.5,
  numClusters = 10,
  significant_PC = 10,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  forceRun = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
biological.group	A character value that specifies the name of the colData() column to use as the main biological group in the Seurat report for marker selection and grouping.
phenotype.groups	A character vector that specifies the names of the colData() columns to use for differential expression in addition to the biological.group parameter.
runClustering	A logical value indicating if Clustering should be run or not in the report. Default is TRUE. If FALSE, parameters plotTSNE and plotUMAP are also set to FALSE.
plotTSNE	A logical value indicating if TSNE plots should be visualized in the clustering section of the report. Default is TRUE.

plotUMAP	A logical value indicating if UMAP plots should be visualized in the clustering section of the report. Default is TRUE.
minResolution	A numeric value indicating the minimum resolution to use for clustering. Default 0.3.
maxResolution	A numeric value indicating the maximum resolution to use for clustering. Default 1.5.
numClusters	temp (to remove)
significant_PC	temp (change to pc.use)
outputFile	Specify the name of the generated output HTML file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
outputPath	Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.
subtitle	A character value specifying the subtitle to use in the report. Default NULL.
authors	A character value specifying the names of the authors to use in the report. Default NULL.
showSession	A logical value indicating if session information should be displayed or not. Default is FALSE.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.
forceRun	A logical value indicating if all computations previously computed should be re-calculated regardless if these computations are available in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

`reportSeuratDimRed` *Generates an HTML report for Seurat Dimensionality Reduction and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for Seurat Dimensionality Reduction and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratDimRed(
  inSCE,
  pc.count = 50,
  runDimRed = TRUE,
  plotJackStraw = FALSE,
```

```
  plotElbowPlot = TRUE,
  plotHeatmaps = TRUE,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  forceRun = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
pc.count	A numeric value indicating the number of principal components to compute. Default is 50.
runDimRed	A logical value indicating if dimenionality reduction should be computed. Default TRUE.
plotJackStraw	A logical value indicating if JackStraw plot should be visualized. Default FALSE.
plotElbowPlot	A logical value indicating if ElbowPlot should be visualized. Default TRUE.
plotHeatmaps	A logical value indicating if heatmaps should be visualized. Default TRUE.
outputFile	Specify the name of the generated output HTML file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
outputPath	Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.
subtitle	A character value specifying the subtitle to use in the report. Default NULL.
authors	A character value specifying the names of the authors to use in the report. Default NULL.
showSession	A logical value indicating if session information should be displayed or not. Default is FALSE.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.
forceRun	A logical value indicating if all computations previously computed should be re-calculated regardless if these computations are available in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

`reportSeuratFeatureSelection` *Generates an HTML report for Seurat Feature Selection and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for Seurat Feature Selection and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratFeatureSelection(
  inSCE,
  variable.features = 2000,
  runHVG = TRUE,
  plotHVG = TRUE,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  forceRun = TRUE
)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`variable.features`	A numeric value indicating the number of top variable features to identify. Default `2000`.
`runHVG`	A logical value indicating if the feature selection algorithm should be run or not. Default `TRUE`.
`plotHVG`	A logical value indicating if the mean-to-variance plot of the top variable feature should be visualized or not. Default `TRUE`.
`outputFile`	Specify the name of the generated output HTML file. If `NULL` then the output file name will be based on the name of the Rmarkdown template. Default `NULL`.
`outputPath`	Specify the name of the output directory to save the rendered HTML file. If `NULL` the file is stored to the current working directory. Default `NULL`.
`subtitle`	A character value specifying the subtitle to use in the report. Default `NULL`.
`authors`	A character value specifying the names of the authors to use in the report. Default `NULL`.
`showSession`	A logical value indicating if session information should be displayed or not. Default is `FALSE`.

pdf — A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.

forceRun — A logical value indicating if all computations previously computed should be re-calculated regardless if these computations are available in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

`reportSeuratMarkerSelection` *Generates an HTML report for Seurat Results (including Clustering & Marker Selection) and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for Seurat Results (including Clustering & Marker Selection) and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratMarkerSelection(
  inSCE,
  biological.group = NULL,
  phenotype.groups = NULL,
  selected.markers = NULL,
  runMarkerSelection = TRUE,
  plotMarkerSelection = TRUE,
  numTopFeatures = 10,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE
)
```

Arguments

inSCE — Input SingleCellExperiment object.

biological.group — A character value that specifies the name of the colData() column to use as the main biological group in the Seurat report for marker selection and grouping.

phenotype.groups
: A character vector that specifies the names of the colData() columns to use for differential expression in addition to the biological.group parameter.

selected.markers
: A character vector containing the user-specified gene symbols or feature names of marker genes that be used to generate gene plots in addition to the gene markers computed from differential expression.

runMarkerSelection
: A logical value indicating if the marker selection computation should be run or not. Default TRUE.

plotMarkerSelection
: A logical value indicating if the gene marker plots should be visualized or not. Default TRUE.

numTopFeatures
: A numeric value indicating the number of top features to visualize in each group. Default 10.

outputFile
: Specify the name of the generated output HTML file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.

outputPath
: Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.

subtitle
: A character value specifying the subtitle to use in the report. Default NULL.

authors
: A character value specifying the names of the authors to use in the report. Default NULL.

showSession
: A logical value indicating if session information should be displayed or not. Default is FALSE.

pdf
: A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.

Value

A SingleCellExperiment object with computations stored.

`reportSeuratNormalization`

Generates an HTML report for Seurat Normalization and returns the SCE object with the results computed and stored inside the object.

Description

Generates an HTML report for Seurat Normalization and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratNormalization(
  inSCE,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  forceRun = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object previously passed through reportSeuratRun().
outputFile	Specify the name of the generated output HTML file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
outputPath	Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.
subtitle	A character value specifying the subtitle to use in the report. Default NULL.
authors	A character value specifying the names of the authors to use in the report. Default NULL.
showSession	A logical value indicating if session information should be displayed or not. Default is FALSE.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.
forceRun	A logical value indicating if all computations previously computed should be re-calculated regardless if these computations are available in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

---

reportSeuratResults *Generates an HTML report for Seurat Results (including Clustering & Marker Selection) and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for Seurat Results (including Clustering & Marker Selection) and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratResults(
  inSCE,
  biological.group = NULL,
  phenotype.groups = NULL,
  selected.markers = NULL,
  clustering.resolution = 0.8,
  pc.count = 50,
  plotTSNE = TRUE,
  plotUMAP = TRUE,
  runClustering = TRUE,
  runMSClusters = TRUE,
  runMSBioGroup = TRUE,
  numTopFeatures = 10,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  forceRun = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object previously passed through reportSeuratRun().
biological.group	A character value that specifies the name of the colData() column to use as the main biological group in the Seurat report for marker selection and grouping.
phenotype.groups	A character vector that specifies the names of the colData() columns to use for differential expression in addition to the biological.group parameter.
selected.markers	A character vector containing the user-specified gene symbols or feature names of marker genes that be used to generate gene plots in addition to the gene markers computed from differential expression.
clustering.resolution	A numeric value indicating the user-specified final resolution to use with clustering. Default is 0.8.
pc.count	A numeric value indicating the number of principal components to use in the analysis workflow. Default is 50.
plotTSNE	A logical value indicating if TSNE plots should be visualized in the clustering section of the report. Default is TRUE.
plotUMAP	A logical value indicating if UMAP plots should be visualized in the clustering section of the report. Default is TRUE.

runClustering	A logical value indicating if Clustering should be run or not in the report. Default is TRUE. If FALSE, parameters plotTSNE and plotUMAP are also set to FALSE.
runMSClusters	A logical value indicating if the marker selection section for identifying marker genes between clusters should be run and visualized in the report. Default TRUE.
runMSBioGroup	A logical value indicating if the marker selection section for identifying marker genes between the biological.group parameter should be run and visualized in the report. Default TRUE.
numTopFeatures	A numeric value indicating the number of top features to visualize in each group. Default 10.
outputFile	Specify the name of the generated output HTML file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
outputPath	Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.
subtitle	A character value specifying the subtitle to use in the report. Default NULL.
authors	A character value specifying the names of the authors to use in the report. Default NULL.
showSession	A logical value indicating if session information should be displayed or not. Default is FALSE.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.
forceRun	A logical value indicating if all computations previously computed should be re-calculated regardless if these computations are available in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

---

reportSeuratRun *Generates an HTML report for Seurat Run (including Normalization, Feature Selection, Dimensionality Reduction & Clustering) and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for Seurat Run (including Normalization, Feature Selection, Dimensionality Reduction & Clustering) and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratRun(
  inSCE,
  biological.group = NULL,
  phenotype.groups = NULL,
  variable.features = 2000,
  pc.count = 50,
  runHVG = TRUE,
  plotHVG = TRUE,
  runDimRed = TRUE,
  plotJackStraw = FALSE,
  plotElbowPlot = TRUE,
  plotHeatmaps = TRUE,
  runClustering = TRUE,
  plotTSNE = TRUE,
  plotUMAP = TRUE,
  minResolution = 0.3,
  maxResolution = 1.5,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  forceRun = TRUE
)
```

Arguments

`inSCE`	Input `SingleCellExperiment` object.
`biological.group`	A character value that specifies the name of the `colData()` column to use as the main biological group in the Seurat report for tSNE & UMAP visualization.
`phenotype.groups`	A character value that specifies the name of the `colData()` column to use as additional phenotype variables in the Seurat report for tSNE & UMAP visualization.
`variable.features`	A numeric value indicating the number of top variable genes to identify in the report. Default is `2000`.
`pc.count`	A numeric value indicating the number of principal components to use in the analysis workflow. Default is `50`.
`runHVG`	A logical value indicating if feature selection should be run in the report. Default `TRUE`.
`plotHVG`	A logical value indicating if the top variable genes should be visualized through a mean-to-variance plot. Default is `TRUE`.

runDimRed	A logical value indicating if PCA should be computed in the report. Default is TRUE.
plotJackStraw	A logical value indicating if the JackStraw plot should be visualized for the principal components. Default is FALSE.
plotElbowPlot	A logical value indicating if the ElbowPlot should be visualized for the principal components. Default is FALSE.
plotHeatmaps	A logical value indicating if the Heatmaps should be visualized for the principal components. Default is FALSE.
runClustering	A logical value indicating if Clustering should be run over multiple resolutions as defined by the minResolution and maxResolution parameters. Default is TRUE.
plotTSNE	A logical value indicating if TSNE plot should be visualized for clusters. Default is TRUE.
plotUMAP	A logical value indicating if UMAP plot should be visualized for clusters. Default is TRUE.
minResolution	A numeric value indicating the minimum resolution to use for clustering. Default 0.3.
maxResolution	A numeric value indicating the maximum resolution to use for clustering. Default 1.5.
outputFile	Specify the name of the generated output HTML file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
outputPath	Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.
subtitle	A character value specifying the subtitle to use in the report. Default NULL.
authors	A character value specifying the names of the authors to use in the report. Default NULL.
showSession	A logical value indicating if session information should be displayed or not. Default is FALSE.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.
forceRun	A logical value indicating if all computations previously computed should be re-calculated regardless if these computations are available in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

reportSeuratScaling *Generates an HTML report for Seurat Scaling and returns the SCE object with the results computed and stored inside the object.*

Description

Generates an HTML report for Seurat Scaling and returns the SCE object with the results computed and stored inside the object.

Usage

```
reportSeuratScaling(
  inSCE,
  outputFile = NULL,
  outputPath = NULL,
  subtitle = NULL,
  authors = NULL,
  showSession = FALSE,
  pdf = FALSE,
  forceRun = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
outputFile	Specify the name of the generated output HTML file. If NULL then the output file name will be based on the name of the Rmarkdown template. Default NULL.
outputPath	Specify the name of the output directory to save the rendered HTML file. If NULL the file is stored to the current working directory. Default NULL.
subtitle	A character value specifying the subtitle to use in the report. Default NULL.
authors	A character value specifying the names of the authors to use in the report. Default NULL.
showSession	A logical value indicating if session information should be displayed or not. Default is FALSE.
pdf	A logical value indicating if a pdf should also be generated for each figure in the report. Default is FALSE.
forceRun	A logical value indicating if all computations previously computed should be re-calculated regardless if these computations are available in the input object. Default is TRUE.

Value

A SingleCellExperiment object with computations stored.

retrieveSCEIndex *Retrieve cell/feature index by giving identifiers saved in col/rowData*

Description

Originally written in retrieveFeatureIndex. Modified for also retrieving cell indices and only working for SingleCellExperiment object. This will return indices of features among the rowData/colData. Partial matching (i.e. grepping) can be used.

Usage

```
retrieveSCEIndex(
  inSCE,
  IDs,
  axis,
  by = NULL,
  exactMatch = TRUE,
  firstMatch = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object. Required
IDs	Character vector of identifiers for features or cells to find in rowData or colData of inSCE
axis	A character scalar to specify whether to search for features or cells. Use "row", "feature" or "gene" for features; "col" or "cell" for cells.
by	Character. In which column to search for features/cells in rowData/colData. Default NULL for search the rownames/colnames
exactMatch	A logical scalar. Whether to only identify exact matches or to identify partial matches using grep. Default TRUE
firstMatch	A logical scalar. Whether to only identify the first matches or to return all plausible matches. Default TRUE

Value

A unique, non-NA numeric vector of indices for the matching features/cells in inSCE.

Author(s)

Yusuke Koga, Joshua Campbell, Yichen Wang

Examples

```
data(scExample, package = "singleCellTK")
retrieveSCEIndex(inSCE = sce, IDs = "ENSG00000205542",
 axis = "row")
```

runBarcodeRankDrops *Identify empty droplets using barcodeRanks.*

Description

Run barcodeRanks on a count matrix provided in a SingleCellExperiment object. Distinguish between droplets containing cells and ambient RNA in a droplet-based single-cell RNA sequencing experiment.

Usage

```
runBarcodeRankDrops(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  lower = 100,
  fitBounds = NULL,
  df = 20
)
```

Arguments

inSCE	A SingleCellExperiment object. Must contain a raw counts matrix before empty droplets have been removed.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
useAssay	A string specifying which assay in the SCE to use. Default "counts"
lower	See barcodeRanks for more information. Default 100.
fitBounds	See barcodeRanks for more information. Default NULL.
df	See barcodeRanks for more information. Default 20.

Value

A SingleCellExperiment object with the barcodeRanks output table appended to the colData slot. The columns include dropletUtils_BarcodeRank_Knee and dropletUtils_barcodeRank_inflection. Please refer to the documentation of barcodeRanks for details.

See Also

barcodeRanks, runDropletQC, plotBarcodeRankDropsResults

Examples

```
data(scExample, package = "singleCellTK")
sce <- runBarcodeRankDrops(inSCE = sce)
```

runBBKNN *Apply BBKNN batch effect correction method to SingleCellExperiment object*

Description

BBKNN, an extremely fast graph-based data integration algorithm. It modifies the neighbourhood construction step to produce a graph that is balanced across all batches of the data.

Usage

```
runBBKNN(
  inSCE,
  useAssay = "logcounts",
  batch = "batch",
  reducedDimName = "BBKNN",
  nComponents = 50L
)
```

Arguments

inSCE	Input SingleCellExperiment object
useAssay	A single character indicating the name of the assay requiring batch correction. Default "logcounts".
batch	A single character indicating a field in colData that annotates the batches of each cell; or a vector/factor with the same length as the number of cells. Default "batch".
reducedDimName	A single character. The name for the corrected low-dimensional representation. Will be saved to reducedDim(inSCE). Default "BBKNN".
nComponents	An integer. Number of principle components or the dimensionality, adopted in the pre-PCA-computation step, the BBKNN step (for how many PCs the algorithm takes into account), and the final UMAP combination step where the value represent the dimensionality of the updated reducedDim. Default 50L.

Value

The input SingleCellExperiment object with reducedDim(inSCE, reducedDimName) updated.

References

Krzysztof Polanski et al., 2020

Examples

```
## Not run:
data('sceBatches', package = 'singleCellTK')
logcounts(sceBatches) <- log1p(counts(sceBatches))
sceBatches <- runBBKNN(sceBatches, useAssay = "logcounts",
                       nComponents = 10)

## End(Not run)
```

---

runBcds *Find doublets/multiplets using bcds.*

Description

A wrapper function for bcds. Annotate doublets/multiplets using a binary classification approach to discriminate artificial doublets from original data. Generate a doublet score for each cell. Infer doublets if `estNdbl` is `TRUE`.

Usage

```
runBcds(
  inSCE,
  sample = NULL,
  seed = 12345,
  ntop = 500,
  srat = 1,
  verb = FALSE,
  retRes = FALSE,
  nmax = "tune",
  varImp = FALSE,
  estNdbl = FALSE,
  useAssay = "counts"
)
```

Arguments

`inSCE`	A SingleCellExperiment object.
`sample`	Character vector or colData variable name. Indicates which sample each cell belongs to. Default `NULL`.
`seed`	Seed for the random number generator, can be `NULL`. Default `12345`.
`ntop`	See bcds for more information. Default `500`.
`srat`	See bcds for more information. Default `1`.
`verb`	See bcds for more information. Default `FALSE`.
`retRes`	See bcds for more information. Default `FALSE`.
`nmax`	See bcds for more information. Default `"tune"`.

varImp	See bcds for more information. Default FALSE.
estNdbl	See bcds for more information. Default FALSE.
useAssay	A string specifying which assay in inSCE to use. Default "counts"

Details

When the argument sample is specified, bcds will be run on cells from each sample separately. If sample = NULL, then all cells will be processed together.

Value

A SingleCellExperiment object with bcds output appended to the colData slot. The columns include *bcds_score* and optionally *bcds_call*. Please refer to the documentation of bcds for details.

See Also

bcds, plotBcdsResults, runCellQC

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runBcds(sce)
```

---

runCellQC *Perform comprehensive single cell QC*

Description

A wrapper function to run several QC algorithms on a SingleCellExperiment object containing cells after empty droplets have been removed.

Usage

```
runCellQC(
  inSCE,
  algorithms = c("QCMetrics", "scDblFinder", "cxds", "bcds", "cxds_bcds_hybrid",
    "decontX", "decontX_bg", "soupX", "soupX_bg"),
  sample = NULL,
  collectionName = NULL,
  geneSetList = NULL,
  geneSetListLocation = "rownames",
  geneSetCollection = NULL,
  mitoRef = "human",
  mitoIDType = "ensembl",
  mitoPrefix = "MT-",
  mitoID = NULL,
```

```
  mitoGeneLocation = "rownames",
  useAssay = "counts",
  background = NULL,
  bgAssayName = NULL,
  bgBatch = NULL,
  seed = 12345,
  paramsList = NULL
)
```

Arguments

inSCE	A SingleCellExperiment object.
algorithms	Character vector. Specify which QC algorithms to run. Available options are "QCMetrics", "scrublet", "doubletFinder", "scDblFinder", "cxds", "bcds", "cxds_bcds_hybrid", "decontX" and "soupX".
sample	Character vector. Indicates which sample each cell belongs to. Algorithms will be run on cells from each sample separately.
collectionName	Character. Name of a GeneSetCollection obtained by using one of the import-GeneSet* functions. Default NULL.
geneSetList	See runPerCellQC. Default NULL.
geneSetListLocation	See runPerCellQC. Default NULL.
geneSetCollection	See runPerCellQC. Default NULL.
mitoRef, mitoIDType, mitoPrefix, mitoID, mitoGeneLocation	Arguments used to import mitochondrial genes and quantify their expression. Please see runPerCellQC for detailed information.
useAssay	A string specifying which assay contains the count matrix for cells.
background	A SingleCellExperiment with the matrix located in the assay slot under bgAssayName. It should have the same structure as inSCE except it contains the matrix of empty droplets instead of cells. When supplied, empirical distribution of transcripts from these empty droplets will be used as the contamination distribution. It is only used in algorithms "decontX" and "soupX". Default NULL.
bgAssayName	Character. Name of the assay to use if background is a SingleCellExperiment. If NULL, the function will use the same value as useAssay. It is only used in algorithms "decontX" and "soupX". Default is NULL.
bgBatch	Batch labels for background. If background is a SingleCellExperiment object, this can be a single character specifying a name that can be found in colData(background) to directly use the barcode annotation Its unique values should be the same as those in sample, such that each batch of cells have their corresponding batch of empty droplets as background, pointed by this parameter. It is only used in algorithms "decontX" and "soupX". Default to NULL.
seed	Seed for the random number generator. Default 12345.
paramsList	A list containing parameters for QC functions. Default NULL.

Value

SingleCellExperiment object containing the outputs of the specified algorithms in the colData of inSCE.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
## Not run:
sce <- runCellQC(sce)

## End(Not run)
```

---

runClusterSummaryMetrics

Run Cluster Summary Metrics

Description

Calculates the mean expression of percent of cells that express the given genes for each cluster

Usage

```
runClusterSummaryMetrics(
  inSCE,
  useAssay = "logcounts",
  featureNames,
  displayName = NULL,
  groupNames = "cluster",
  scale = FALSE
)
```

Arguments

inSCE	The single cell experiment to use.
useAssay	The assay to use.
featureNames	A string or vector of strings with each gene to aggregate.
displayName	A string that is the name of the column used for genes.
groupNames	The name of a colData entry that can be used as groupNames.
scale	Option to scale the data. Default: /codeFALSE. Selected assay will not be scaled.

