

# Package ‘scDDboost’

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**Type** Package

**Title** A compositional model to assess expression changes from single-cell rna-seq data

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## Description

scDDboost is an R package to analyze changes in the distribution of single-cell expression data between two experimental conditions. Compared to other methods that assess differential expression, scDDboost benefits uniquely from information conveyed by the clustering of cells into cellular subtypes. Through a novel empirical Bayesian formulation it calculates gene-specific posterior probabilities that the marginal expression distribution is the same (or different) between the two conditions. The implementation in scDDboost treats gene-level expression data within each condition as a mixture of negative binomial distributions.

**License** GPL (>= 2)

**Imports** Rcpp (>= 0.12.11), RcppEigen (>= 0.3.2.9.0), EBSeq, BiocParallel, mclust, SingleCellExperiment, cluster, Oscope, SummarizedExperiment, stats, methods

**biocViews** SingleCell, Software, Clustering, Sequencing, GeneExpression, DifferentialExpression, Bayesian

**Depends** R (>= 4.2), ggplot2

**LinkingTo** Rcpp, RcppEigen, BH

**Suggests** knitr, rmarkdown, BiocStyle, testthat

**SystemRequirements** c++11

**Roxygen** list(wrap=FALSE)

**RoxygenNote** 7.1.2

**VignetteBuilder** knitr

**BugReports** <https://github.com/wiscstatman/scDDboost/issues>

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## R topics documented:

scDDboost-package . . . . .	2
calD . . . . .	3
clusHelper . . . . .	4
detK . . . . .	5
EBS . . . . .	5
extractInfo . . . . .	6
gCl . . . . .	7
genRClus . . . . .	7
getDD . . . . .	8
getsizeofDD . . . . .	8
getZ1Z2 . . . . .	9
gRef . . . . .	9
isRef . . . . .	10
LL . . . . .	10
lpt1t2 . . . . .	11
lpzgt . . . . .	11
mdd . . . . .	12
pat . . . . .	12
pdd . . . . .	13
pddAggregate . . . . .	14
pddRandom . . . . .	15
rwMle . . . . .	15
sim_dat . . . . .	16

**Index**

**17**

## Description

scDDboost is an R package to analyze changes in the distribution of single-cell expression data between two experimental conditions. Compared to other methods that assess differential expression, scDDboost benefits uniquely from information conveyed by the clustering of cells into cellular subtypes. Through a novel empirical Bayesian formulation it calculates gene-specific posterior probabilities that the marginal expression distribution is the same (or different) between the two conditions. The implementation in scDDboost treats gene-level expression data within each condition as a mixture of negative binomial distributions.

## Details

The DESCRIPTION file: This package was not yet installed at build time.

Index: This package was not yet installed at build time.

Package used to score evidence of differential distribution in single-cell RNA-seq data

## Author(s)

NA

Maintainer: NA

## References

<https://projecteuclid.org/journals/annals-of-applied-statistics/volume-15/issue-2/A-compositional-model-to-assess-expression-changes-from-single-cell/10.1214/20-AOAS1423.short>

## See Also

<https://github.com/wiscstatman/scDDboost/blob/master/DESCRIPTION>

## Examples

```
data(sim_dat)
dat = extractInfo(sim_dat)
data_counts = dat$count_matrix
cd = dat$condition
bp <- BiocParallel::MulticoreParam(4)
D_c = calD(data_counts, bp)
pDD = pdd(data_counts, cd, bp, D_c)
```

---

calD

*calculate distance matrix*

---

## Description

calculate distance matrix

**Usage**

```
calD(data, bp)
```

**Arguments**

data	transcripts
bp	bioc parallel parameter

**Value**

distance matrix

**Examples**

```
data(sim_dat)
dat <- extractInfo(sim_dat)
data_counts <- dat$count_matrix
bp <- BiocParallel::MulticoreParam(4)
D_c <- calD(data_counts, bp)
```

<i>clusHelper</i>	<i>function to get intra and inter distance for clusters</i>
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**Description**

function to get intra and inter distance for clusters

**Usage**

```
clusHelper(D, i)
```

**Arguments**

D	distance matrix
i	number of clusters

**Value**

vector of intra and inter distance

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detK	<i>determine the number of clusters</i>
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---

## Description

determine the number of clusters

## Usage

```
detK(D, epi = 1)
```

## Arguments

D	distance matrix
epi	threshold for cutting off

## Value

number of clusters

## Examples

```
data(sim_dat)
dat <- extractInfo(sim_dat)
data_counts <- dat$count_matrix
bp <- BiocParallel::MulticoreParam(4)
D_c <- calD(data_counts, bp)
detK(D_c)
```

---

EBS	<i>accelerated empirical bayesian</i>
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---