Value

A dataframe with mean expression and percent of cells in cluster that express for each cluster.

Examples

```
data("scExample")
runClusterSummaryMetrics(inSCE=sce, useAssay="counts", featureNames=c("B2M", "MALAT1"),
displayName="feature_name", groupNames="type")
```

`runComBatSeq` *Apply ComBat-Seq batch effect correction method to SingleCellExperiment object*

Description

The ComBat-Seq batch adjustment approach assumes that batch effects represent non-biological but systematic shifts in the mean or variability of genomic features for all samples within a processing batch. It uses either parametric or non-parametric empirical Bayes frameworks for adjusting data for batch effects.

Usage

```
runComBatSeq(
  inSCE,
  useAssay = "counts",
  batch = "batch",
  covariates = NULL,
  bioCond = NULL,
  useSVA = FALSE,
  assayName = "ComBatSeq",
  shrink = FALSE,
  shrinkDisp = FALSE,
  nGene = NULL
)
```

Arguments

`inSCE`	Input SingleCellExperiment object
`useAssay`	A single character indicating the name of the assay requiring batch correction. Default `"counts"`.
`batch`	A single character indicating a field in `colData` that annotates the batches. Default `"batch"`.
`covariates`	A character vector indicating the fields in `colData` that annotates other covariates, such as the cell types. Default `NULL`.
`bioCond`	A single character indicating a field in `colData` that annotates the biological conditions. Default `NULL`.
`useSVA`	A logical scalar. Whether to estimate surrogate variables and use them as an empirical control. Default `FALSE`.

assayName	A single characeter. The name for the corrected assay. Will be saved to assay. Default "ComBat".
shrink	A logical scalar. Whether to apply shrinkage on parameter estimation. Default FALSE.
shrinkDisp	A logical scalar. Whether to apply shrinkage on dispersion. Default FALSE.
nGene	An integer. Number of random genes to use in empirical Bayes estimation, only useful when shrink is set to TRUE. Default NULL.

Details

For the parameters covariates and useSVA, when the cell type information is known, it is recommended to specify the cell type annotation to the argument covariates; if the cell types are unknown but expected to be balanced, it is recommended to run with default settings, yet informative covariates could still be useful. If the cell types are unknown and are expected to be unbalanced, it is recommended to set useSVA to TRUE.

Value

The input SingleCellExperiment object with assay(inSCE, assayName) updated.

Examples

```
data('sceBatches', package = 'singleCellTK')
sceBatches <- sample(sceBatches, 40)
# Cell type known
sceBatches <- runComBatSeq(sceBatches, "counts", "batch",
                           covariates = "cell_type",
                           assayName = "ComBat_cell_seq")
# Cell type unknown but balanced
#sceBatches <- runComBatSeq(sceBatches, "counts", "batch",
#                           assayName = "ComBat_seq")
# Cell type unknown and unbalanced
#sceBatches <- runComBatSeq(sceBatches, "counts", "batch",
#                           useSVA = TRUE,
#                           assayName = "ComBat_sva_seq")
```

---

runCxds *Find doublets/multiplets using cxds.*

Description

A wrapper function for cxds. Annotate doublets/multiplets using co-expression based approach. Generate a doublet score for each cell. Infer doublets if estNdbl is TRUE.

Usage

```
runCxds(
  inSCE,
  sample = NULL,
  seed = 12345,
  ntop = 500,
  binThresh = 0,
  verb = FALSE,
  retRes = FALSE,
  estNdbl = FALSE,
  useAssay = "counts"
)
```

Arguments

inSCE	A SingleCellExperiment object.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
seed	Seed for the random number generator, can be NULL. Default 12345.
ntop	See cxds for more information. Default 500.
binThresh	See cxds for more information. Default 0.
verb	See cxds for more information. Default FALSE.
retRes	See cxds for more information. Default FALSE.
estNdbl	See cxds for more information. Default FALSE.
useAssay	A string specifying which assay in the SCE to use. Default "counts"

Details

When the argument sample is specified, cxds will be run on cells from each sample separately. If sample = NULL, then all cells will be processed together.

Value

A SingleCellExperiment object with cxds output appended to the colData slot. The columns include *cxds_score* and optionally *cxds_call*.

See Also

cxds, plotCxdsResults, runCellQC

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runCxds(sce)
```

runCxdsBcdsHybrid *Find doublets/multiplets using cxds_bcds_hybrid.*

Description

A wrapper function for cxds_bcds_hybrid. Annotate doublets/multiplets using a binary classification approach to discriminate artificial doublets from original data. Generate a doublet score for each cell. Infer doublets if `estNdbl` is `TRUE`.

Usage

```
runCxdsBcdsHybrid(
  inSCE,
  sample = NULL,
  seed = 12345,
  nTop = 500,
  cxdsArgs = list(),
  bcdsArgs = list(),
  verb = FALSE,
  estNdbl = FALSE,
  force = FALSE,
  useAssay = "counts"
)
```

Arguments

`inSCE`	A SingleCellExperiment object. Needs `counts` in assays slot.
`sample`	Character vector. Indicates which sample each cell belongs to. cxds_bcds_hybrid will be run on cells from each sample separately. If NULL, then all cells will be processed together. Default NULL.
`seed`	Seed for the random number generator. Default 12345.
`nTop`	The number of top varialbe genes to consider. Used in both `csds` and `bcds`. Default `500`.
`cxdsArgs`	See cxds_bcds_hybrid for more information. Default `NULL`.
`bcdsArgs`	See cxds_bcds_hybrid for more information. Default `NULL`.
`verb`	See cxds_bcds_hybrid for more information. Default `FALSE`.
`estNdbl`	See cxds_bcds_hybrid for more information. Default `FALSE`.
`force`	See cxds_bcds_hybrid for more information. Default `FALSE`.
`useAssay`	A string specifying which assay in the SCE to use.

Value

A SingleCellExperiment object with cxds_bcds_hybrid output appended to the colData slot. The columns include *hybrid_score* and optionally *hybrid_call*. Please refer to the documentation of cxds_bcds_hybrid for details.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runCxdsBcdsHybrid(sce)
```

`runDEAnalysis` *Perform differential expression analysis on SCE object*

Description

Perform differential expression analysis on SCE object

Usage

```
runDEAnalysis(inSCE, method = "wilcox", ...)

runDESeq2(
  inSCE,
  useAssay = "counts",
  useReducedDim = NULL,
  index1 = NULL,
  index2 = NULL,
  class = NULL,
  classGroup1 = NULL,
  classGroup2 = NULL,
  analysisName,
  groupName1,
  groupName2,
  covariates = NULL,
  fullReduced = TRUE,
  onlyPos = FALSE,
  log2fcThreshold = NULL,
  fdrThreshold = NULL,
  minGroup1MeanExp = NULL,
  maxGroup2MeanExp = NULL,
  minGroup1ExprPerc = NULL,
  maxGroup2ExprPerc = NULL,
  overwrite = FALSE,
  verbose = TRUE
)

runLimmaDE(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  index1 = NULL,
  index2 = NULL,
```

```
  class = NULL,
  classGroup1 = NULL,
  classGroup2 = NULL,
  analysisName,
  groupName1,
  groupName2,
  covariates = NULL,
  onlyPos = FALSE,
  log2fcThreshold = NULL,
  fdrThreshold = NULL,
  minGroup1MeanExp = NULL,
  maxGroup2MeanExp = NULL,
  minGroup1ExprPerc = NULL,
  maxGroup2ExprPerc = NULL,
  overwrite = FALSE,
  verbose = TRUE
)

runANOVA(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  index1 = NULL,
  index2 = NULL,
  class = NULL,
  classGroup1 = NULL,
  classGroup2 = NULL,
  analysisName,
  groupName1,
  groupName2,
  covariates = NULL,
  onlyPos = FALSE,
  log2fcThreshold = NULL,
  fdrThreshold = NULL,
  minGroup1MeanExp = NULL,
  maxGroup2MeanExp = NULL,
  minGroup1ExprPerc = NULL,
  maxGroup2ExprPerc = NULL,
  overwrite = FALSE,
  verbose = TRUE
)

runMAST(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  index1 = NULL,
  index2 = NULL,
```

```
  class = NULL,
  classGroup1 = NULL,
  classGroup2 = NULL,
  analysisName,
  groupName1,
  groupName2,
  covariates = NULL,
  onlyPos = FALSE,
  log2fcThreshold = NULL,
  fdrThreshold = NULL,
  minGroup1MeanExp = NULL,
  maxGroup2MeanExp = NULL,
  minGroup1ExprPerc = NULL,
  maxGroup2ExprPerc = NULL,
  overwrite = FALSE,
  check_sanity = TRUE,
  verbose = TRUE
)

runWilcox(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  index1 = NULL,
  index2 = NULL,
  class = "cluster",
  classGroup1 = c(1),
  classGroup2 = c(2),
  analysisName = "cluster1_VS_2",
  groupName1 = "cluster1",
  groupName2 = "cluster2",
  covariates = NULL,
  onlyPos = FALSE,
  log2fcThreshold = NULL,
  fdrThreshold = NULL,
  minGroup1MeanExp = NULL,
  maxGroup2MeanExp = NULL,
  minGroup1ExprPerc = NULL,
  maxGroup2ExprPerc = NULL,
  overwrite = FALSE,
  verbose = TRUE
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
method	Character. Specify which method to use when using `runDEAnalysis()`. Choose from `"wilcox"`, `"MAST"`, `"DESeq2"`, `"Limma"`, `"ANOVA"`. Default `"wilcox"`.

...	Arguments to pass to specific methods when using the generic runDEAnalysis().
useAssay	character. A string specifying which assay to use for the DE regression. Ignored when useReducedDim is specified. Default "counts" for DESeq2, "logcounts" for other methods.
useReducedDim	character. A string specifying which reducedDim to use for DE analysis. Will treat the dimensions as features. Default NULL.
index1	Any type of indices that can subset a SingleCellExperiment inherited object by cells. Specifies which cells are of interests. Default NULL.
index2	Any type of indices that can subset a SingleCellExperiment inherited object by cells. specifies the control group against those specified by index1. If NULL when using index specification, index1 cells will be compared with all other cells. Default NULL.
class	A vector/factor with ncol(inSCE) elements, or a character scalar that specifies a column name of colData(inSCE). Default "cluster".
classGroup1	a vector specifying which "levels" given in class are of interests. Default c(1).
classGroup2	a vector specifying which "levels" given in class is the control group against those specified by classGroup1. If NULL when using annotation specification, classGroup1 cells will be compared with all other cells. Default c(2).
analysisName	A character scalar naming the DEG analysis. Default "cluster1_VS_2".
groupName1	A character scalar naming the group of interests. Default "cluster1".
groupName2	A character scalar naming the control group. Default "cluster2".
covariates	A character vector of additional covariates to use when building the model. All covariates must exist in names(colData(inSCE)). Default NULL.
fullReduced	Logical, DESeq2 only argument. Whether to apply LRT (Likelihood ratio test) with a 'full' model. Default TRUE.
onlyPos	Whether to only output DEG with positive log2_FC value. Default FALSE.
log2fcThreshold	Only out put DEGs with the absolute values of log2FC greater than this value. Default NULL.
fdrThreshold	Only out put DEGs with FDR value less than this value. Default NULL.
minGroup1MeanExp	Only out put DEGs with mean expression in group1 greater then this value. Default NULL.
maxGroup2MeanExp	Only out put DEGs with mean expression in group2 less then this value. Default NULL.
minGroup1ExprPerc	Only out put DEGs expressed in greater then this fraction of cells in group1. Default NULL.
maxGroup2ExprPerc	Only out put DEGs expressed in less then this fraction of cells in group2. Default NULL.
overwrite	A logical scalar. Whether to overwrite result if exists. Default FALSE.
verbose	A logical scalar. Whether to show messages. Default TRUE.
check_sanity	Logical, MAST only argument. Whether to perform MAST's sanity check to see if the counts are logged. Default TRUE.

Details

SCTK provides Limma, MAST, DESeq2, ANOVA and Wilcoxon test for differential expression analysis, where DESeq2 expects non-negtive integer assay input while others expect logcounts.

Condition specification allows two methods: 1. Index level selection. Only use arguments `index1` and `index2`. 2. Annotation level selection. Only use arguments `class`, `classGroup1` and `classGroup2`.

Value

The input SingleCellExperiment object, where `metadata(inSCE)$diffExp` is updated with a list named by `analysisName`, with elements of:

`$groupNames`	the naming of the two conditions
`$useAssay`, `$useReducedDim`	the matrix name that was used for calculation
`$select`	the cell selection indices (logical) for each condition
`$result`	a `data.frame` of the DEGs table
`$method`	the method used

See Also

See `plotDEGHeatmap`, `plotDEGRegression`, `plotDEGViolin` and `plotDEGVolcano` for visualization method after running DE analysis.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- scaterlogNormCounts(sce, assayName = "logcounts")
sce <- runDEAnalysis(method = "Limma", inSCE = sce, groupName1 = "group1",
 groupName2 = "group2", index1 = seq(20), index2 = seq(21,40),
 analysisName = "Limma")
```

---

runDecontX *Detecting contamination with DecontX.*

Description

A wrapper function for decontX. Identify potential contamination from experimental factors such as ambient RNA.

Usage

```
runDecontX(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  background = NULL,
  bgAssayName = NULL,
  bgBatch = NULL,
  z = NULL,
  maxIter = 500,
  delta = c(10, 10),
  estimateDelta = TRUE,
  convergence = 0.001,
  iterLogLik = 10,
  varGenes = 5000,
  dbscanEps = 1,
  seed = 12345,
  logfile = NULL,
  verbose = TRUE
)
```

Arguments

inSCE	A SingleCellExperiment object.
sample	A single character specifying a name that can be found in colData(inSCE) to directly use the cell annotation; or a character vector with as many elements as cells to indicates which sample each cell belongs to. Default NULL. decontX will be run on cells from each sample separately.
useAssay	A string specifying which assay in the SCE to use. Default 'counts'.
background	A SingleCellExperiment with the matrix located in the assay slot under bgAssayName. It should have the same structure as inSCE except it contains the matrix of empty droplets instead of cells. When supplied, empirical distribution of transcripts from these empty droplets will be used as the contamination distribution. Default NULL.
bgAssayName	Character. Name of the assay to use if background is a SingleCellExperiment. If NULL, the function will use the same value as useAssay. Default is NULL.
bgBatch	Batch labels for background. If background is a SingleCellExperiment object, this can be a single character specifying a name that can be found in colData(background) to directly use the barcode annotation; or a numeric / character vector that has as many elements as barcodes to indicate which sample each barcode belongs to. Its unique values should be the same as those in sample, such that each batch of cells have their corresponding batch of empty droplets as background, pointed by this parameter. Default to NULL.
z	Numeric or character vector. Cell cluster labels. If NULL, PCA will be used to reduce the dimensionality of the dataset initially, 'umap' from the 'uwot' package will be used to further reduce the dataset to 2 dimenions and the 'dbscan'

function from the 'dbscan' package will be used to identify clusters of broad cell types. Default NULL.

`maxIter` Integer. Maximum iterations of the EM algorithm. Default 500.

`delta` Numeric Vector of length 2. Concentration parameters for the Dirichlet prior for the contamination in each cell. The first element is the prior for the native counts while the second element is the prior for the contamination counts. These essentially act as pseudocounts for the native and contamination in each cell. If `estimateDelta = TRUE`, this is only used to produce a random sample of proportions for an initial value of contamination in each cell. Then `fit_dirichlet` is used to update `delta` in each iteration. If `estimateDelta = FALSE`, then `delta` is fixed with these values for the entire inference procedure. Fixing `delta` and setting a high number in the second element will force `decontX` to be more aggressive and estimate higher levels of contamination at the expense of potentially removing native expression. Default `c(10, 10)`.

`estimateDelta` Boolean. Whether to update `delta` at each iteration.

`convergence` Numeric. The EM algorithm will be stopped if the maximum difference in the contamination estimates between the previous and current iterations is less than this. Default 0.001.

`iterLogLik` Integer. Calculate log likelihood every `iterLogLik` iteration. Default 10.

`varGenes` Integer. The number of variable genes to use in dimensionality reduction before clustering. Variability is calcualted using `modelGeneVar` function from the 'scran' package. Used only when z is not provided. Default 5000.

`dbscanEps` Numeric. The clustering resolution parameter used in 'dbscan' to estimate broad cell clusters. Used only when z is not provided. Default 1.

`seed` Integer. Passed to with_seed. For reproducibility, a default value of 12345 is used. If NULL, no calls to with_seed are made.

`logfile` Character. Messages will be redirected to a file named 'logfile'. If NULL, messages will be printed to stdout. Default NULL.

`verbose` Logical. Whether to print log messages. Default TRUE.

Value

A SingleCellExperiment object with 'decontX_Contamination' and 'decontX_Clusters' added to the colData slot. Additionally, the decontaminated counts will be added as an assay called 'decontXCounts'.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runDecontX(sce[,sample(ncol(sce),20)])
```

runDimReduce *Generic Wrapper function for running dimensionality reduction*

Description

Generic Wrapper function for running dimensionality reduction

Usage

```
runDimReduce(
  inSCE,
 method = c("scaterPCA", "seuratPCA", "seuratICA", "scanpyPCA", "rTSNE", "seuratTSNE",
    "scaterUMAP", "seuratUMAP", "scanpyUMAP", "scanpyTSNE"),
  useAssay = NULL,
  useReducedDim = NULL,
  useAltExp = NULL,
  reducedDimName = method,
  nComponents = 20,
  useFeatureSubset = NULL,
  scale = FALSE,
  seed = 12345,
  ...
)
```

Arguments

inSCE	Input SingleCellExperiment object.
method	One from "scaterPCA", "seuratPCA", "seuratICA", "rTSNE", "seuratTSNE", "scaterUMAP", "seuratUMAP", "scanpyPCA", "scanpyUMAP" and "scanpyTSNE".
useAssay	Assay to use for computation. If useAltExp is specified, useAssay has to exist in assays(altExp(inSCE, useAltExp)). Default "counts".
useReducedDim	The low dimension representation to use for embedding computation. Default NULL.
useAltExp	The subset to use for computation, usually for the selected variable features. Default NULL.
reducedDimName	The name of the result matrix. Required.
nComponents	Specify the number of dimensions to compute with the selected method in case of PCA/ICA and the number of components to use in the case of TSNE/UMAP methods.
useFeatureSubset	Subset of feature to use for dimension reduction. A character string indicating a rowData variable that stores the logical vector of HVG selection, or a vector that can subset the rows of inSCE. Default NULL.
scale	Logical scalar, whether to standardize the expression values. Default TRUE.

seed	Random seed for reproducibility of results. Default NULL will use global seed in use by the R environment.
...	The other arguments for running a specific algorithm. Please refer to the one you use.

Details

Wrapper function to run one of the available dimensionality reduction algorithms integrated within SCTK from scaterPCA, runSeuratPCA, runSeuratICA, runTSNE, runSeuratTSNE, runUMAP and runSeuratUMAP. Users can use an assay by specifying useAssay, use the assay in an altExp by specifying both useAltExp and useAssay, or use a low-dimensionality representation by specifying useReducedDim.

Value

The input SingleCellExperiment object with reducedDim updated with the result.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runNormalization(sce, useAssay = "counts",
                        outAssayName = "logcounts",
                        normalizationMethod = "logNormCounts")
sce <- runDimReduce(inSCE = sce, method = "scaterPCA",
                    useAssay = "logcounts", scale = TRUE,
                    reducedDimName = "PCA")
```

---

runDoubletFinder *Generates a doublet score for each cell via doubletFinder*

Description

Uses doubletFinder to determine cells within the dataset suspected to be doublets.

Usage

```
runDoubletFinder(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  seed = 12345,
  seuratNfeatures = 2000,
  seuratPcs = seq(15),
  seuratRes = 1.5,
  formationRate = 0.075,
  nCores = NULL,
  verbose = FALSE
)
```

Arguments

inSCE	inSCE A SingleCellExperiment object.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
useAssay	A string specifying which assay in the SCE to use. Default "counts".
seed	Seed for the random number generator, can be set to NULL. Default 12345.
seuratNfeatures	Integer. Number of highly variable genes to use. Default 2000.
seuratPcs	Numeric vector. The PCs used in seurat function to determine number of clusters. Default 1:15.
seuratRes	Numeric vector. The resolution parameter used in Seurat, which adjusts the number of clusters determined via the algorithm. Default 1.5.
formationRate	Doublet formation rate used within algorithm. Default 0.075.
nCores	Number of cores used for running the function. Default NULL.
verbose	Boolean. Wheter to print messages from Seurat and DoubletFinder. Default FALSE.

Value

SingleCellExperiment object containing the doublet_finder_doublet_score variable in colData slot.

See Also

runCellQC, plotDoubletFinderResults

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runDoubletFinder(sce)
```

---

runDropletQC *Perform comprehensive droplet QC*

Description

A wrapper function to run several QC algorithms for determining empty droplets in single cell RNA-seq data

Usage

```
runDropletQC(
  inSCE,
  algorithms = c("QCMetrics", "emptyDrops", "barcodeRanks"),
  sample = NULL,
  useAssay = "counts",
  paramsList = NULL
)
```

Arguments

inSCE	A SingleCellExperiment object containing the full droplet count matrix
algorithms	Character vector. Specify which QC algorithms to run. Available options are "emptyDrops" and "barcodeRanks".
sample	Character vector. Indicates which sample each cell belongs to. Algorithms will be run on cells from each sample separately.
useAssay	A string specifying which assay contains the count matrix for droplets.
paramsList	A list containing parameters for QC functions. Default NULL.

Value

SingleCellExperiment object containing the outputs of the specified algorithms in the colData of inSCE.

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runDropletQC(sce)

## End(Not run)
```

---

runEmptyDrops *Identify empty droplets using emptyDrops.*

Description

Run emptyDrops on the count matrix in the provided \linkS4classSingleCellExperiment object. Distinguish between droplets containing cells and ambient RNA in a droplet-based single-cell RNA sequencing experiment.

Usage

```
runEmptyDrops(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  lower = 100,
  niters = 10000,
  testAmbient = FALSE,
  ignore = NULL,
  alpha = NULL,
  retain = NULL,
  barcodeArgs = list(),
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

inSCE	A SingleCellExperiment object. Must contain a raw counts matrix before empty droplets have been removed.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
useAssay	A string specifying which assay in the SCE to use. Default "counts"
lower	See emptyDrops for more information. Default 100.
niters	See emptyDrops for more information. Default 10000.
testAmbient	See emptyDrops for more information. Default FALSE.
ignore	See emptyDrops for more information. Default NULL.
alpha	See emptyDrops for more information. Default NULL.
retain	See emptyDrops for more information. Default NULL.
barcodeArgs	See emptyDrops for more information. Default list().
BPPARAM	See emptyDrops for more information. Default BiocParallel::SerialParam().

Value

A SingleCellExperiment object with the emptyDrops output table appended to the colData slot. The columns include emptyDrops_total, emptyDrops_logprob, emptyDrops_pvalue, emptyDrops_limited, emptyDrops_fdr. Please refer to the documentation of emptyDrops for details.

See Also

runDropletQC, plotEmptyDropsResults, plotEmptyDropsScatter

Examples

```
data(scExample, package = "singleCellTK")
sce <- runEmptyDrops(inSCE = sce)
```

`runEnrichR` *Run EnrichR on SCE object*

Description

Run EnrichR on SCE object

Usage

```
runEnrichR(
  inSCE,
  features,
  analysisName,
  db = NULL,
  by = "rownames",
  featureName = NULL
)
```

Arguments

`inSCE`	A SingleCellExperiment object.
`features`	Character vector, selected genes for enrichment analysis.
`analysisName`	A string that identifies each specific analysis.
`db`	Character vector. Selected database name(s) from the enrichR database list. If `NULL` then EnrichR will be run on all the available databases on the enrichR database. See details. Default `NULL`
`by`	Character. From where should we find the `features`? `"rownames"` for from `rownames(inSCE)`, otherwise, from a column of feature metadata (`rowData(inSCE)[[by]]`). See details. Default `"rownames"`.
`featureName`	Character. Indicates the actual feature identifiers to be passed to EnrichR. Can be `"rownames"`, a column in feature metadata (`rowData(inSCE)[[featureName]]`), or a character vector with its length equals to `nrow(inSCE)`. See details. Default `"rownames"`.

Details

EnrichR works by querying the specified `features` to its online databases, thus it requires the Internet connection.

Available db options could be shown by running `enrichR::listEnrichrDbs()$libraryName`

This function checks for the existence of features in the SCE object. When `features` do not have a match in `rownames(inSCE)`, users may try to specify by to pass the check.

EnrichR expects gene symbols/names as the input (i.e. Ensembl ID might not work). When specified `features` are not qualified for this, users may try to specify `featureName` to change the identifier type to pass to EnrichR.

Value

Updates inSCE metadata with a data.frame of enrichment terms overlapping in the respective databases along with p-values, z-scores etc.

See Also

getEnrichRResult

Examples

```
data("mouseBrainSubsetSCE")
if (Biobase::testBioCConnection()) {
  mouseBrainSubsetSCE <- runEnrichR(mouseBrainSubsetSCE, features = "Cmtm5",
                                    db = "GO_Cellular_Component_2017",
                                    analysisName = "analysis1")
}
```

---

runFastMNN *Apply a fast version of the mutual nearest neighbors (MNN) batch effect correction method to SingleCellExperiment object*

Description

fastMNN is a variant of the classic MNN method, modified for speed and more robust performance. For introduction of MNN, see runMNNCorrect.

Usage

```
runFastMNN(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  batch = "batch",
  reducedDimName = "fastMNN",
  k = 20,
  propK = NULL,
  ndist = 3,
  minBatchSkip = 0,
  cosNorm = TRUE,
  nComponents = 50,
  weights = NULL,
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

inSCE	Input SingleCellExperiment object
useAssay	A single character indicating the name of the assay requiring batch correction. Default "logcounts".
useReducedDim	A single character indicating the dimension reduction used for batch correction. Will ignore useAssay when using. Default NULL.
batch	A single character indicating a field in colData that annotates the batches of each cell; or a vector/factor with the same length as the number of cells. Default "batch".
reducedDimName	A single character. The name for the corrected low-dimensional representation. Default "fastMNN".
k	An integer scalar specifying the number of nearest neighbors to consider when identifying MNNs. See "See Also". Default 20.
propK	A numeric scalar in (0, 1) specifying the proportion of cells in each dataset to use for mutual nearest neighbor searching. See "See Also". Default NULL.
ndist	A numeric scalar specifying the threshold beyond which neighbours are to be ignored when computing correction vectors. See "See Also". Default 3.
minBatchSkip	Numeric scalar specifying the minimum relative magnitude of the batch effect, below which no correction will be performed at a given merge step. See "See Also". Default 0.
cosNorm	A logical scalar indicating whether cosine normalization should be performed on useAssay prior to PCA. See "See Also". Default TRUE.
nComponents	An integer scalar specifying the number of dimensions to produce. See "See Also". Default 50.
weights	The weighting scheme to use. Passed to multiBatchPCA. Default NULL.
BPPARAM	A BiocParallelParam object specifying whether the SVD should be parallelized.

Value

The input SingleCellExperiment object with reducedDim(inSCE, reducedDimName) updated.

References

Lun ATL, et al., 2016

See Also

fastMNN for using useAssay, and reducedMNN for using useReducedDim

Examples

```
data('sceBatches', package = 'singleCellTK')
logcounts(sceBatches) <- log1p(counts(sceBatches))
sceCorr <- runFastMNN(sceBatches, useAssay = 'logcounts')
```

runFeatureSelection *Run Variable Feature Detection Methods*

Description

Wrapper function to run all of the feature selection methods integrated within the singleCellTK package including three methods from Seurat ("vst", "mean.var.plot" or dispersion) and the Scran modelGeneVar method.

This function does not return the names of the variable features but only computes the metrics, which will be stored in the rowData slot. To set a HVG list for downstream use, users should call setTopHVG after computing the metrics. To get the names of the variable features, users should call getTopHVG function after computing the metrics.

Usage

```
runFeatureSelection(inSCE, useAssay, method = "vst")
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Specify the name of the assay that should be used. Should use raw counts for "vst" method, or a normalized assay for other methods.
method	Specify the method to use for variable gene selection. Options include "vst", "mean.var.plot" or "dispersion" from Seurat and "modelGeneVar" from Scran. Default "vst"

Value

The input SingleCellExperiment object that contains the computed statistics in the rowData slot

See Also

runModelGeneVar, runSeuratFindHVG, getTopHVG, plotTopHVG

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runFeatureSelection(mouseBrainSubsetSCE,
                                           "logcounts",
                                           "modelGeneVar")
```

runFindMarker *Find the marker gene set for each cluster*

Description

With an input SingleCellExperiment object and specifying the clustering labels, this function iteratively call the differential expression analysis on each cluster against all the others. runFindMarker will be deprecated in the future.

Usage

```
runFindMarker(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  method = "wilcox",
  cluster = "cluster",
  covariates = NULL,
  log2fcThreshold = NULL,
  fdrThreshold = 0.05,
  minClustExprPerc = NULL,
  maxCtrlExprPerc = NULL,
  minMeanExpr = NULL,
  detectThresh = 0
)

findMarkerDiffExp(
  inSCE,
  useAssay = "logcounts",
  useReducedDim = NULL,
  method = c("wilcox", "MAST", "DESeq2", "Limma", "ANOVA"),
  cluster = "cluster",
  covariates = NULL,
  log2fcThreshold = NULL,
  fdrThreshold = 0.05,
  minClustExprPerc = NULL,
  maxCtrlExprPerc = NULL,
  minMeanExpr = NULL,
  detectThresh = 0
)
```

Arguments

inSCE	SingleCellExperiment inherited object.
useAssay	character. A string specifying which assay to use for the MAST calculations. Default "logcounts".

useReducedDim character. A string specifying which reducedDim to use for MAST calculations. Set useAssay to NULL when using. Required.

method A single character for specific differential expression analysis method. Choose from 'wilcox', 'MAST', 'DESeq2', 'Limma', and 'ANOVA'. Default "wilcox".

cluster One single character to specify a column in colData(inSCE) for the clustering label. Alternatively, a vector or a factor is also acceptable. Default "cluster".

covariates A character vector of additional covariates to use when building the model. All covariates must exist in names(colData(inSCE)). Not applicable when method is "MAST" method. Default NULL.

log2fcThreshold
Only out put DEGs with the absolute values of log2FC larger than this value. Default NULL

fdrThreshold Only out put DEGs with FDR value smaller than this value. Default NULL

minClustExprPerc
A numeric scalar. The minimum cutoff of the percentage of cells in the cluster of interests that expressed the marker gene. From 0 to 1. Default NULL.

maxCtrlExprPerc
A numeric scalar. The maximum cutoff of the percentage of cells out of the cluster (control group) that expressed the marker gene. From 0 to 1. Default NULL.

minMeanExpr A numeric scalar. The minimum cutoff of the mean expression value of the marker in the cluster of interests. Default NULL.

detectThresh A numeric scalar, above which a matrix value will be treated as expressed when calculating cluster/control expression percentage. Default 0.

Details

The returned marker table, in the metadata slot, consists of 8 columns: "Gene", "Log2_FC", "Pvalue", "FDR", cluster, "clusterExprPerc", "ControlExprPerc" and "clusterAveExpr".

"clusterExprPerc" is the fraction of cells, that has marker value (e.g. gene expression counts) larger than detectThresh, in the cell population of the cluster. As for each cluster, we set all cells out of this cluster as control. Similarly, "ControlExprPerc" is the fraction of cells with marker value larger than detectThresh in the control cell group.

Value

The input SingleCellExperiment object with metadata(inSCE)$findMarker updated with a data.table of the up- regulated DEGs for each cluster.

See Also

runDEAnalysis, getFindMarkerTopTable, plotFindMarkerHeatmap

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runFindMarker(mouseBrainSubsetSCE,
                                     useAssay = "logcounts",
                                     cluster = "level1class")
```

---

runGSVA *Run GSVA analysis on a SingleCellExperiment object*

Description

Run GSVA analysis on a SingleCellExperiment object

Usage

```
runGSVA(
  inSCE,
  useAssay = "logcounts",
  resultNamePrefix = NULL,
  geneSetCollectionName,
  ...
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Indicate which assay to use. The default is "logcounts"
resultNamePrefix	Character. Prefix to the name the GSVA results which will be stored in the reducedDim slot of inSCE. The names of the output matrix will be resultNamePrefix_Scores. If this parameter is set to NULL, then "GSVA_geneSetCollectionName_" will be used. Default NULL.
geneSetCollectionName	Character. The name of the gene set collection to use.
...	Parameters to pass to gsva()

Value

A SingleCellExperiment object with pathway activity scores from GSVA stored in reducedDim as GSVA_geneSetCollectionName_Scores.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- scaterlogNormCounts(sce, assayName = "logcounts")
gs1 <- rownames(sce)[seq(10)]
gs2 <- rownames(sce)[seq(11,20)]
gs <- list("geneset1" = gs1, "geneset2" = gs2)

sce <- importGeneSetsFromList(inSCE = sce,geneSetList = gs,
                                          by = "rownames")
sce <- runGSVA(inSCE = sce,
               geneSetCollectionName = "GeneSetCollection",
               useAssay = "logcounts")
```

---

runHarmony *Apply Harmony batch effect correction method to SingleCellExperiment object*

Description

Harmony is an algorithm that projects cells into a shared embedding in which cells group by cell type rather than dataset-specific conditions.

Usage

```
runHarmony(
  inSCE,
  useAssay = NULL,
  useReducedDim = NULL,
  batch = "batch",
  reducedDimName = "HARMONY",
  nComponents = 50,
  lambda = 0.1,
  theta = 5,
  sigma = 0.1,
  nIter = 10,
  seed = 12345,
  verbose = TRUE,
  ...
)
```

Arguments

inSCE	Input SingleCellExperiment object
useAssay	A single character indicating the name of the assay requiring batch correction. Default NULL. It is recommended to use a reducedDim such as PCA through the 'useReducedDim' parameter of this function.

useReducedDim	A single character indicating the name of the reducedDim to be used. It is recommended to use a reducedDim instead of a full assay as using an assay might cause the algorithm to not converge and throw error. Specifying this will ignore useAssay. Default NULL.
batch	A single character indicating a field in colData that annotates the batches of each cell; or a vector/factor with the same length as the number of cells. Default "batch".
reducedDimName	A single character. The name for the corrected low-dimensional representation. Will be saved to reducedDim(inSCE). Default "HARMONY".
nComponents	An integer. The number of PCs to use and generate. Default 50L.
lambda	A Numeric scalar. Ridge regression penalty parameter. Must be strictly positive. Smaller values result in more aggressive correction. Default 0.1.
theta	A Numeric scalar. Diversity clustering penalty parameter. Larger values of theta result in more diverse clusters. theta=0 does not encourage any diversity. Default 5.
sigma	A Numeric scalar. Width of soft kmeans clusters. Larger values of sigma result in cells assigned to more clusters. Smaller values of sigma make soft kmeans cluster approach hard clustering. Default 0.1.
nIter	An integer. The max number of iterations to perform. Default 10L.
seed	Set seed for reproducibility. Default is 12345.
verbose	Whether to print progress messages. Default TRUE.
...	Other arguments passed to HarmonyMatrix. See details.

Details

Since some of the arguments of HarmonyMatrix is controlled by this wrapper function. The additional arguments users can work with only include: nclust, tau, block.size, max.iter.cluster, epsilon.cluster, epsilon.harmony, plot_convergence, reference_values and cluster_prior.

Value

The input SingleCellExperiment object with reducedDim(inSCE, reducedDimName) updated.

References

Ilya Korsunsky, et al., 2019

Examples

```
data('sceBatches', package = 'singleCellTK')
logcounts(sceBatches) <- log1p(counts(sceBatches))
## Not run:
if (require("harmony"))
    sceCorr <- runHarmony(sceBatches)

## End(Not run)
```

runKMeans *Get clustering with KMeans*

Description

Perform KMeans clustering on a SingleCellExperiment object, with kmeans.

Usage

```
runKMeans(
  inSCE,
  nCenters,
  useReducedDim = "PCA",
  clusterName = "KMeans_cluster",
  nComp = 10,
  nIter = 10,
  nStart = 1,
  seed = 12345,
  algorithm = c("Hartigan-Wong", "Lloyd", "MacQueen")
)
```

Arguments

inSCE	A SingleCellExperiment object.
nCenters	An integer, the number of centroids (clusters).
useReducedDim	A single character, specifying which low-dimension representation to perform the clustering algorithm on. Default "PCA".
clusterName	A single character, specifying the name to store the cluster label in colData. Default "KMeans_cluster".
nComp	An integer. The number of components to use for K-Means. Default 10. See Detail.
nIter	An integer, the maximum number of iterations allowed. Default 10.
nStart	An integer, the number of random sets to choose. Default 1.
seed	An integer. The seed for the random number generator. Default 12345.
algorithm	A single character. Choose from "Hartigan-Wong", "Lloyd", "MacQueen". May be abbreviated. Default "Hartigan-Wong".

Value

The input SingleCellExperiment object with factor cluster labeling updated in colData(inSCE)[[clusterName]].

Examples

```
data("mouseBrainSubsetSCE")
mouseBrainSubsetSCE <- runKMeans(mouseBrainSubsetSCE,
                                 useReducedDim = "PCA_logcounts",
                                 nCenters = 2)
```

runLimmaBC *Apply Limma's batch effect correction method to SingleCellExperiment object*

Description

Limma's batch effect removal function fits a linear model to the data, then removes the component due to the batch effects.

Usage

```
runLimmaBC(inSCE, useAssay = "logcounts", assayName = "LIMMA", batch = "batch")
```

Arguments

inSCE	Input SingleCellExperiment object
useAssay	A single character indicating the name of the assay requiring batch correction. Default "logcounts".
assayName	A single characeter. The name for the corrected assay. Will be saved to assay. Default "LIMMA".
batch	A single character indicating a field in colData that annotates the batches of each cell; or a vector/factor with the same length as the number of cells. Default "batch".

Value

The input SingleCellExperiment object with assay(inSCE, assayName) updated.

References

Gordon K Smyth, et al., 2003

Examples

```
data('sceBatches', package = 'singleCellTK')
logcounts(sceBatches) <- log1p(counts(sceBatches))
sceCorr <- runLimmaBC(sceBatches)
```

runMNNCorrect *Apply the mutual nearest neighbors (MNN) batch effect correction method to SingleCellExperiment object*

Description

MNN is designed for batch correction of single-cell RNA-seq data where the batches are partially confounded with biological conditions of interest. It does so by identifying pairs of MNN in the high-dimensional log-expression space. For each MNN pair, a pairwise correction vector is computed by applying a Gaussian smoothing kernel with bandwidth 'sigma'.

Usage

```
runMNNCorrect(
  inSCE,
  useAssay = "logcounts",
  batch = "batch",
  assayName = "MNN",
  k = 20L,
  propK = NULL,
  sigma = 0.1,
  cosNormIn = TRUE,
  cosNormOut = TRUE,
  varAdj = TRUE,
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

inSCE	Input SingleCellExperiment object
useAssay	A single character indicating the name of the assay requiring batch correction. Default "logcounts".
batch	A single character indicating a field in colData that annotates the batches of each cell; or a vector/factor with the same length as the number of cells. Default "batch".
assayName	A single characeter. The name for the corrected assay. Will be saved to assay. Default "MNN".
k	An integer scalar specifying the number of nearest neighbors to consider when identifying MNNs. See "See Also". Default 20.
propK	A numeric scalar in (0, 1) specifying the proportion of cells in each dataset to use for mutual nearest neighbor searching. See "See Also". Default NULL.
sigma	A numeric scalar specifying the bandwidth of the Gaussian smoothing kernel used to compute the correction vector for each cell. See "See Also". Default 0.1.

cosNormIn	A logical scalar indicating whether cosine normalization should be performed on the input data prior to calculating distances between cells. See "See Also". Default TRUE.
cosNormOut	A logical scalar indicating whether cosine normalization should be performed prior to computing corrected expression values. See "See Also". Default TRUE.
varAdj	A logical scalar indicating whether variance adjustment should be performed on the correction vectors. See "See Also". Default TRUE.
BPPARAM	A BiocParallelParam object specifying whether the PCA and nearest-neighbor searches should be parallelized.