## Description

accelerated empirical bayesian

## Usage

```
EBS(data, conditions, gclus, sf, iter = 10, hyper, PP, stp1, stp2)
```

**Arguments**

<code>data</code>	single cell expression matrix, row as genes column as cells
<code>conditions</code>	partition of cells
<code>gclus</code>	partition of genes
<code>sf</code>	size factors
<code>iter</code>	maximum iteration step of EM
<code>hyper</code>	hyper parameters for beta distributions
<code>PP</code>	pattern of partitions
<code>stp1</code>	step size of hyperparameter alpha (shared by all units) in one step EM
<code>stp2</code>	step size of hyperparameter beta (unit specific) in one step EM

**Value**

posterior probability of mean expression pattern

**extractInfo**

*extract count matrix from SingleCellExperiment object*

**Description**

extract count matrix from SingleCellExperiment object

**Usage**

```
extractInfo(data)
```

**Arguments**

<code>data</code>	SingleCellExperiment object
-------------------	-----------------------------

**Value**

list of count matrix and condition vector

**Examples**

```
data(sim_dat)
dat <- extractInfo(sim_dat)
```

---

gCl	<i>gene_level cluster</i>
-----	---------------------------

---

**Description**

gene\_level cluster

**Usage**

```
gCl(data, bp)
```

**Arguments**

data	transcripts
bp	bioc parallel parameter

**Value**

return a matrix whose row represent gene specific cluster

---

genRClus	<i>generate random clusterings</i>
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---

**Description**

generate random clusterings

**Usage**

```
genRClus(D, a, K)
```

**Arguments**

D	distance matrix of cells
a	paramter for weights
K	number of subtypes

**Value**

random generated clustering of cells

**getDD** *index of DD genes under FDR control*

### Description

index of DD genes under FDR control

### Usage

```
getDD(pDD, FDR = 0.01)
```

### Arguments

pDD	probability of genes being DD
FDR	fdr to be controlled

### Value

index of positive genes

### Examples

```
p_dd <- c(0.01, 0.99, 0.7, 0.5)
getDD(p_dd)
```

**getsizeofDD** *number of DD genes under FDR control*

### Description

number of DD genes under FDR control

### Usage

```
getsizeofDD(pDD, FDR = 0.01)
```

### Arguments

pDD	estimated probability of being DD
FDR	fdr to be controlled

### Value

number of positive genes

**Examples**

```
p_dd <- c(0.1,0.99,1,0.05,0.05)
getSizeofDD(p_dd)
```

---

getZ1Z2

*function to get counts of cluster sizes at two conditions*

---

**Description**

function to get counts of cluster sizes at two conditions

**Usage**

```
getZ1Z2(ccl, cd)
```

**Arguments**

ccl	clustering label
cd	condition label

**Value**

return list of counts

---

gRef

*generate reference matrix*

---

**Description**

generate reference matrix

**Usage**

```
gRef(Posp)
```

**Arguments**

Posp	possible partition of data
------	----------------------------

**Value**

return a matrix indicate the refinement relation between different partitions.

<code>isRef</code>	<i>check refinement relation between two clusters</i>
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**Description**

check refinement relation between two clusters

**Usage**

```
isRef(x, y)
```

**Arguments**

<code>x</code>	a cluster
<code>y</code>	a cluster

**Value**

whether `x` refines `y`

<code>LL</code>	<i>likelihood function for hyperparameters estimation</i>
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**Description**

likelihood function for hyperparameters estimation

**Usage**

```
LL(param, x, d0)
```

**Arguments**

<code>param</code>	parameters to be determined by MLE
<code>x</code>	distance matrix of cells
<code>d0</code>	rate parameter of prior of 1 / true distance

**Value**

return hyperparameteres a.

<code>lpt1t2</code>	<i>log likelihood of z1,z2 given t1,t2</i>
---------------------	--------------------------------------------

### Description

log likelihood of z1,z2 given t1,t2

### Usage

```
lpt1t2(z1, z2, pp, alpha1, alpha2)
```

### Arguments

<code>z1</code>	counts of each group in condition 1
<code>z2</code>	counts of each group in condition 2
<code>pp</code>	a partition
<code>alpha1</code>	parameter of double dirichlet prior
<code>alpha2</code>	parameter of double dirichlet prior

### Value

log likelihood of z1,z2 given t1,t2

<code>lpzgt</code>	<i>log likelihood of aggregated multinomial counts z given aggregated proportions t</i>
--------------------	-----------------------------------------------------------------------------------------

### Description

log likelihood of aggregated multinomial counts z given aggregated proportions t

### Usage

```
lpzgt(z, pp, alpha)
```

### Arguments

<code>z</code>	counts of each group in one condition
<code>pp</code>	a partition
<code>alpha</code>	parameter of double dirichlet prior