Value

The input SingleCellExperiment object with `assay(inSCE, assayName)` updated.

References

Haghverdi L, Lun ATL, et. al., 2018

See Also

mnnCorrect

Examples

```
data('sceBatches', package = 'singleCellTK')
logcounts(sceBatches) <- log1p(counts(sceBatches))
sceCorr <- runMNNCorrect(sceBatches)
```

---

runModelGeneVar *Calculate Variable Genes with Scran modelGeneVar*

Description

Generates and stores variability data in the input SingleCellExperiment object, using modelGeneVar method.

Also selects a specified number of top HVGs and store the logical selection in rowData.

Usage

```
runModelGeneVar(inSCE, useAssay = "logcounts")
```

Arguments

inSCE	A SingleCellExperiment object
useAssay	A character string to specify an assay to compute variable features from. Default "logcounts".

Value

inSCE updated with variable feature metrics in rowData

Author(s)

Irzam Sarfraz

See Also

runFeatureSelection, runSeuratFindHVG, getTopHVG, plotTopHVG

Examples

```
data("scExample", package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- scaterlogNormCounts(sce, "logcounts")
sce <- runModelGeneVar(sce)
hvf <- getTopHVG(sce, method = "modelGeneVar", hvgNumber = 10,
          useFeatureSubset = NULL)
```

---

runNormalization *Run normalization/transformation with various methods*

Description

Wrapper function to run any of the integrated normalization/transformation methods in the singleCellTK. The available methods include 'LogNormalize', 'CLR', 'RC' and 'SCTransform' from Seurat, 'logNormCounts and 'CPM' from Scater. Additionally, users can 'scale' using Z.Score, 'transform' using log, log1p and sqrt, add 'pseudocounts' and trim the final matrices between a range of values.

Usage

```
runNormalization(
  inSCE,
  useAssay = "counts",
  outAssayName = "logcounts",
  normalizationMethod = "logNormCounts",
  scale = FALSE,
  seuratScaleFactor = 10000,
  transformation = NULL,
  pseudocountsBeforeNorm = NULL,
  pseudocountsBeforeTransform = NULL,
  trim = NULL,
  verbose = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Specify the name of the assay that should be used.
outAssayName	Specify the name of the new output assay.
normalizationMethod	Specify a normalization method from 'LogNormalize', 'CLR', 'RC' and 'SCTransform' from Seurat or 'logNormCounts' and 'CPM' from scater packages. Default NULL is set which will not run any normalization method.
scale	Logical value indicating if the data should be scaled using Z.Score. Default FALSE.
seuratScaleFactor	Specify the 'scaleFactor' argument if a Seurat normalization method is selected. Default is 10000. This parameter will not be used if methods other than seurat are selected.
transformation	Specify the transformation options to run on the selected assay. Options include 'log2' (base 2 log transformation), 'log1p' (natural log + 1 transformation) and 'sqrt' (square root). Default value is NULL, which will not run any transformation.
pseudocountsBeforeNorm	Specify a numeric pseudo value that should be added to the assay before normalization is performed. Default is NULL, which will not add any value.
pseudocountsBeforeTransform	Specify a numeric pseudo value that should be added to the assay before transformation is run. Default is NULL, which will not add any value.
trim	Specify a vector of two numeric values that should be used as the upper and lower trim values to trim the assay between these two values. For example, c(10,-10) will trim the values between 10 and -10. Default is NULL, which will not trim the data assay.
verbose	Logical value indicating if progress messages should be displayed to the user. Default is TRUE.

Value

Output SCE object with new normalized/transformed assay stored.

Examples

```
data(sce_chcl, package = "scds")
sce_chcl <- runNormalization(
 inSCE = sce_chcl,
 normalizationMethod = "LogNormalize",
 useAssay = "counts",
 outAssayName = "logcounts")
```

runPerCellQC *Wrapper for calculating QC metrics with scater.*

Description

A wrapper function for addPerCellQC. Calculate general quality control metrics for each cell in the count matrix.

Usage

```
runPerCellQC(
  inSCE,
  useAssay = "counts",
  mitoGeneLocation = "rownames",
  mitoRef = c(NULL, "human", "mouse"),
  mitoIDType = c("ensembl", "symbol", "entrez", "ensemblTranscriptID"),
  mitoPrefix = "MT-",
  mitoID = NULL,
  collectionName = NULL,
  geneSetList = NULL,
  geneSetListLocation = "rownames",
  geneSetCollection = NULL,
  percent_top = c(50, 100, 200, 500),
  use_altexps = FALSE,
  flatten = TRUE,
  detectionLimit = 0,
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

inSCE	A SingleCellExperiment object.
useAssay	A string specifying which assay in the SCE to use. Default "counts".
mitoGeneLocation	Character. Describes the location within inSCE where the gene identifiers in the mitochondrial gene sets should be located. If set to "rownames" then the features will be searched for among rownames(inSCE). This can also be set to one of the column names of rowData(inSCE) in which case the gene identifies will be mapped to that column in the rowData of inSCE. See featureIndex for more information. If this parameter is set to NULL, then no mitochondrial metrics will be calculated. Default "rownames".
mitoRef	Character. The species used to extract mitochondrial genes ID from build-in mitochondrial geneset in SCTK. Available species options are "human" and "mouse". Default is "human".
mitoIDType	Character. Types of mitochondrial gene id. SCTK supports "symbol", "entrez", "ensembl" and "ensemblTranscriptID". It is used with mitoRef to extract mitochondrial genes from build-in mitochondrial geneset in SCTK. Default NULL.

mitoPrefix
: Character. The prefix used to get mitochondrial gene from either rownames(inSCE) or columns of rowData(inSCE) specified by mitoGeneLocation. This parameter is usually used to extract mitochondrial genes from the gene symbol. For example, mitoPrefix = "^MT-" can be used to detect mito gene symbols like "MT-ND4". Note that case is ignored so "mt-" will still match "MT-ND4". Default "^MT-".

mitoID
: Character. A vector of mitochondrial genes to be quantified.

collectionName
: Character. Name of a GeneSetCollection obtained by using one of the importGeneSet* functions. Default NULL.

geneSetList
: List of gene sets to be quantified. The genes in the assays will be matched to the genes in the list based on geneSetListLocation. Default NULL.

geneSetListLocation
: Character or numeric vector. If set to 'rownames', then the genes in geneSetList will be looked up in rownames(inSCE). If another character is supplied, then genes will be looked up in the column names of rowData(inSCE). A character vector with the same length as geneSetList can be supplied if the IDs for different gene sets are found in different places, including a mixture of 'rownames' and rowData(inSCE). An integer or integer vector can be supplied to denote the column index in rowData(inSCE). Default 'rownames'.

geneSetCollection
: Class of GeneSetCollection from package GSEABase. The location of the gene IDs in inSCE should be in the description slot of each gene set and should follow the same notation as geneSetListLocation. The function getGmt can be used to read in gene sets from a GMT file. If reading a GMT file, the second column for each gene set should be the description denoting the location of the gene IDs in inSCE. These gene sets will be included with those from geneSetList if both parameters are provided.

percent_top
: An integer vector. Each element is treated as a number of top genes to compute the percentage of library size occupied by the most highly expressed genes in each cell. Default c(50, 100, 200, 500).

use_altexps
: Logical scalar indicating whether QC statistics should be computed for alternative Experiments in inSCE (altExps(inSCE)). If TRUE, statistics are computed for all alternative experiments. Alternatively, an integer or character vector specifying the alternative Experiments to use to compute QC statistics. Alternatively NULL, in which case alternative experiments are not used. Default FALSE.

flatten
: Logical scalar indicating whether the nested DataFrame-class in the output should be flattened. Default TRUE.

detectionLimit
: A numeric scalar specifying the lower detection limit for expression. Default 0

BPPARAM
: A BiocParallelParam object specifying whether the QC calculations should be parallelized. Default BiocParallel::SerialParam().

Details

This function allows multiple ways to import mitochondrial genes and quantify their expression in cells. mitoGeneLocation is required for all methods to point to the location within inSCE object that stores the mitochondrial gene IDs or Symbols. The various ways mito genes can be specified are:

- A combination of `mitoRef` and `mitoIDType` parameters can be used to load pre-built mitochondrial gene sets stored in the SCTK package. These parameters are used in the importMitoGeneSet function.
- The `mitoPrefix` parameter can be used to search for features matching a particular pattern. The default pattern is an "MT-" at the beginning of the ID.
- The `mitoID` parameter can be used to directy supply a vector of mitochondrial gene IDs or names. Only features that exactly match items in this vector will be included in the mitochondrial gene set.

Value

A SingleCellExperiment object with cell QC metrics added to the colData slot.

See Also

addPerCellQC, link{plotRunPerCellQCResults}, runCellQC

Examples

```
data(scExample, package = "singleCellTK")
mito.ix = grep("^MT-", rowData(sce)$feature_name)
geneSet <- list("Mito"=rownames(sce)[mito.ix])
sce <- runPerCellQC(sce, geneSetList = geneSet)
```

---

runSCANORAMA *Apply the mutual nearest neighbors (MNN) batch effect correction method to SingleCellExperiment object*

Description

SCANORAMA is analogous to computer vision algorithms for panorama stitching that identify images with overlapping content and merge these into a larger panorama.

Usage

```
runSCANORAMA(
  inSCE,
  useAssay = "logcounts",
  batch = "batch",
  assayName = "SCANORAMA",
  SIGMA = 15,
  ALPHA = 0.1,
  KNN = 20,
  approx = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object
useAssay	A single character indicating the name of the assay requiring batch correction. Scanorama requires a transformed normalized expression assay. Default "logcounts".
batch	A single character indicating a field in colData that annotates the batches of each cell; or a vector/factor with the same length as the number of cells. Default "batch".
assayName	A single characeter. The name for the corrected assay. Will be saved to assay. Default "SCANORAMA".
SIGMA	A numeric scalar. Algorithmic parameter, correction smoothing parameter on Gaussian kernel. Default 15.
ALPHA	A numeric scalar. Algorithmic parameter, alignment score minimum cutoff. Default 0.1.
KNN	An integer. Algorithmic parameter, number of nearest neighbors to use for matching. Default 20.
approx	Boolean. Use approximate nearest neighbors, greatly speeds up matching runtime. Default TRUE.

Value

The input SingleCellExperiment object with assay(inSCE, assayName) updated.

References

Brian Hie et al, 2019

Examples

```
## Not run:
data('sceBatches', package = 'singleCellTK')
logcounts(sceBatches) <- log1p(counts(sceBatches))
sceCorr <- runSCANORAMA(sceBatches, "ScaterLogNormCounts")

## End(Not run)
```

---

runScanpyFindClusters *runScanpyFindClusters Computes the clusters from the input sce object and stores them back in sce object*

Description

runScanpyFindClusters Computes the clusters from the input sce object and stores them back in sce object

Usage

```
runScanpyFindClusters(
  inSCE,
  useAssay = "scanpyScaledData",
  useReducedDim = "scanpyPCA",
  nNeighbors = 10,
  dims = 40,
  method = c("leiden", "louvain"),
  colDataName = NULL,
  resolution = 1,
  niterations = -1,
  flavor = "vtraag",
  use_weights = FALSE,
  cor_method = "pearson",
  inplace = TRUE,
  externalReduction = NULL,
  seed = 12345
)
```

Arguments

inSCE	(sce) object from which clusters should be computed and stored in
useAssay	Assay containing scaled counts to use for clustering.
useReducedDim	Reduction method to use for computing clusters. Default "scanpyPCA".
nNeighbors	The size of local neighborhood (in terms of number of neighboring data points) used for manifold approximation. Larger values result in more global views of the manifold, while smaller values result in more local data being preserved. Default 10.
dims	numeric value of how many components to use for computing clusters. Default 40.
method	selected method to compute clusters. One of "louvain", and "leiden". Default louvain.
colDataName	Specify the name to give to this clustering result. Default is NULL that will generate a meaningful name automatically.
resolution	A parameter value controlling the coarseness of the clustering. Higher values lead to more clusters Default 1.
niterations	How many iterations of the Leiden clustering method to perform. Positive values above 2 define the total number of iterations to perform, -1 has the method run until it reaches its optimal clustering. Default -1.
flavor	Choose between to packages for computing the clustering. Default vtraag
use_weights	Boolean. Use weights from knn graph. Default FALSE
cor_method	correlation method to use. Options are 'pearson', 'kendall', and 'spearman'. Default pearson.
inplace	If True, adds dendrogram information to annData object, else this function returns the information. Default TRUE

externalReduction
: Pass DimReduce object if PCA computed through other libraries. Default NULL.

seed
: Specify numeric value to set as a seed. Default 12345.

Value

Updated sce object which now contains the computed clusters

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")

## End(Not run)
```

---

runScanpyFindHVG *runScanpyFindHVG Find highly variable genes and store in the input sce object*

Description

runScanpyFindHVG Find highly variable genes and store in the input sce object

Usage

```
runScanpyFindHVG(
  inSCE,
  useAssay = "scanpyNormData",
  method = c("seurat", "cell_ranger", "seurat_v3"),
  altExpName = "featureSubset",
  altExp = FALSE,
  hvgNumber = 2000,
  minMean = 0.0125,
  maxMean = 3,
  minDisp = 0.5,
  maxDisp = Inf
)
```

Arguments

inSCE
: (sce) object to compute highly variable genes from and to store back to it

useAssay
: Specify the name of the assay to use for computation of variable genes. It is recommended to use log normalized data, except when flavor='seurat_v3', in which counts data is expected.

method	selected method to use for computation of highly variable genes. One of 'seurat', 'cell_ranger', or 'seurat_v3'. Default "seurat".
altExpName	Character. Name of the alternative experiment object to add if returnAsAltExp = TRUE. Default featureSubset.
altExp	Logical value indicating if the input object is an altExperiment. Default FALSE.
hvgNumber	numeric value of how many genes to select as highly variable. Default 2000
minMean	If n_top_genes unequals None, this and all other cutoffs for the means and the normalized dispersions are ignored. Ignored if flavor='seurat_v3'. Default 0.0125
maxMean	If n_top_genes unequals None, this and all other cutoffs for the means and the normalized dispersions are ignored. Ignored if flavor='seurat_v3'. Default 3
minDisp	If n_top_genes unequals None, this and all other cutoffs for the means and the normalized dispersions are ignored. Ignored if flavor='seurat_v3'. Default 0.5
maxDisp	If n_top_genes unequals None, this and all other cutoffs for the means and the normalized dispersions are ignored. Ignored if flavor='seurat_v3'. Default Inf

Value

Updated SingleCellExperiment object with highly variable genes computation stored getTopHVG, plotTopHVG

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
g <- getTopHVG(sce, method = "seurat", hvgNumber = 500)

## End(Not run)
```

---

runScanpyFindMarkers *runScanpyFindMarkers*

Description

runScanpyFindMarkers

Usage

```
runScanpyFindMarkers(
  inSCE,
  nGenes = NULL,
  useAssay = "scanpyNormData",
  colDataName,
  group1 = "all",
```

```
  group2 = "rest",
  test = c("wilcoxon", "t-test", "t-test_overestim_var", "logreg"),
  corr_method = c("benjamini-hochberg", "bonferroni")
)
```

Arguments

inSCE	Input SingleCellExperiment object.
nGenes	The number of genes that appear in the returned tables. Defaults to all genes.
useAssay	Specify the name of the assay to use for computation of marker genes. It is recommended to use log normalized assay.
colDataName	colData to use as the key of the observations grouping to consider.
group1	Name of group1. Subset of groups, to which comparison shall be restricted, or 'all' (default), for all groups.
group2	Name of group2. If 'rest', compare each group to the union of the rest of the group. If a group identifier, compare with respect to this group. Default is 'rest'
test	Test to use for DE. Default "t-test".
corr_method	p-value correction method. Used only for 't-test', 't-test_overestim_var', and 'wilcoxon'.

Value

A SingleCellExperiment object that contains marker genes populated in a data.frame stored inside metadata slot.

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyFindMarkers(sce, colDataName = "Scanpy_louvain_1" )

## End(Not run)
```

---

runScanpyNormalizeData

runScanpyNormalizeData Wrapper for NormalizeData() function from scanpy library Normalizes the sce object according to the input parameters

---

Description

runScanpyNormalizeData Wrapper for NormalizeData() function from scanpy library Normalizes the sce object according to the input parameters

Usage

```
runScanpyNormalizeData(
  inSCE,
  useAssay,
  targetSum = 10000,
  maxFraction = 0.05,
  normAssayName = "scanpyNormData"
)
```

Arguments

inSCE	(sce) object to normalize
useAssay	Assay containing raw counts to use for normalization.
targetSum	If NULL, after normalization, each observation (cell) has a total count equal to the median of total counts for observations (cells) before normalization. Default 1e4
maxFraction	Include cells that have more counts than max_fraction of the original total counts in at least one cell. Default 0.05
normAssayName	Name of new assay containing normalized data. Default scanpyNormData.

Value

Normalized SingleCellExperiment object

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
rownames(sce) <- rowData(sce)$feature_name
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")

## End(Not run)
```

---

runScanpyPCA *runScanpyPCA Computes PCA on the input sce object and stores the calculated principal components within the sce object*

Description

runScanpyPCA Computes PCA on the input sce object and stores the calculated principal components within the sce object

Usage

```
runScanpyPCA(
  inSCE,
  useAssay = "scanpyScaledData",
  reducedDimName = "scanpyPCA",
  nPCs = 50,
  method = c("arpack", "randomized", "auto", "lobpcg"),
  use_highly_variable = TRUE,
  seed = 12345
)
```

Arguments

inSCE	(sce) object on which to compute PCA
useAssay	Assay containing scaled counts to use in PCA. Default "scanpyScaledData".
reducedDimName	Name of new reducedDims object containing Scanpy PCA. Default scanpyPCA.
nPCs	numeric value of how many components to compute. Default 50.
method	selected method to use for computation of pca. One of 'arpack', 'randomized', 'auto' or 'lobpcg'. Default "arpack".
use_highly_variable	boolean value of whether to use highly variable genes only. By default uses them if they have been determined beforehand.
seed	Specify numeric value to set as a seed. Default 12345.

Value

Updated SingleCellExperiment object which now contains the computed principal components

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")

## End(Not run)
```

---

runScanpyScaleData *runScanpyScaleData Scales the input sce object according to the input parameters*

Description

runScanpyScaleData Scales the input sce object according to the input parameters

Usage

```
runScanpyScaleData(
  inSCE,
  useAssay = "scanpyNormData",
  scaledAssayName = "scanpyScaledData"
)
```

Arguments

inSCE	(sce) object to scale
useAssay	Assay containing normalized counts to scale.
scaledAssayName	Name of new assay containing scaled data. Default scanpyScaledData.

Value

Scaled SingleCellExperiment object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")

## End(Not run)
```

---

runScanpyTSNE *runScanpyTSNE Computes tSNE from the given sce object and stores the tSNE computations back into the sce object*

Description

runScanpyTSNE Computes tSNE from the given sce object and stores the tSNE computations back into the sce object

Usage

```
runScanpyTSNE(
  inSCE,
  useAssay = NULL,
  useReducedDim = "scanpyPCA",
  reducedDimName = "scanpyTSNE",
  dims = 40,
  perplexity = 30,
  externalReduction = NULL,
  seed = 12345
)
```

Arguments

inSCE	(sce) object on which to compute the tSNE
useAssay	Specify name of assay to use. Default is NULL, so useReducedDim param will be used instead.
useReducedDim	selected reduction method to use for computing tSNE. Default "scanpyPCA".
reducedDimName	Name of new reducedDims object containing Scanpy tSNE Default scanpyTSNE.
dims	Number of reduction components to use for tSNE computation. Default 40.
perplexity	Adjust the perplexity tuneable parameter for the underlying tSNE call. Default 30.
externalReduction	Pass DimReduc object if PCA computed through other libraries. Default NULL.
seed	Specify numeric value to set as a seed. Default 12345.

Value

Updated sce object with tSNE computations stored

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyTSNE(sce, useReducedDim = "scanpyPCA")

## End(Not run)
```

---

runScanpyUMAP *runScanpyUMAP Computes UMAP from the given sce object and stores the UMAP computations back into the sce object*

Description

runScanpyUMAP Computes UMAP from the given sce object and stores the UMAP computations back into the sce object

Usage

```
runScanpyUMAP(
  inSCE,
  useAssay = NULL,
  useReducedDim = "scanpyPCA",
  reducedDimName = "scanpyUMAP",
```

```
  dims = 40,
  minDist = 0.5,
  nNeighbors = 10,
  spread = 1,
  alpha = 1,
  gamma = 1,
  externalReduction = NULL,
  seed = 12345
)
```

Arguments

inSCE	(sce) object on which to compute the UMAP
useAssay	Specify name of assay to use. Default is NULL, so useReducedDim param will be used instead.
useReducedDim	Reduction to use for computing UMAP. Default is "scanpyPCA".
reducedDimName	Name of new reducedDims object containing Scanpy UMAP Default scanpyUMAP.
dims	Numerical value of how many reduction components to use for UMAP computation. Default 40.
minDist	Sets the "min_dist" parameter to the underlying UMAP call. Default 0.5.
nNeighbors	Sets the "n_neighbors" parameter to the underlying UMAP call. Default 10.
spread	Sets the "spread" parameter to the underlying UMAP call. Default 1.
alpha	Sets the "alpha" parameter to the underlying UMAP call. Default 1.
gamma	Sets the "gamma" parameter to the underlying UMAP call. Default 1.
externalReduction	Pass DimReduce object if PCA computed through other libraries. Default NULL.
seed	Specify numeric value to set as a seed. Default 12345.

Value

Updated sce object with UMAP computations stored

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runScanpyNormalizeData(sce, useAssay = "counts")
sce <- runScanpyFindHVG(sce, useAssay = "scanpyNormData", method = "seurat")
sce <- runScanpyScaleData(sce, useAssay = "scanpyNormData")
sce <- runScanpyPCA(sce, useAssay = "scanpyScaledData")
sce <- runScanpyFindClusters(sce, useReducedDim = "scanpyPCA")
sce <- runScanpyUMAP(sce, useReducedDim = "scanpyPCA")

## End(Not run)
```

`runScDblFinder` *Detect doublet cells using scDblFinder.*

Description

A wrapper function for scDblFinder. Identify potential doublet cells based on simulations of putative doublet expression profiles. Generate a doublet score for each cell.

Usage

```
runScDblFinder(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  nNeighbors = 50,
  simDoublets = max(10000, ncol(inSCE)),
  seed = 12345,
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

`inSCE`	A SingleCellExperiment object.
`sample`	Character vector or colData variable name. Indicates which sample each cell belongs to. Default `NULL`.
`useAssay`	A string specifying which assay in the SCE to use. Default `"counts"`.
`nNeighbors`	Number of nearest neighbors used to calculate density for doublet detection. Default `50`.
`simDoublets`	Number of simulated doublets created for doublet detection. Default `10000`.
`seed`	Seed for the random number generator, can be set to `NULL`. Default `12345`.
`BPPARAM`	A `BiocParallelParam-class` object specifying whether the neighbour searches should be parallelized. Default `BiocParallel::SerialParam()`.

Details

This function is a wrapper function for scDblFinder. `runScDblFinder` runs scDblFinder for each sample within `inSCE` iteratively. The resulting doublet scores for all cells will be appended to the `colData` of `inSCE`.

Value

A SingleCellExperiment object with the scDblFinder QC outputs added to the colData slot.

References

Lun ATL (2018). Detecting doublet cells with scran. https://ltla.github.io/SingleCellThoughts/software/doublet_detection/bycell.html

See Also

scDblFinder, plotScDblFinderResults, runCellQC

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runScDblFinder(sce)
```

---

runSCMerge *Apply scMerge batch effect correction method to SingleCellExperiment object*

Description

The scMerge method leverages factor analysis, stably expressed genes (SEGs) and (pseudo-) replicates to remove unwanted variations and merge multiple scRNA-Seq data.

Usage

```
runSCMerge(
  inSCE,
  useAssay = "logcounts",
  batch = "batch",
  assayName = "scMerge",
  hvgExprs = "counts",
  seg = NULL,
  kmeansK = NULL,
  cellType = NULL,
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

inSCE	Input SingleCellExperiment object
useAssay	A single character indicating the name of the assay requiring batch correction. Default "logcounts".
batch	A single character indicating a field in colData that annotates the batches. Default "batch".
assayName	A single characeter. The name for the corrected assay. Will be saved to assay. Default "scMerge".
hvgExprs	A single characeter. The assay that to be used for highly variable genes identification. Default "counts".
seg	A vector of gene names or indices that specifies SEG (Stably Expressed Genes) set as negative control. Pre-defined dataset with human and mouse SEG lists is available with segList or segList_ensemblGeneID. Default NULL, and this value will be auto-detected by default with scSEGIndex.

kmeansK	An integer vector. Indicating the kmeans' K-value for each batch (i.e. how many subclusters in each batch should exist), in order to construct pseudo-replicates. The length of codekmeansK needs to be the same as the number of batches. Default NULL, and this value will be auto-detected by default, depending on cellType.
cellType	A single character. A string indicating a field in colData(inSCE) that defines different cell types. Default 'cell_type'.
BPPARAM	A BiocParallelParam object specifying whether should be parallelized. Default BiocParallel::SerialParam().

Value

The input SingleCellExperiment object with assay(inSCE, assayName) updated.

References

Hoa, et al., 2020

Examples

```
data('sceBatches', package = 'singleCellTK')
## Not run:
logcounts(sceBatches) <- log1p(counts(sceBatches))
sceCorr <- runSCMerge(sceBatches)

## End(Not run)
```

---

runScranSNN *Get clustering with SNN graph*

Description

Perform SNN graph clustering on a SingleCellExperiment object, with graph construction by buildSNNGraph and graph clustering by "igraph" package.

Usage

```
runScranSNN(
  inSCE,
  useReducedDim = "PCA",
  useAssay = NULL,
  useAltExp = NULL,
  altExpAssay = "counts",
  altExpRedDim = NULL,
  clusterName = "cluster",
  k = 14,
  nComp = 10,
```

```
  weightType = "jaccard",
 algorithm = c("louvain", "leiden", "walktrap", "infomap", "fastGreedy", "labelProp",
   "leadingEigen"),
  BPPARAM = BiocParallel::SerialParam(),
  seed = 12345,
  ...
)
```

Arguments

inSCE	A SingleCellExperiment object.
useReducedDim	A single character, specifying which low-dimension representation (reducedDim) to perform the clustering algorithm on. Default "PCA".
useAssay	A single character, specifying which assay to perform the clustering algorithm on. Default NULL.
useAltExp	A single character, specifying the assay which altExp to perform the clustering algorithm on. Default NULL.
altExpAssay	A single character, specifying which assay in the chosen altExp to work on. Only used when useAltExp is set. Default "counts".
altExpRedDim	A single character, specifying which reducedDim within the altExp specified by useAltExp to use. Only used when useAltExp is set. Default NULL.
clusterName	A single character, specifying the name to store the cluster label in colData. Default "cluster".
k	An integer, the number of nearest neighbors used to construct the graph. Smaller value indicates higher resolution and larger number of clusters. Default 14.
nComp	An integer. The number of components to use for graph construction. Default 10. See Detail.
weightType	A single character, that specifies the edge weighing scheme when constructing the Shared Nearest-Neighbor (SNN) graph. Choose from "rank", "number", "jaccard". Default "jaccard".
algorithm	A single character, that specifies the community detection algorithm to work on the SNN graph. Choose from "leiden", "louvain", "walktrap", "infomap", "fastGreedy", "labelProp", "leadingEigen". Default "louvain". See Detail.
BPPARAM	A BiocParallelParam object to use for processing the SNN graph generation step in parallel.
seed	Random seed for reproducibility of results. Default NULL will use global seed in use by the R environment.
...	Other optional parameters passed to the igraph clustering functions. See Details.

Details

Different graph based clustering algorithms have diverse sets of parameters that users can tweak. The help information can be found here:

- for "louvain", see function help cluster_louvain
- for "leiden", see function help cluster_leiden
- for "walktrap", see function help cluster_walktrap
- for "infomap", see function help cluster_infomap
- for "fastGreedy", see function help cluster_fast_greedy
- for "labelProp", see function help cluster_label_prop
- for "leadingEigen", see function help cluster_leading_eigen

The Scran SNN building method can work on specified nComp components. When users specify input matrix by useAssay or useAltExp + altExpAssay, the method will generate nComp components and use them all. When specifying useReducedDim or useAltExp + altExpRedDim, this function will subset the top nComp components and pass them to the method.

Value

The input SingleCellExperiment object with factor cluster labeling updated in colData(inSCE)[[clusterName]].

References

Aaron Lun and et. al., 2016

Examples

```
data("mouseBrainSubsetSCE")
mouseBrainSubsetSCE <- runScranSNN(mouseBrainSubsetSCE,
                                   useReducedDim = "PCA_logcounts")
```

---

runScrublet *Find doublets using* scrublet.

Description

A wrapper function that calls scrub_doublets from python module scrublet. Simulates doublets from the observed data and uses a k-nearest-neighbor classifier to calculate a continuous scrublet_score (between 0 and 1) for each transcriptome. The score is automatically thresholded to generate scrublet_call, a boolean array that is TRUE for predicted doublets and FALSE otherwise.

Usage

```
runScrublet(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  simDoubletRatio = 2,
  nNeighbors = NULL,
```

```
  minDist = NULL,
  expectedDoubletRate = 0.1,
  stdevDoubletRate = 0.02,
  syntheticDoubletUmiSubsampling = 1,
  useApproxNeighbors = TRUE,
  distanceMetric = "euclidean",
  getDoubletNeighborParents = FALSE,
  minCounts = 3,
  minCells = 3L,
  minGeneVariabilityPctl = 85,
  logTransform = FALSE,
  meanCenter = TRUE,
  normalizeVariance = TRUE,
  nPrinComps = 30L,
  tsneAngle = NULL,
  tsnePerplexity = NULL,
  verbose = TRUE,
  seed = 12345
)
```

Arguments

inSCE	A SingleCellExperiment object.
sample	Character vector or colData variable name. Indicates which sample each cell belongs to. Default NULL.
useAssay	A string specifying which assay in the SCE to use. Default "counts".
simDoubletRatio	Numeric. Number of doublets to simulate relative to the number of observed transcriptomes. Default 2.0.
nNeighbors	Integer. Number of neighbors used to construct the KNN graph of observed transcriptomes and simulated doublets. If NULL, this is set to round(0.5 * sqrt(n_cells)). Default NULL.
minDist	Float Determines how tightly UMAP packs points together. If NULL, this is set to 0.1. Default NULL.
expectedDoubletRate	The estimated doublet rate for the experiment. Default 0.1.
stdevDoubletRate	Uncertainty in the expected doublet rate. Default 0.02.
syntheticDoubletUmiSubsampling	Numeric. Rate for sampling UMIs when creating synthetic doublets. If 1.0, each doublet is created by simply adding the UMIs from two randomly sampled observed transcriptomes. For values less than 1, the UMI counts are added and then randomly sampled at the specified rate. Defuault 1.0.
useApproxNeighbors	Boolean. Use approximate nearest neighbor method (annoy) for the KNN classifier. Default TRUE.

distanceMetric Character. Distance metric used when finding nearest neighbors. See detail. Default "euclidean".

getDoubletNeighborParents
Boolean. If TRUE, return the parent transcriptomes that generated the doublet neighbors of each observed transcriptome. This information can be used to infer the cell states that generated a given doublet state. Default FALSE.

minCounts Numeric. Used for gene filtering prior to PCA. Genes expressed at fewer than minCounts in fewer than minCells are excluded. Default 3.

minCells Integer. Used for gene filtering prior to PCA. Genes expressed at fewer than minCounts in fewer than minCells are excluded. Default 3.

minGeneVariabilityPctl
Numeric. Used for gene filtering prior to PCA. Keep the most highly variable genes (in the top minGeneVariabilityPctl percentile), as measured by the v-statistic (Klein et al., Cell 2015). Default 85.

logTransform Boolean. If TRUE, log-transform the counts matrix (log1p(TPM)). sklearn.decomposition.TruncatedS will be used for dimensionality reduction, unless meanCenter is TRUE. Default FALSE.

meanCenter If TRUE, center the data such that each gene has a mean of 0. sklearn.decomposition.PCA will be used for dimensionality reduction. Default TRUE.

normalizeVariance
Boolean. If TRUE, normalize the data such that each gene has a variance of 1. sklearn.decomposition.TruncatedSVD will be used for dimensionality reduction, unless meanCenter is TRUE. Default TRUE.

nPrinComps Integer. Number of principal components used to embed the transcriptomes prior to k-nearest-neighbor graph construction. Default 30.

tsneAngle Float. Determines angular size of a distant node as measured from a point in the t-SNE plot. If NULL, it is set to 0.5. Default NULL.

tsnePerplexity Integer. The number of nearest neighbors that is used in other manifold learning algorithms. If NULL, it is set to 30. Default NULL.

verbose Boolean. If TRUE, print progress updates. Default TRUE.

seed Seed for the random number generator, can be set to NULL. Default 12345.

Details

For the list of valid values for distanceMetric, see the documentation for annoy (if useApproxNeighbors is TRUE) or sklearn.neighbors.NearestNeighbors (if useApproxNeighbors is FALSE).

Value

A SingleCellExperiment object with scrub_doublets output appended to the colData slot. The columns include scrublet_score and scrublet_call.

See Also

plotScrubletResults, runCellQC

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- runScrublet(sce)

## End(Not run)
```

---

runSeuratFindClusters *runSeuratFindClusters Computes the clusters from the input sce object and stores them back in sce object*

Description

runSeuratFindClusters Computes the clusters from the input sce object and stores them back in sce object

Usage

```
runSeuratFindClusters(
  inSCE,
  useAssay = "seuratNormData",
  useReduction = c("pca", "ica"),
  dims = 10,
  algorithm = c("louvain", "multilevel", "SLM"),
  groupSingletons = TRUE,
  resolution = 0.8,
  seed = 12345,
  externalReduction = NULL,
  verbose = TRUE
)
```

Arguments

inSCE	(sce) object from which clusters should be computed and stored in
useAssay	Assay containing scaled counts to use for clustering.
useReduction	Reduction method to use for computing clusters. One of "pca" or "ica". Default "pca".
dims	numeric value of how many components to use for computing clusters. Default 10.
algorithm	selected algorithm to compute clusters. One of "louvain", "multilevel", or "SLM". Use louvain for "original Louvain algorithm" and multilevel for "Louvain algorithm with multilevel refinement". Default louvain.
groupSingletons	boolean if singletons should be grouped together or not. Default TRUE.

resolution	Set the resolution parameter to find larger (value above 1) or smaller (value below 1) number of communities. Default 0.8.
seed	Specify the seed value. Default 12345.
externalReduction	Pass DimReduc object if PCA/ICA computed through other libraries. Default NULL.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Value

Updated sce object which now contains the computed clusters

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")
sce <- runSeuratFindClusters(sce, useAssay = "counts")

## End(Not run)
```

---

runSeuratFindHVG *runSeuratFindHVG Find highly variable genes and store in the input sce object*

Description

runSeuratFindHVG Find highly variable genes and store in the input sce object

Usage

```
runSeuratFindHVG(
  inSCE,
  useAssay = "counts",
  method = c("vst", "dispersion", "mean.var.plot"),
  hvgNumber = 2000,
  createFeatureSubset = "hvf",
  altExp = FALSE,
  verbose = TRUE
)
```

Arguments

inSCE	(sce) object to compute highly variable genes from and to store back to it
useAssay	Specify the name of the assay to use for computation of variable genes. It is recommended to use a raw counts assay with the "vst" method and normalized assay with all other methods. Default is "counts".
method	selected method to use for computation of highly variable genes. One of 'vst', 'dispersion', or 'mean.var.plot'. Default "vst" which uses the raw counts. All other methods use normalized counts.
hvgNumber	numeric value of how many genes to select as highly variable. Default 2000
createFeatureSubset	Specify a name of the subset to create for the identified variable features. Default is "hvf". Leave it NULL if you do not want to create a subset of variable features.
altExp	Logical value indicating if the input object is an altExperiment. Default FALSE.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Value

Updated SingleCellExperiment object with highly variable genes computation stored

See Also

runFeatureSelection, runModelGeneVar, getTopHVG, plotTopHVG

Examples

```
data(scExample, package = "singleCellTK")
sce <- runSeuratFindHVG(sce)
```

---

runSeuratFindMarkers *runSeuratFindMarkers*

Description

runSeuratFindMarkers

Usage

```
runSeuratFindMarkers(
  inSCE,
  cells1 = NULL,
  cells2 = NULL,
  group1 = NULL,
  group2 = NULL,
  allGroup = NULL,
  conserved = FALSE,
```

```
  test = "wilcox",
  onlyPos = FALSE,
  minPCT = 0.1,
  threshUse = 0.25,
  verbose = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
cells1	A list of sample names included in group1.
cells2	A list of sample names included in group2.
group1	Name of group1.
group2	Name of group2.
allGroup	Name of all groups.
conserved	Logical value indicating if markers conserved between two groups should be identified. Default is FALSE.
test	Test to use for DE. Default "wilcox".
onlyPos	Logical value indicating if only positive markers should be returned.
minPCT	Numeric value indicating the minimum fraction of min.pct cells in which genes are detected. Default is 0.1.
threshUse	Numeric value indicating the logFC threshold value on which on average, at least X-fold difference (log-scale) between the two groups of cells exists. Default is 0.25.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Value

A SingleCellExperiment object that contains marker genes populated in a data.frame stored inside metadata slot.

---

runSeuratHeatmap *runSeuratHeatmap Computes the heatmap plot object from the pca slot in the input sce object*

Description

runSeuratHeatmap Computes the heatmap plot object from the pca slot in the input sce object

Usage

```
runSeuratHeatmap(
  inSCE,
  useAssay,
  useReduction = c("pca", "ica"),
  dims = NULL,
  nfeatures = 30,
  cells = NULL,
  ncol = NULL,
  balanced = TRUE,
  fast = TRUE,
  combine = TRUE,
  raster = TRUE,
  externalReduction = NULL
)
```

Arguments

inSCE	(sce) object from which to compute heatmap (pca should be computed)
useAssay	Specify name of the assay that will be scaled by this function. The output scaled assay will be used for computation of the heatmap.
useReduction	Reduction method to use for computing clusters. One of "pca" or "ica". Default "pca".
dims	Number of components to generate heatmap plot objects. If NULL, a heatmap will be generated for all components. Default NULL.
nfeatures	Number of features to include in the heatmap. Default 30.
cells	Numeric value indicating the number of top cells to plot. Default is NULL which indicates all cells.
ncol	Numeric value indicating the number of columns to use for plot. Default is NULL which will automatically compute accordingly.
balanced	Plot equal number of genes with positive and negative scores. Default is TRUE.
fast	See DimHeatmap for more information. Default TRUE.
combine	See DimHeatmap for more information. Default TRUE.
raster	See DimHeatmap for more information. Default TRUE.
externalReduction	Pass DimReduc object if PCA/ICA computed through other libraries. Default NULL.

Value

plot object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")
heatmap <- runSeuratHeatmap(sce, useAssay = "counts")
plotSeuratHeatmap(heatmap)

## End(Not run)
```

`runSeuratICA` *runSeuratICA Computes ICA on the input sce object and stores the calculated independent components within the sce object*

Description

runSeuratICA Computes ICA on the input sce object and stores the calculated independent components within the sce object

Usage

```
runSeuratICA(
  inSCE,
  useAssay = "seuratScaledData",
  useFeatureSubset = NULL,
  scale = TRUE,
  reducedDimName = "seuratICA",
  nics = 20,
  seed = 12345,
  verbose = FALSE
)
```

Arguments

`inSCE`	(sce) object on which to compute ICA
`useAssay`	Assay containing scaled counts to use in ICA.
`useFeatureSubset`	Subset of feature to use for dimension reduction. A character string indicating a `rowData` variable that stores the logical vector of HVG selection, or a vector that can subset the rows of `inSCE`. Default `NULL`.
`scale`	Logical scalar, whether to standardize the expression values using `ScaleData`. Default `TRUE`.
`reducedDimName`	Name of new reducedDims object containing Seurat ICA Default `seuratICA`.
`nics`	Number of independent components to compute. Default `20`.

seed	Random seed for reproducibility of results. Default NULL will use global seed in use by the R environment.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Details

For features used for computation, it can be controlled by features or useFeatureSubset. When features is specified, the scaling and dimensionality reduction will only be processed with these features. When features is NULL but useFeatureSubset is specified, will use the features that the HVG list points to. If both parameters are NULL, the function will see if any Seurat's variable feature detection has been ever performed, and use them if found. Otherwise, all features are used.

Value

Updated SingleCellExperiment object which now contains the computed independent components

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratICA(sce, useAssay = "counts")

## End(Not run)
```

---

runSeuratIntegration *runSeuratIntegration A wrapper function to Seurat Batch-Correction/Integration workflow.*

Description

runSeuratIntegration A wrapper function to Seurat Batch-Correction/Integration workflow.

Usage

```
runSeuratIntegration(
  inSCE,
  useAssay = "counts",
  batch,
  newAssayName = "SeuratIntegratedAssay",
  kAnchor,
  kFilter,
  kWeight,
  ndims = 10
)
```

Arguments

inSCE	Input SingleCellExperiment object that contains the assay to batch-correct.
useAssay	Assay to batch-correct.
batch	Batch variable from colData slot of SingleCellExperiment object.
newAssayName	Assay name for the batch-corrected output assay.
kAnchor	Number of neighbours to use for finding the anchors in the FindIntegrationAnchors function.
kFilter	Number of neighbours to use for filtering the anchors in the FindIntegrationAnchors function.
kWeight	Number of neighbours to use when weigthing the anchors in the IntegrateData function.
ndims	Number of dimensions to use. Default 10.

Value

A SingleCellExperiment object that contains the batch-corrected assay inside the altExp slot of the object

---

runSeuratJackStraw *runSeuratJackStraw Compute jackstraw plot and store the computations in the input sce object*

Description

runSeuratJackStraw Compute jackstraw plot and store the computations in the input sce object

Usage

```
runSeuratJackStraw(
  inSCE,
  useAssay,
  dims = NULL,
  numReplicate = 100,
  propFreq = 0.025,
  externalReduction = NULL
)
```

Arguments

inSCE	(sce) object on which to compute and store jackstraw plot
useAssay	Specify name of the assay to use for scaling. Assay name provided against this parameter is scaled by the function and used for the computation of JackStraw scores along with the reduced dimensions specified by the dims parameter.

dims — Number of components to test in Jackstraw. If NULL, then all components are used. Default NULL.

numReplicate — Numeric value indicating the number of replicate samplings to perform. Default value is 100.

propFreq — Numeric value indicating the proportion of data to randomly permute for each replicate. Default value is 0.025.

externalReduction — Pass DimReduc object if PCA/ICA computed through other libraries. Default NULL.

Value

Updated SingleCellExperiment object with jackstraw computations stored in it

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")
sce <- runSeuratJackStraw(sce, useAssay = "counts")

## End(Not run)
```

---

runSeuratNormalizeData

runSeuratNormalizeData Wrapper for NormalizeData() function from seurat library Normalizes the sce object according to the input parameters

---

Description

runSeuratNormalizeData Wrapper for NormalizeData() function from seurat library Normalizes the sce object according to the input parameters

Usage

```
runSeuratNormalizeData(
  inSCE,
  useAssay,
  normAssayName = "seuratNormData",
  normalizationMethod = "LogNormalize",
  scaleFactor = 10000,
  verbose = TRUE
)
```

Arguments

inSCE	(sce) object to normalize
useAssay	Assay containing raw counts to use for normalization.
normAssayName	Name of new assay containing normalized data. Default seuratNormData.
normalizationMethod	selected normalization method. Default "LogNormalize".
scaleFactor	numeric value that represents the scaling factor. Default 10000.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Value

Normalized SingleCellExperiment object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")

## End(Not run)
```

---

runSeuratPCA *runSeuratPCA Computes PCA on the input sce object and stores the calculated principal components within the sce object*

Description

runSeuratPCA Computes PCA on the input sce object and stores the calculated principal components within the sce object

Usage

```
runSeuratPCA(
  inSCE,
  useAssay = "seuratNormData",
  useFeatureSubset = "hvf",
  scale = TRUE,
  reducedDimName = "seuratPCA",
  nPCs = 20,
  seed = 12345,
  verbose = TRUE
)
```

Arguments

inSCE	(sce) object on which to compute PCA
useAssay	Assay containing scaled counts to use in PCA. Default "seuratNormData".
useFeatureSubset	Subset of feature to use for dimension reduction. A character string indicating a rowData variable that stores the logical vector of HVG selection, or a vector that can subset the rows of inSCE. Default "hvf".
scale	Logical scalar, whether to standardize the expression values using ScaleData. Default TRUE.
reducedDimName	Name of new reducedDims object containing Seurat PCA. Default seuratPCA.
nPCs	numeric value of how many components to compute. Default 20.
seed	Random seed for reproducibility of results. Default NULL will use global seed in use by the R environment.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Details

For features used for computation, it can be controlled by features or useFeatureSubset. When features is specified, the scaling and dimensionality reduction will only be processed with these features. When features is NULL but useFeatureSubset is specified, will use the features that the HVG list points to. If both parameters are NULL, the function will see if any Seurat's variable feature detection has been ever performed, and use them if found. Otherwise, all features are used.

Value

Updated SingleCellExperiment object which now contains the computed principal components

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- setTopHVG(sce, method = "vst", featureSubsetName = "hvf")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")

## End(Not run)
```

runSeuratScaleData *runSeuratScaleData Scales the input sce object according to the input parameters*

Description

runSeuratScaleData Scales the input sce object according to the input parameters

Usage

```
runSeuratScaleData(
  inSCE,
  useAssay = "seuratNormData",
  scaledAssayName = "seuratScaledData",
  model = "linear",
  scale = TRUE,
  center = TRUE,
  scaleMax = 10,
  verbose = TRUE
)
```

Arguments

inSCE	(sce) object to scale
useAssay	Assay containing normalized counts to scale.
scaledAssayName	Name of new assay containing scaled data. Default seuratScaledData.
model	selected model to use for scaling data. Default "linear".
scale	boolean if data should be scaled or not. Default TRUE.
center	boolean if data should be centered or not. Default TRUE
scaleMax	maximum numeric value to return for scaled data. Default 10.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Value

Scaled SingleCellExperiment object

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")

## End(Not run)
```

runSeuratSCTransform *runSeuratSCTransform Runs the SCTransform function to transform/normalize the input data*

Description

runSeuratSCTransform Runs the SCTransform function to transform/normalize the input data

Usage

```
runSeuratSCTransform(
  inSCE,
  normAssayName = "SCTCounts",
  useAssay = "counts",
  verbose = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object
normAssayName	Name for the output data assay. Default "SCTCounts".
useAssay	Name for the input data assay. Default "counts".
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Value

Updated SingleCellExperiment object containing the transformed data

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runSeuratSCTransform(mouseBrainSubsetSCE)
```

runSeuratTSNE *runSeuratTSNE Computes tSNE from the given sce object and stores the tSNE computations back into the sce object*

Description

runSeuratTSNE Computes tSNE from the given sce object and stores the tSNE computations back into the sce object

Usage

```
runSeuratTSNE(
  inSCE,
  useReduction = c("pca", "ica"),
  reducedDimName = "seuratTSNE",
  dims = 10,
  perplexity = 30,
  externalReduction = NULL,
  seed = 1
)
```

Arguments

inSCE	(sce) object on which to compute the tSNE
useReduction	selected reduction algorithm to use for computing tSNE. One of "pca" or "ica". Default "pca".
reducedDimName	Name of new reducedDims object containing Seurat tSNE Default seuratTSNE.
dims	Number of reduction components to use for tSNE computation. Default 10.
perplexity	Adjust the perplexity tuneable parameter for the underlying tSNE call. Default 30.
externalReduction	Pass DimReduc object if PCA/ICA computed through other libraries. Default NULL.
seed	Random seed for reproducibility of results. Default 1.

Value

Updated sce object with tSNE computations stored

---

runSeuratUMAP *runSeuratUMAP Computes UMAP from the given sce object and stores the UMAP computations back into the sce object*

Description

runSeuratUMAP Computes UMAP from the given sce object and stores the UMAP computations back into the sce object

Usage

```
runSeuratUMAP(
  inSCE,
  useReduction = c("pca", "ica"),
  reducedDimName = "seuratUMAP",
  dims = 10,
```

```
  minDist = 0.3,
  nNeighbors = 30L,
  spread = 1,
  externalReduction = NULL,
  seed = 42,
  verbose = TRUE
)
```

Arguments

inSCE	(sce) object on which to compute the UMAP
useReduction	Reduction to use for computing UMAP. One of "pca" or "ica". Default is "pca".
reducedDimName	Name of new reducedDims object containing Seurat UMAP Default seuratUMAP.
dims	Numerical value of how many reduction components to use for UMAP computation. Default 10.
minDist	Sets the "min.dist" parameter to the underlying UMAP call. See RunUMAP for more information. Default 0.3.
nNeighbors	Sets the "n.neighbors" parameter to the underlying UMAP call. See RunUMAP for more information. Default 30L.
spread	Sets the "spread" parameter to the underlying UMAP call. See RunUMAP for more information. Default 1.
externalReduction	Pass DimReduc object if PCA/ICA computed through other libraries. Default NULL.
seed	Random seed for reproducibility of results. Default 42.
verbose	Logical value indicating if informative messages should be displayed. Default is TRUE.

Value

Updated sce object with UMAP computations stored

Examples

```
data(scExample, package = "singleCellTK")
## Not run:
sce <- runSeuratNormalizeData(sce, useAssay = "counts")
sce <- runSeuratFindHVG(sce, useAssay = "counts")
sce <- runSeuratScaleData(sce, useAssay = "counts")
sce <- runSeuratPCA(sce, useAssay = "counts")
sce <- runSeuratFindClusters(sce, useAssay = "counts")
sce <- runSeuratUMAP(sce, useReduction = "pca")

## End(Not run)
```

runSingleR *Label cell types with SingleR*

Description

SingleR works with a reference dataset where the cell type labeling is given. Given a reference dataset of samples (single-cell or bulk) with known labels, it assigns those labels to new cells from a test dataset based on similarities in their expression profiles.

Usage

```
runSingleR(
  inSCE,
  useAssay = "logcounts",
  useSCERef = NULL,
  labelColName = NULL,
  useBltinRef = c("hpca", "bpe", "mp", "dice", "immgen", "mouse", "zeisel"),
  level = "fine",
  featureType = c("symbol", "ensembl"),
  labelByCluster = NULL
)
```

Arguments

inSCE	SingleCellExperiment inherited object. Required.
useAssay	character. A string specifying which assay to use for expression profile identification. Required.
useSCERef	SingleCellExperiment inherited object. An optional customized reference dataset. Default NULL.
labelColName	A single character. A string specifying the column in colData(useSCERef) that stores the cell type labeling. Default NULL.
useBltinRef	A single character. A string that specifies a reference provided by SingleR. Choose from "hpca", "bpe", "mp", "dice", "immgen","mouse", "zeisel". See detail. Default "hpca".
level	A string for cell type labeling level. Used only when using some of the SingleR built-in references. Choose from "main", "fine","ont". Default "fine".
featureType	A string for whether to use gene symbols or Ensembl IDs when using a SingleR built-in reference. Should be set based on the type of rownames of inSCE. Choose from "symbol", "ensembl". Default "symbol".
labelByCluster	A single character. A string specifying the column name in colData(inSCE) that stores clustering labels. Use this when users want to only label cells on cluster level, instead of performing calculation on each cell. Default NULL.

Value

Input SCE object with cell type labeling updated in colData(inSCE), together with scoring metrics.

Examples

```
data("sceBatches")
logcounts(sceBatches) <- log1p(counts(sceBatches))
#sceBatches <- runSingleR(sceBatches, useBltinRef = "mp")
```

---

runSoupX *Detecting and correct contamination with SoupX*

Description

A wrapper function for autoEstCont and adjustCounts. Identify potential contamination from experimental factors such as ambient RNA. Visit their vignette for better understanding.

Usage

```
runSoupX(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  background = NULL,
  bgAssayName = NULL,
  bgBatch = NULL,
  assayName = ifelse(is.null(background), "SoupX", "SoupX_bg"),
  cluster = NULL,
  reducedDimName = ifelse(is.null(background), "SoupX_UMAP_", "SoupX_bg_UMAP_"),
  tfidfMin = 1,
  soupQuantile = 0.9,
  maxMarkers = 100,
  contaminationRange = c(0.01, 0.8),
  rhoMaxFDR = 0.2,
  priorRho = 0.05,
  priorRhoStdDev = 0.1,
  forceAccept = FALSE,
  adjustMethod = c("subtraction", "soupOnly", "multinomial"),
  roundToInt = FALSE,
  tol = 0.001,
  pCut = 0.01
)
```

Arguments

inSCE	A SingleCellExperiment object.
sample	A single character specifying a name that can be found in `colData(inSCE)` to directly use the cell annotation; or a character vector with as many elements as cells to indicates which sample each cell belongs to. SoupX will be run on cells from each sample separately. Default `NULL`.

useAssay	A single character string specifying which assay in inSCE to use. Default 'counts'.
background	A numeric matrix of counts or a SingleCellExperiment object with the matrix in assay slot. It should have the same structure as inSCE except it contains the matrix including empty droplets. Default NULL.
bgAssayName	A single character string specifying which assay in background to use when background is a SingleCellExperiment object. If NULL, the function will use the same value as useAssay. Default NULL.
bgBatch	The same thing as sample but for background. Can be a single character only when background is a SingleCellExperiment object. Default NULL.
assayName	A single character string of the output corrected matrix. Default "SoupX" when not using a background, otherwise, "SoupX_bg".
cluster	Prior knowledge of clustering labels on cells. A single character string for specifying clustering label stored in colData(inSCE), or a character vector with as many elements as cells. When not supplied, quickCluster method will be applied.
reducedDimName	A single character string of the prefix of output corrected embedding matrix for each sample. Default "SoupX_UMAP_" when not using a background, otherwise, "SoupX_bg_UMAP_".
tfidfMin	Numeric. Minimum value of tfidf to accept for a marker gene. Default 1. See ?SoupX::autoEstCont.
soupQuantile	Numeric. Only use genes that are at or above this expression quantile in the soup. This prevents inaccurate estimates due to using genes with poorly constrained contribution to the background. Default 0.9. See ?SoupX::autoEstCont.
maxMarkers	Integer. If we have heaps of good markers, keep only the best maxMarkers of them. Default 100. See ?SoupX::autoEstCont.
contaminationRange	Numeric vector of two elements. This constrains the contamination fraction to lie within this range. Must be between 0 and 1. The high end of this range is passed to estimateNonExpressingCells as maximumContamination. Default c(0.01, 0.8). See ?SoupX::autoEstCont.
rhoMaxFDR	Numeric. False discovery rate passed to estimateNonExpressingCells, to test if rho is less than maximumContamination. Default 0.2. See ?SoupX::autoEstCont.
priorRho	Numeric. Mode of gamma distribution prior on contamination fraction. Default 0.05. See ?SoupX::autoEstCont.
priorRhoStdDev	Numeric. Standard deviation of gamma distribution prior on contamination fraction. Default 0.1. See ?SoupX::autoEstCont.
forceAccept	Logical. Should we allow very high contamination fractions to be used. Passed to setContaminationFraction. Default FALSE. See ?SoupX::autoEstCont.
adjustMethod	Character. Method to use for correction. One of 'subtraction', 'soupOnly', or 'multinomial'. Default 'subtraction'. See ?SoupX::adjustCounts.
roundToInt	Logical. Should the resulting matrix be rounded to integers? Default FALSE. See ?SoupX::adjustCounts.
tol	Numeric. Allowed deviation from expected number of soup counts. Don't change this. Default 0.001. See ?SoupX::adjustCounts.

pCut — Numeric. The p-value cut-off used when method = 'soupOnly'. Default 0.01. See ?SoupX::adjustCounts.

Value

The input inSCE object with soupX_nUMIs, soupX_clustrers, soupX_contamination appended to colData slot; soupX_{sample}_est and soupX_{sample}_counts for each sample appended to rowData slot; and other computational metrics at getSoupX(inSCE). Replace "soupX" to "soupX_bg" when background is used.

Author(s)

Yichen Wang

See Also

plotSoupXResults

Examples

```
## Not run:
# SoupX does not work for toy example,
sce <- importExampleData("pbmc3k")
sce <- runSoupX(sce, sample = "sample")
plotSoupXResults(sce, sample = "sample")

## End(Not run)
```

---

runTSCAN *Run TSCAN to obtain pseudotime values for cells*

Description

Wrapper for obtaining a pseudotime ordering of the cells by projecting them onto the minimum spanning tree (MST)

Usage

```
runTSCAN(
  inSCE,
  useReducedDim = "PCA",
  cluster = NULL,
  starter = NULL,
  seed = 12345
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useReducedDim	Character. A low-dimension representation in reducedDims, will be used for both clustering if cluster not specified and MST construction. Default "PCA".
cluster	Grouping for each cell in inSCE. A vector with equal length to the number of the cells in inSCE, or a single character for retriving colData variable. Default NULL, will run runScranSNN to obtain.
starter	Character. Specifies the starting node from which to compute the pseudotime. Default NULL, will select an arbitrary node.
seed	An integer. Random seed for clustering if cluster is not specified. Default 12345.

Value

The input inSCE object with pseudotime ordering of the cells along the paths and the cluster label stored in colData, and other unstructured information in metadata.

Author(s)

Nida Pervaiz

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
```

---

runTSCANClusterDEAnalysis

Find DE genes between all TSCAN paths rooted from given cluster

---

Description

This function finds all paths that root from a given cluster useCluster, and performs tests to identify significant features for each path, and are not significant and/or changing in the opposite direction in the other paths. Using a branching cluster (i.e. a node with degree > 2) may highlight features which are responsible for the branching event. MST has to be pre-calculated with runTSCAN.

Usage

```
runTSCANClusterDEAnalysis(
  inSCE,
  useCluster,
  useAssay = "logcounts",
  fdrThreshold = 0.05
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useCluster	The cluster to be regarded as the root, has to existing in colData(inSCE)$TSCAN_clusters.
useAssay	Character. The name of the assay to use. This assay should contain log normalized counts. Default "logcounts".
fdrThreshold	Only out put DEGs with FDR value smaller than this value. Default 0.05.

Value

The input inSCE with results updated in metadata.

Author(s)

Nida Pervaiz

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
mouseBrainSubsetSCE <- runTSCANClusterDEAnalysis(inSCE = mouseBrainSubsetSCE,
                                         useCluster = 1)
```

---

runTSCANDEG *Test gene expression changes along a TSCAN trajectory path*

Description

Wrapper for identifying genes with significant changes with respect to one of the TSCAN pseudotime paths

Usage

```
runTSCANDEG(inSCE, pathIndex, useAssay = "logcounts", discardCluster = NULL)
```

Arguments

inSCE	Input SingleCellExperiment object.
pathIndex	Path index for which the pseudotime values should be used. This corresponds to the terminal node of specific path from the root node to the terminal node. Run listTSCANTerminalNodes(inSCE) for available options.
useAssay	Character. The name of the assay to use for testing the expression change. Should be log-normalized. Default "logcounts"
discardCluster	Cluster(s) which are not of use or masks other interesting effects can be discarded. Default NULL.

Value

The input `inSCE` with results updated in `metadata`.

Author(s)

Nida Pervaiz

Examples

```
data("mouseBrainSubsetSCE", package = "singleCellTK")
mouseBrainSubsetSCE <- runTSCAN(inSCE = mouseBrainSubsetSCE,
                                useReducedDim = "PCA_logcounts")
terminalNodes <- listTSCANTerminalNodes(mouseBrainSubsetSCE)
mouseBrainSubsetSCE <- runTSCANDEG(inSCE = mouseBrainSubsetSCE,
                                   pathIndex = terminalNodes[1])
```

---

runTSNE *Run t-SNE embedding with Rtsne method*

Description

T-Stochastic Neighbour Embedding (t-SNE) algorithm is commonly for 2D visualization of single-cell data. This function wraps the Rtsne `Rtsne` function.

With this funciton, users can create tSNE embedding directly from raw count matrix, with necessary preprocessing including normalization, scaling, dimension reduction all automated. Yet we still recommend having the PCA as input, so that the result can match with the clustering based on the same input PCA, and will be much faster.

Usage

```
runTSNE(
  inSCE,
  useReducedDim = "PCA",
  useAssay = NULL,
  useAltExp = NULL,
  reducedDimName = "TSNE",
  logNorm = TRUE,
  useFeatureSubset = NULL,
  nTop = 2000,
  center = TRUE,
  scale = TRUE,
  pca = TRUE,
  partialPCA = FALSE,
  initialDims = 25,
  theta = 0.5,
  perplexity = 30,
  nIterations = 1000,
```

```
  numThreads = 1,
  seed = 12345
)

runQuickTSNE(inSCE, useAssay = "counts", ...)

getTSNE(
  inSCE,
  useReducedDim = "PCA",
  useAssay = NULL,
  useAltExp = NULL,
  reducedDimName = "TSNE",
  logNorm = TRUE,
  useFeatureSubset = NULL,
  nTop = 2000,
  center = TRUE,
  scale = TRUE,
  pca = TRUE,
  partialPCA = FALSE,
  initialDims = 25,
  theta = 0.5,
  perplexity = 30,
  nIterations = 1000,
  numThreads = 1,
  seed = 12345
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useReducedDim	The low dimension representation to use for UMAP computation. Default "PCA".
useAssay	Assay to use for tSNE computation. If useAltExp is specified, useAssay has to exist in assays(altExp(inSCE, useAltExp)). Default NULL.
useAltExp	The subset to use for tSNE computation, usually for the selected.variable features. Default NULL.
reducedDimName	a name to store the results of the dimension reductions. Default "TSNE".
logNorm	Whether the counts will need to be log-normalized prior to generating the tSNE via scaterlogNormCounts. Ignored when using useReducedDim. Default TRUE.
useFeatureSubset	Subset of feature to use for dimension reduction. A character string indicating a rowData variable that stores the logical vector of HVG selection, or a vector that can subset the rows of inSCE. Default NULL.
nTop	Automatically detect this number of variable features to use for dimension reduction. Ignored when using useReducedDim or using useFeatureSubset. Default 2000.
center	Whether data should be centered before PCA is applied. Ignored when using useReducedDim. Default TRUE.

scale	Whether data should be scaled before PCA is applied. Ignored when using useReducedDim. Default TRUE.
pca	Whether an initial PCA step should be performed. Ignored when using useReducedDim. Default TRUE.
partialPCA	Whether truncated PCA should be used to calculate principal components (requires the irlba package). This is faster for large input matrices. Ignored when using useReducedDim. Default FALSE.
initialDims	Number of dimensions from PCA to use as input in tSNE. Default 25.
theta	Numeric value for speed/accuracy trade-off (increase for less accuracy), set to 0.0 for exact TSNE. Default 0.5.
perplexity	perplexity parameter. Should not be bigger than 3 * perplexity < ncol(inSCE) - 1. Default 30. See Rtsne details for interpretation.
nIterations	maximum iterations. Default 1000.
numThreads	Integer, number of threads to use using OpenMP, Default 1. 0 corresponds to using all available cores.
seed	Random seed for reproducibility of tSNE results. Default NULL will use global seed in use by the R environment.
...	Other parameters to be passed to runTSNE

Value

A SingleCellExperiment object with tSNE computation updated in reducedDim(inSCE, reducedDimName).

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
# Run from raw counts
sce <- runQuickTSNE(sce)
## Not run:
# Run from PCA
sce <- scaterlogNormCounts(sce, "logcounts")
sce <- runModelGeneVar(sce)
sce <- setTopHVG(sce, method = "modelGeneVar", hvgNumber = 2000,
                 featureSubsetName = "HVG_modelGeneVar2000")
sce <- scaterPCA(sce, useAssay = "logcounts",
                 useFeatureSubset = "HVG_modelGeneVar2000", scale = TRUE)
sce <- runTSNE(sce, useReducedDim = "PCA")

## End(Not run)
```

runUMAP *Run UMAP embedding with scater method*

Description

Uniform Manifold Approximation and Projection (UMAP) algorithm is commonly for 2D visualization of single-cell data. These functions wrap the scater `calculateUMAP` function.

Users can use `runQuickUMAP` to directly create UMAP embedding from raw count matrix, with necessary preprocessing including normalization, variable feature selection, scaling, dimension reduction all automated. Therefore, `useReducedDim` is disabled for `runQuickUMAP`.

In a complete analysis, we still recommend having dimension reduction such as PCA created beforehand and select proper numbers of dimensions for using `runUMAP`, so that the result can match with the clustering based on the same input PCA.

Usage

```
runUMAP(
  inSCE,
  useReducedDim = "PCA",
  useAssay = NULL,
  useAltExp = NULL,
  sample = NULL,
  reducedDimName = "UMAP",
  logNorm = TRUE,
  useFeatureSubset = NULL,
  nTop = 2000,
  scale = TRUE,
  pca = TRUE,
  initialDims = 10,
  nNeighbors = 30,
  nIterations = 200,
  alpha = 1,
  minDist = 0.01,
  spread = 1,
  seed = 12345,
  verbose = TRUE,
  BPPARAM = SerialParam()
)

runQuickUMAP(inSCE, useAssay = "counts", sample = "sample", ...)

getUMAP(
  inSCE,
  useReducedDim = "PCA",
  useAssay = NULL,
  useAltExp = NULL,
```

```
  sample = NULL,
  reducedDimName = "UMAP",
  logNorm = TRUE,
  useFeatureSubset = NULL,
  nTop = 2000,
  scale = TRUE,
  pca = TRUE,
  initialDims = 25,
  nNeighbors = 30,
  nIterations = 200,
  alpha = 1,
  minDist = 0.01,
  spread = 1,
  seed = 12345,
  BPPARAM = SerialParam()
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useReducedDim	The low dimension representation to use for UMAP computation. If useAltExp is specified, useReducedDim has to exist in reducedDims(altExp(inSCE, useAltExp)). Default "PCA".
useAssay	Assay to use for UMAP computation. If useAltExp is specified, useAssay has to exist in assays(altExp(inSCE, useAltExp)). Ignored when using useReducedDim. Default NULL.
useAltExp	The subset to use for UMAP computation, usually for the selected variable features. Default NULL.
sample	Character vector. Indicates which sample each cell belongs to. If given a single character, will take the annotation from colData. Default NULL.
reducedDimName	A name to store the results of the UMAP embedding coordinates obtained from this method. Default "UMAP".
logNorm	Whether the counts will need to be log-normalized prior to generating the UMAP via scaterlogNormCounts. Ignored when using useReducedDim. Default TRUE.
useFeatureSubset	Subset of feature to use for dimension reduction. A character string indicating a rowData variable that stores the logical vector of HVG selection, or a vector that can subset the rows of inSCE. Default NULL.
nTop	Automatically detect this number of variable features to use for dimension reduction. Ignored when using useReducedDim or using useFeatureSubset. Default 2000.
scale	Whether useAssay matrix will need to be standardized. Default TRUE.
pca	Logical. Whether to perform dimension reduction with PCA before UMAP. Ignored when using useReducedDim. Default TRUE.
initialDims	Number of dimensions from PCA to use as input in UMAP. Default 10.

nNeighbors	The size of local neighborhood used for manifold approximation. Larger values result in more global views of the manifold, while smaller values result in more local data being preserved. Default 30. See calculateUMAP for more information.
nIterations	The number of iterations performed during layout optimization. Default is 200.
alpha	The initial value of "learning rate" of layout optimization. Default is 1.
minDist	The effective minimum distance between embedded points. Smaller values will result in a more clustered/clumped embedding where nearby points on the manifold are drawn closer together, while larger values will result on a more even dispersal of points. Default 0.01. See calculateUMAP for more information.
spread	The effective scale of embedded points. In combination with minDist, this determines how clustered/clumped the embedded points are. Default 1. See calculateUMAP for more information.
seed	Random seed for reproducibility of UMAP results. Default NULL will use global seed in use by the R environment.
verbose	Logical. Whether to print log messages. Default TRUE.
BPPARAM	A BiocParallelParam object specifying whether the PCA should be parallelized.
...	Parameters passed to runUMAP

Value

A SingleCellExperiment object with UMAP computation updated in reducedDim(inSCE, reducedDimName).

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
# Run from raw counts
sce <- runQuickUMAP(sce)
plotDimRed(sce, "UMAP")
```

---

runVAM *Run VAM to score gene sets in single cell data*

Description

Wrapper for the Variance-adjusted Mahalanobis (VAM), which is a fast and accurate method for cell-specific gene set scoring of single cell data. This algorithm computes distance statistics and one-sided p-values for all cells in the specified single cell gene expression matrix. Gene sets should already be imported and stored in the meta data using functions such as importGeneSetsFromList or importGeneSetsFromMSigDB

Usage

```
runVAM(
  inSCE,
  geneSetCollectionName = "H",
  useAssay = "logcounts",
  resultNamePrefix = NULL,
  center = FALSE,
  gamma = TRUE
)
```

Arguments

inSCE	Input SingleCellExperiment object.
geneSetCollectionName	Character. The name of the gene set collection to use. Default "H".
useAssay	Character. The name of the assay to use. This assay should contain log normalized counts. Default "logcounts".
resultNamePrefix	Character. Prefix to the name the VAM results which will be stored in the reducedDim slot of inSCE. The names of the output matrices will be resultNamePrefix_Distance and resultNamePrefix_CDF. If this parameter is set to NULL, then "VAM_geneSetCollectionName_" will be used. Default NULL.
center	Boolean. If TRUE, values will be mean centered when computing the Mahalanobis statistic. Default FALSE.
gamma	Boolean. If TRUE, a gamma distribution will be fit to the non-zero squared Mahalanobis distances computed from a row-permuted version of the gene expression matrix. The estimated gamma distribution will be used to compute a one-sided p-value for each cell. If FALSE, the p-value will be computed using the standard chi-square approximation for the squared Mahalanobis distance (or non-central if center = FALSE). Default TRUE.

Value

A SingleCellExperiment object with VAM metrics stored in reducedDim as VAM_NameOfTheGeneset_Distance and VAM_NameOfTheGeneset_CDF.

Author(s)

Nida Pervaiz

See Also

importGeneSetsFromList, importGeneSetsFromMSigDB, importGeneSetsFromGMT, importGeneSetsFromCollection for importing gene sets. sctkListGeneSetCollections, getPathwayResultNames and getGenesetNamesFromCollection for available related information in inSCE.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- scaterlogNormCounts(sce, assayName = "logcounts")
gs1 <- rownames(sce)[seq(10)]
gs2 <- rownames(sce)[seq(11,20)]
gs <- list("geneset1" = gs1, "geneset2" = gs2)
sce <- importGeneSetsFromList(inSCE = sce,geneSetList = gs,
                              by = "rownames")
sce <- runVAM(inSCE = sce,
              geneSetCollectionName = "GeneSetCollection",
              useAssay = "logcounts")
```

`runZINBWaVE` *Apply ZINBWaVE Batch effect correction method to SingleCellExperiment object*

Description

A general and flexible zero-inflated negative binomial model that can be used to provide a low-dimensional representations of scRNAseq data. The model accounts for zero inflation (dropouts), over-dispersion, and the count nature of the data. The model also accounts for the difference in library sizes and optionally for batch effects and/or other covariates.

Usage

```
runZINBWaVE(
  inSCE,
  useAssay = "counts",
  batch = "batch",
  nHVG = 1000L,
  nComponents = 50L,
  epsilon = 1000,
  nIter = 10L,
  reducedDimName = "zinbwave",
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

`inSCE`	Input SingleCellExperiment object
`useAssay`	A single character indicating the name of the assay requiring batch correction. Note that ZINBWaVE works for counts (integer) input rather than logcounts that other methods prefer. Default `"counts"`.
`batch`	A single character indicating a field in `colData` that annotates the batches. Default `"batch"`.

nHVG	An integer. Number of highly variable genes to use when fitting the model. Default 1000L.
nComponents	An integer. The number of principle components or dimensionality to generate in the resulting matrix. Default 50L.
epsilon	An integer. Algorithmic parameter. Empirically, a high epsilon is often required to obtained a good low-level representation. Default 1000L.
nIter	An integer, The max number of iterations to perform. Default 10L.
reducedDimName	A single character. The name for the corrected low-dimensional representation. Will be saved to reducedDim(inSCE). Default "zinbwave".
BPPARAM	A BiocParallelParam object specifying whether should be parallelized. Default BiocParallel::SerialParam().

Value

The input SingleCellExperiment object with reducedDim(inSCE, reducedDimName) updated.

References

Pollen, Alex A et al., 2014

Examples

```
data('sceBatches', package = 'singleCellTK')
## Not run:
    sceCorr <- runZINBWaVE(sceBatches, nIter = 5)

## End(Not run)
```

---

sampleSummaryStats *Generate table of SCTK QC outputs.*

Description

Creates a table of QC metrics generated from QC algorithms, which is stored within the metadata slot of the input SingleCellExperiment object.

Usage

```
sampleSummaryStats(
  inSCE,
  sample = NULL,
  useAssay = "counts",
  simple = TRUE,
  statsName = "qc_table"
)
```

Arguments

inSCE	Input SingleCellExperiment object with saved assay data and/or colData data. Required.
sample	Character vector. Indicates which sample each cell belongs to.
useAssay	A string specifying which assay in the SCE to use. Default 'counts'.
simple	Boolean. Indicates whether to generate a table of only basic QC stats (ex. library size), or to generate a summary table of all QC stats stored in the inSCE.
statsName	Character. The name of the slot that will store the QC stat table. Default "qc_table".

Value

A SingleCellExperiment object with a summary table for QC statistics in the 'sample_summary' slot of metadata.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- sampleSummaryStats(sce, simple = TRUE)
getSampleSummaryStatsTable(sce, statsName = "qc_table")
```

---

scaterCPM *scaterCPM Uses CPM from scater library to compute counts-per-million.*

Description

scaterCPM Uses CPM from scater library to compute counts-per-million.

Usage

```
scaterCPM(inSCE, assayName = "ScaterCPMCounts", useAssay = "counts")
```

Arguments

inSCE	Input SingleCellExperiment object
assayName	New assay name for cpm data.
useAssay	Input assay

Value

inSCE Updated SingleCellExperiment object

Author(s)

Irzam Sarfraz

Examples

```
data(sce_chcl, package = "scds")
sce_chcl <- scaterCPM(sce_chcl,"countsCPM", "counts")
```

`scaterlogNormCounts` *scaterlogNormCounts Uses logNormCounts to log normalize input data*

Description

scaterlogNormCounts Uses logNormCounts to log normalize input data

Usage

```
scaterlogNormCounts(
  inSCE,
  assayName = "ScaterLogNormCounts",
  useAssay = "counts"
)
```

Arguments

`inSCE`	Input SingleCellExperiment object
`assayName`	New assay name for log normalized data
`useAssay`	Input assay

Value

inSCE Updated SingleCellExperiment object that contains the new log normalized data

Author(s)

Irzam Sarfraz

Examples

```
data(sce_chcl, package = "scds")
sce_chcl <- scaterlogNormCounts(sce_chcl,"logcounts", "counts")
```

scaterPCA *Perform scater PCA on a SingleCellExperiment Object*

Description

A wrapper to runPCA function to compute principal component analysis (PCA) from a given SingleCellExperiment object.

Usage

```
scaterPCA(
  inSCE,
  useAssay = "logcounts",
  useFeatureSubset = "hvg2000",
  scale = TRUE,
  reducedDimName = "PCA",
  nComponents = 50,
  ntop = 2000,
  useAltExp = NULL,
  seed = 12345,
  BPPARAM = BiocParallel::SerialParam()
)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Assay to use for PCA computation. If useAltExp is specified, useAssay has to exist in assays(altExp(inSCE, useAltExp)). Default "logcounts"
useFeatureSubset	Subset of feature to use for dimension reduction. A character string indicating a rowData variable that stores the logical vector of HVG selection, or a vector that can subset the rows of inSCE. Default "hvg2000".
scale	Logical scalar, whether to standardize the expression values. Default TRUE.
reducedDimName	Name to use for the reduced output assay. Default "PCA".
nComponents	Number of principal components to obtain from the PCA computation. Default 50.
ntop	Automatically detect this number of variable features to use for dimension reduction. Ignored when using useReducedDim or using useFeatureSubset. Default 2000.
useAltExp	The subset to use for PCA computation, usually for the selected.variable features. Default NULL.
seed	Integer, random seed for reproducibility of PCA results. Default NULL.
BPPARAM	A BiocParallelParam object specifying whether the PCA should be parallelized.

Value

A SingleCellExperiment object with PCA computation updated in `reducedDim(inSCE, reducedDimName)`.

Examples

```
data(scExample, package = "singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- scaterlogNormCounts(sce, "logcounts")

# Example of ranking variable genes, selecting the top variable features,
# and running PCA. Make sure to increase the number of highly variable
# features (hvgNumber) and the number of principal components (nComponents)
# for real datasets
sce <- runModelGeneVar(sce, useAssay = "logcounts")
sce <- setTopHVG(sce, method = "modelGeneVar", hvgNumber = 100,
                 featureSubsetName = "hvf")
sce <- scaterPCA(sce, useAssay = "logcounts", scale = TRUE,
                 useFeatureSubset = "hvf", nComponents = 5)

# Alternatively, let the scater PCA function select the top variable genes
sce <- scaterPCA(sce, useAssay = "logcounts", scale = TRUE,
                 useFeatureSubset = NULL, ntop = 100, nComponents = 5)
```

---

sce *Example Single Cell RNA-Seq data in SingleCellExperiment Object, subset of 10x public dataset*

Description

https://support.10xgenomics.com/single-cell-gene-expression/datasets/2.1.0/pbmc4k A subset of 390 barcodes and top 200 genes were included in this example. Within 390 barcodes, 195 barcodes are empty droplet, 150 barcodes are cell barcode and 45 barcodes are doublets predicted by scrublet and doubletFinder package. This example only serves as a proof of concept and a tutoriol on how to run the functions in this package. The results should not be used for drawing scientific conclusions.

Usage

```
data("scExample")
```

Format

A SingleCellExperiment object.

Value

Example Single Cell RNA-Seq data in SingleCellExperiment Object, subset of 10x public dataset

Examples

```
data("scExample")
```

sceBatches *Example Single Cell RNA-Seq data in SingleCellExperiment object, with different batches annotated*

Description

Two batches of pancreas scRNAseq dataset are combined with their original counts. Cell types and batches are annotated in 'colData(sceBatches)'. Two batches came from Wang, et al., 2016, annotated as ''w''; and Xin, et al., 2016, annotated as ''x''. Two common cell types, ''alpha'' and ''beta'', that could be found in both original studies with relatively large population were kept for cleaner demonstration.

Usage

```
data('sceBatches')
```

Format

An object of class `SingleCellExperiment` with 100 rows and 250 columns.

Value

Example Single Cell RNA-Seq data in SingleCellExperiment object, with different batches annotated

sctkListGeneSetCollections *Lists imported GeneSetCollections*

Description

Returns a vector of GeneSetCollections that have been imported and stored in `metadata(inSCE)$sctk$genesets`.

Usage

```
sctkListGeneSetCollections(inSCE)
```

Arguments

`inSCE`	A SingleCellExperiment object.

Value

Character vector.

Author(s)

Joshua D. Campbell

See Also

importGeneSetsFromList for importing from lists, importGeneSetsFromGMT for importing from GMT files, GeneSetCollection objects, and importGeneSetsFromMSigDB for importing MSigDB gene sets.

Examples

```
data(scExample)
gs1 <- GSEABase::GeneSet(setName = "geneset1",
                         geneIds = rownames(sce)[seq(10)])
gs2 <- GSEABase::GeneSet(setName = "geneset2",
                         geneIds = rownames(sce)[seq(11,20)])
gsc1 <- GSEABase::GeneSetCollection(gs1)
gsc2 <- GSEABase::GeneSetCollection(gs2)
sce <- importGeneSetsFromCollection(inSCE = sce,
                                    geneSetCollection = gsc1,
                                    by = "rownames",
                                    collectionName = "Collection1")
sce <- importGeneSetsFromCollection(inSCE = sce,
                                    geneSetCollection = gsc2,
                                    by = "rownames",
                                    collectionName = "Collection2")
collections <- sctkListGeneSetCollections(sce)
```

---

sctkPythonInstallConda

Installs Python packages into a Conda environment

Description

Install all Python packages used in the singleCellTK package using conda_install from package reticulate. This will create a new Conda environment with the name envname if not already present. Note that Anaconda or Miniconda already need to be installed on the local system.

Usage

```
sctkPythonInstallConda(
  envname = "sctk-reticulate",
  conda = "auto",
  packages = c("scipy", "numpy", "astroid", "six"),
 pipPackages = c("scrublet", "scanpy", "louvain", "leidenalg", "bbknn", "scanorama",
    "anndata"),
  selectConda = TRUE,
  forge = FALSE,
```

```
  pipIgnoreInstalled = TRUE,
  pythonVersion = NULL,
  ...
)
```

Arguments

envname	Character. Name of the conda environment to create.
conda	Character. Path to conda executable. Usue "auto" to find conda using the PATH and other conventional install locations. Default 'auto'.
packages	Character Vector. List of packages to install from Conda.
pipPackages	Character Vector. List of packages to install into the Conda environment using 'pip'.
selectConda	Boolean. Run selectSCTKConda after installing all packages to select the Conda environment. Default TRUE.
forge	Boolean. Include the Conda Forge repository.
pipIgnoreInstalled	Boolean. Ignore installed versions when using pip. This is TRUE by default so that specific package versions can be installed even if they are downgrades. The FALSE option is useful for situations where you don't want a pip install to attempt an overwrite of a conda binary package (e.g. SciPy on Windows which is very difficult to install via pip due to compilation requirements).
pythonVersion	Passed to python_version variable in conda_install. Default NULL.
...	Other parameters to pass to conda_install.

Value

None. Installation of Conda environment.

See Also

See conda_create for more information on creating a Conda environment. See conda_install for more description of the installation parameters. See https://rstudio.github.io/reticulate/ for more information on package reticulate. See selectSCTKConda for reloading the Conda environment if R is restarted without going through the whole installation process again. See https://docs.conda.io/en/latest/ for more information on Conda environments.

Examples

```
## Not run:
sctkPythonInstallConda(envname = "sctk-reticulate")

## End(Not run)
```

sctkPythonInstallVirtualEnv *Installs Python packages into a virtual environment*

Description

Install all Python packages used in the singleCellTK package using virtualenv_install from package reticulate. This will create a new virtual environment with the name envname if not already present.

Usage

```
sctkPythonInstallVirtualEnv(
  envname = "sctk-reticulate",
 packages = c("scipy", "numpy", "astroid", "six", "scrublet", "scanpy", "louvain",
    "leidenalg", "scanorama", "bbknn", "anndata"),
  selectEnvironment = TRUE,
  python = NULL
)
```

Arguments

envname	Character. Name of the virtual environment to create.
packages	Character Vector. List of packages to install.
selectEnvironment	Boolean. Run selectSCTKVirtualEnvironment after installing all packages to select the virtual environment. Default TRUE.
python	The path to a Python interpreter, to be used with the created virtual environment. When NULL, the Python interpreter associated with the current session will be used. Default NULL.

Value

None. Installation of virtual environment.

See Also

See virtualenv_create for more information on creating a Conda environment. See virtualenv_install for more description of the installation parameters. See https://rstudio.github.io/reticulate/ for more information on package reticulate. See selectSCTKVirtualEnvironment for reloading the virtual environment if R is restarted without going through the whole installation process again.

Examples

```
## Not run:
sctkPythonInstallVirtualEnv(envname = "sctk-reticulate")

## End(Not run)
```

SEG *Stably Expressed Gene (SEG) list obect, with SEG sets for human and mouse.*

Description

The two gene sets came from dataset called 'segList' of package 'scMerge'.

Usage

```
data('SEG')
```

Format

list, with two entries "human" and "mouse", each is a charactor vector.

Value

Stably Expressed Gene (SEG) list obect, with SEG sets for human and mouse.

Source

```
data('segList', package='scMerge')
```

Examples

```
data('SEG')
humanSEG <- SEG$human
```

---

selectSCTKConda *Selects a Conda environment*

Description

Selects a Conda environment with Python packages used in singleCellTK.

Usage

```
selectSCTKConda(envname = "sctk-reticulate")
```

Arguments

envname	Character. Name of the conda environment to activate.

Value

None. Selects Conda environment.

See Also

conda-tools for more information on using Conda environments with package reticulate. See https://rstudio.github.io/reticulate/ for more information on package reticulate.

See sctkPythonInstallConda for installation of Python modules into a Conda environment. Seeconda-tools for more information on using Conda environments with package reticulate. See https://rstudio.github.io/reticulate/ for more information on package reticulate. See https://docs.conda.io/en/latest/ for more information on Conda environments.

Examples

```
## Not run:
sctkPythonInstallConda(envname = "sctk-reticulate", selectConda = FALSE)
selectSCTKConda(envname = "sctk-reticulate")

## End(Not run)
```

---

selectSCTKVirtualEnvironment

Selects a virtual environment

Description

Selects a virtual environment with Python packages used in singleCellTK

Usage

```
selectSCTKVirtualEnvironment(envname = "sctk-reticulate")
```

Arguments

envname	Character. Name of the virtual environment to activate.

Value

None. Selects virtual environment.

See Also

See sctkPythonInstallVirtualEnv for installation of Python modules into a virtual environment. Seevirtualenv-tools for more information on using virtual environments with package reticulate. See https://rstudio.github.io/reticulate/ for more information on package reticulate.

Examples

```
## Not run:
sctkPythonInstallVirtualEnv(envname = "sctk-reticulate", selectEnvironment = FALSE)
selectSCTKVirtualEnvironment(envname = "sctk-reticulate")

## End(Not run)
```

---

setRowNames *Set rownames of SCE with a character vector or a rowData column*

Description

Users can set rownames of an SCE object with either a character vector where the length equals to `nrow(x)`, or a single character specifying a column in `rowData(x)`. Also applicable to matrix like object where `rownames<-` method works, but only allows full size name vector. Users can set `dedup = TRUE` to remove duplicated entries in the specification, by adding `-1`, `-2`, `...`, `-i` suffix to the duplication of the same identifier.

Usage

```
setRowNames(x, rowNames, dedup = TRUE)
```

Arguments

`x`	Input object where the rownames will be modified.
`rowNames`	Character vector of the rownames. If `x` is an SingleCellExperiment object, a single character specifying a column in `rowData(x)`.
`dedup`	Logical. Whether to deduplicate the specified rowNames. Default `TRUE`

Value

The input SCE object with rownames updated.

Examples

```
data("scExample", package = "singleCellTK")
head(rownames(sce))
sce <- setRowNames(sce, "feature_name")
head(rownames(sce))
```

setSCTKDisplayRow *Indicates which rowData to use for visualization*

Description

This function is to be used to specify which

Usage

```
setSCTKDisplayRow(inSCE, featureDisplayRow)
```

Arguments

inSCE	Input SingleCellExperiment object with saved dimension reduction components or a variable with saved results. Required.
featureDisplayRow	Indicates which column name of rowData to be used for plots.

Value

A SingleCellExperiment object with the specific column name of rowData to be used for plotting stored in metadata.

Examples

```
data(scExample, package="singleCellTK")
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
sce <- setSCTKDisplayRow(inSCE = sce, featureDisplayRow = "feature_name")
plotSCEViolinAssayData(inSCE = sce, feature = "ENSG00000019582")
```

singleCellTK *Run the single cell analysis app*

Description

Use this function to run the single cell analysis app.

Usage

```
singleCellTK(inSCE = NULL, includeVersion = TRUE, theme = "yeti")
```

Arguments

inSCE	Input SingleCellExperiment object.
includeVersion	Include the version number in the SCTK header. The default is TRUE.
theme	The bootswatch theme to use for the singleCellTK UI. The default is 'flatly'.

Value

The shiny app will open

Examples

```
## Not run:
#Upload data through the app
singleCellTK()

# Load the app with a SingleCellExperiment object
data("mouseBrainSubsetSCE")
singleCellTK(mouseBrainSubsetSCE)

## End(Not run)
```

---

subDiffEx *Passes the output of generateSimulatedData() to differential expression tests, picking either t-tests or ANOVA for data with only two conditions or multiple conditions, respectively.*

Description

Passes the output of generateSimulatedData() to differential expression tests, picking either t-tests or ANOVA for data with only two conditions or multiple conditions, respectively.

Usage

```
subDiffEx(tempData)

subDiffExttest(countMatrix, class.labels, test.type = "t.equalvar")

subDiffExANOVA(countMatrix, condition)
```

Arguments

tempData	Matrix. The output of generateSimulatedData(), where the first row contains condition labels.
countMatrix	Matrix. A simulated counts matrix, sans labels.
class.labels	Factor. The condition labels for the simulated cells. Will be coerced into 1's and 0's.
test.type	Type of test to perform. The default is t.equalvar.
condition	Factor. The condition labels for the simulated cells.

Value

subDiffEx(): A vector of fdr-adjusted p-values for all genes. Nonviable results (such as for genes with 0 counts in a simulated dataset) are coerced to 1.

subDiffExttest(): A vector of fdr-adjusted p-values for all genes. Nonviable results (such as for genes with 0 counts in a simulated dataset) are coerced to 1.

subDiffExANOVA(): A vector of fdr-adjusted p-values for all genes. Nonviable results (such as for genes with 0 counts in a simulated dataset) are coerced to 1.

Functions

- `subDiffEx()`:
- `subDiffExttest()`: Runs t-tests on all genes in a simulated dataset with 2 conditions, and adjusts for FDR.
- `subDiffExANOVA()`: Runs ANOVA on all genes in a simulated dataset with more than 2 conditions, and adjusts for FDR.

Examples

```
data("mouseBrainSubsetSCE")
res <- generateSimulatedData(
         totalReads = 1000, cells=10,
         originalData = assay(mouseBrainSubsetSCE, "counts"),
         realLabels = colData(mouseBrainSubsetSCE)[, "level1class"])
tempSigDiff <- subDiffEx(res)

data("mouseBrainSubsetSCE")
#sort first 100 expressed genes
ord <- rownames(mouseBrainSubsetSCE)[
  order(rowSums(assay(mouseBrainSubsetSCE, "counts")),
        decreasing = TRUE)][seq(100)]
#subset to those first 100 genes
subset <- mouseBrainSubsetSCE[ord, ]
res <- generateSimulatedData(totalReads = 1000, cells=10,
                             originalData = assay(subset, "counts"),
                             realLabels = colData(subset)[, "level1class"])
realLabels <- res[1, ]
output <- res[-1, ]
fdr <- subDiffExttest(output, realLabels)

data("mouseBrainSubsetSCE")
#sort first 100 expressed genes
ord <- rownames(mouseBrainSubsetSCE)[
  order(rowSums(assay(mouseBrainSubsetSCE, "counts")),
        decreasing = TRUE)][seq(100)]
# subset to those first 100 genes
subset <- mouseBrainSubsetSCE[ord, ]
res <- generateSimulatedData(totalReads = 1000, cells=10,
                             originalData = assay(subset, "counts"),
                             realLabels = colData(subset)[, "level2class"])
realLabels <- res[1, ]
```

```
output <- res[-1, ]
fdr <- subDiffExANOVA(output, realLabels)
```

`subsetSCECols` *Subset a SingleCellExperiment object by columns*

Description

Used to peform subsetting of a SingleCellExperiment object using a variety of methods that indicate the correct columns to keep. The various methods, `index`, `bool`, and `colData`, can be used in conjunction with one another.

Usage

```
subsetSCECols(inSCE, index = NULL, bool = NULL, colData = NULL)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`index`	Integer vector. Vector of indicies indicating which columns to keep. If `NULL`, this will not be used for subsetting. Default `NULL`.
`bool`	Boolean vector. Vector of `TRUE` or `FALSE` indicating which columns should be kept. Needs to be the same length as the number of columns in `inSCE`. If `NULL`, this will not be used for subsetting. Default `NULL`.
`colData`	Character. An expression that will identify a subset of columns using variables found in the `colData` of `inSCE`. For example, if `x` is a numeric vector in `colData`, then `"x < 5"` will return all columns with x less than 5. Single quotes should be used for character strings. For example, `"y == 'yes'"` will return all columns where y is "yes". Multiple expressions can be evaluated by placing them in a vector. For example `c("x < 5", "y =='yes'")` will apply both operations for subsetting. If `NULL`, this will not be used for subsetting. Default `NULL`.

Value

A SingleCellExperiment object that has been subsetted by colData.

Author(s)

Joshua D. Campbell

Examples

```
data(scExample)
sce <- subsetSCECols(sce, colData = "type != 'EmptyDroplet'")
```

subsetSCERows *Subset a SingleCellExperiment object by rows*

Description

Used to peform subsetting of a SingleCellExperiment object using a variety of methods that indicate the correct rows to keep. The various methods, `index`, `bool`, and `rowData`, can be used in conjunction with one another. If `returnAsAltExp` is set to `TRUE`, then the returned object will have the same number of rows as the input `inSCE` as the subsetted object will be stored in the `altExp` slot.

Usage

```
subsetSCERows(
  inSCE,
  index = NULL,
  bool = NULL,
  rowData = NULL,
  returnAsAltExp = TRUE,
  altExpName = "subset",
  prependAltExpName = TRUE
)
```

Arguments

`inSCE`	Input SingleCellExperiment object.
`index`	Integer vector. Vector of indicies indicating which rows to keep. If `NULL`, this will not be used for subsetting. Default `NULL`.
`bool`	Boolean vector. Vector of `TRUE` or `FALSE` indicating which rows should be kept. Needs to be the same length as the number of rows in `inSCE`. If `NULL`, this will not be used for subsetting. Default `NULL`.
`rowData`	Character. An expression that will identify a subset of rows using variables found in the `rowData` of `inSCE`. For example, if `x` is a numeric vector in `rowData`, then `"x < 5"` will return all rows with x less than 5. Single quotes should be used for character strings. For example, `"y == 'yes'"` will return all rows where y is "yes". Multiple expressions can be evaluated by placing them in a vector. For example `c("x < 5", "y =='yes'")` will apply both operations for subsetting. If `NULL`, this will not be used for subsetting. Default `NULL`.
`returnAsAltExp`	Boolean. If `TRUE`, the subsetted SingleCellExperiment object will be returned in the `altExp` slot of `inSCE`. If `FALSE`, the subsetted SingleCellExperiment object will be directly returned.
`altExpName`	Character. Name of the alternative experiment object to add if `returnAsAltExp = TRUE`. Default `subset`.
`prependAltExpName`	Boolean. If `TRUE`, `altExpName` will be added to the beginning of the assay names in the `altExp` object. This is only utilized if `returnAsAltExp = TRUE`. Default `TRUE`.

Value

A SingleCellExperiment object that has been subsetted by rowData.

Author(s)

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Examples

```
data(scExample)

# Set a variable up in the rowData indicating mitochondrial genes
rowData(sce)$isMito <- ifelse(grepl("^MT-", rowData(sce)$feature_name),
                              "yes", "no")
sce <- subsetSCERows(sce, rowData = "isMito == 'yes'")
```

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summarizeSCE *Summarize an assay in a SingleCellExperiment*

Description

Creates a table of summary metrics from an input SingleCellExperiment

Usage

```
summarizeSCE(inSCE, useAssay = NULL, sampleVariableName = NULL)
```

Arguments

inSCE	Input SingleCellExperiment object.
useAssay	Indicate which assay to summarize. If NULL, then the first assay in inSCE will be used. Default NULL.
sampleVariableName	Variable name in colData denoting which sample each cell belongs to. If NULL, all cells will be assumed to come from the same sample. Default "sample".

Value

A data.frame object of summary metrics.

Examples

```
data("mouseBrainSubsetSCE")
summarizeSCE(mouseBrainSubsetSCE, sample = NULL)
```

`trimCounts` *Trim Counts*

Description

Trims an input count matrix such that each value greater than a threshold value and each value less than a provided lower threshold value is trimmed to the lower treshold value.

Usage

```
trimCounts(counts, trimValue = c(10, -10))
```

Arguments

`counts`	matrix
`trimValue`	where trimValue[1] for upper threshold and trimValue[2] as lower threshold. Default is `c(10,-10)`

Value

trimmed counts matrix

Examples

```
data(sce_chcl, package = "scds")
assay(sce_chcl, "countsTrimmed") <- trimCounts(assay(sce_chcl, "counts"),
                                               c(10, -10))
```

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