### Value

log likelihood of aggregated multinomial counts z given aggregated proportions t

<code>mdd</code>	<i>posterior of proportion change given mixture double dirichlet prior</i>
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### Description

posterior of proportion change given mixture double dirichlet prior

### Usage

```
mdd(z1, z2, pat, alpha1, alpha2)
```

### Arguments

<code>z1</code>	counts of each group in condition 1
<code>z2</code>	counts of each group in condition 2
<code>pat</code>	partition patterns
<code>alpha1</code>	parameter of double dirichlet prior
<code>alpha2</code>	parameter of double dirichlet prior

### Value

posterior of proportion change

<code>pat</code>	<i>generating partition patterns</i>
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### Description

generating partition patterns

### Usage

```
pat(K)
```

### Arguments

<code>K</code>	number of elements
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### Value

all possible partition of K elements

### Examples

```
pat(3)
```

---

pdd	<i>calculate posterior probabilities of a gene to be differential distributed</i>
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---

**Description**

calculate posterior probabilities of a gene to be differential distributed

**Usage**

```
pdd(
  data,
  cd,
  bp,
  D,
  random = TRUE,
  norm = TRUE,
  epi = 1,
  Upper = 1000,
  nrandom = 50,
  iter = 20,
  reltol = 0.001,
  stp1 = 1e-06,
  stp2 = 0.01,
  K = 0
)
```

**Arguments**

data	normalized preprocessed transcripts
cd	conditions label
bp	bioc parallel parameter
D	distance matrix of cells or cluster of cells or a given clustering
random	boolean indicator of whether randomization has been implemented on distance matrix
norm	boolean indicator of whether the input expression data is normalized
epi	tol for change of validity score in determining number of clusters
Upper	bound for hyper parameters optimization
nrandom	number of random generated distance matrix
iter	max number of iterations for EM
reltol	relative tolerance for optim on weighting paramters
stp1	step size of hyperparameter alpha (shared by all units) in one step EM
stp2	step size of hyperparameter beta (unit specific) in one step EM
K	number of subtypes, could be user specified or determined internally(set to 0)

**Value**

posterior probabilities of a gene to be differential distributed

**Examples**

```
data(sim_dat)
dat <- extractInfo(sim_dat)
data_counts <- dat$count_matrix
cd <- dat$condition
bp <- BiocParallel::MulticoreParam(4)
D_c <- calD(data_counts, bp)
pDD <- pdd(data_counts, cd, bp, D_c)
```

**pddAggregate**

*function to aggregate intermediate results and get prob of DD*

**Description**

function to aggregate intermediate results and get prob of DD

**Usage**

```
pddAggregate(z1, z2, Posp, DE, K, REF)
```

**Arguments**

<code>z1</code>	counts of cluster sizes in condition 1
<code>z2</code>	counts of cluster sizes in condition 2
<code>Posp</code>	partition of cells
<code>DE</code>	posterior probabilities of DE patterns
<code>K</code>	number of clusters
<code>REF</code>	reference matrix indicating relation of nested partitions

**Value**

return vector of prob of DD

---

pddRandom	<i>calculate PDD when add random noise in distance matrix</i>
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---

**Description**

calculate PDD when add random noise in distance matrix

**Usage**

```
pddRandom(data, cd, K, D, a, sz, hp, Posp, iter, REF, stp1, stp2)
```

**Arguments**

data	normalized preprocessed transcripts
cd	condition label
K	number of subgroups
D	distance matrix of cells
a	shape param for weights
sz	size factors
hp	hyper parameters for EBSeq
Posp	partition patterns
iter	max number of iterations for EM in EBSeq
REF	refinement relation matrix
stp1	step size of hyperparameter alpha (shared by all units) in one step EM
stp2	step size of hyperparameter beta (unit specific) in one step EM

**Value**

posterior probabilities under random distance matrix

---

rwMle	<i>MLE for random weighting parameter</i>
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**Description**

MLE for random weighting parameter

**Usage**

```
rwMle(D, reltol)
```

**Arguments**

D, distance matrix of cells  
reltol, tolerance of convergence

**Value**

MLE of random weighting parameter

---

sim\_dat *scDDboost*

---

**Description**

simulated data for demonstration, data are mixture negative binomial distributed

**Usage**

```
data(sim_dat)
```

**Format**

An object of class "list".

**Examples**

```
data(sim_dat)
```

# Index

- \* **Empirical Bayes, clustering, random weighting, local false discovery rate**
  - scDDboost-package, 2
- \* **datasets**
  - sim\_dat, 16
- \* **internal**
  - pddRandom, 15

calD, 3  
clusHelper, 4  
  
detK, 5  
  
EBS, 5  
extractInfo, 6  
  
gC1, 7  
genRClus, 7  
getDD, 8  
getsizeofDD, 8  
getZ1Z2, 9  
gRef, 9  
  
isRef, 10  
  
LL, 10  
lpt1t2, 11  
lpzgt, 11  
  
mdd, 12  
  
pat, 12  
pdd, 13  
pddAggregate, 14  
pddRandom, 15  
  
rwMle, 15  
  
scDDboost (scDDboost-package), 2  
scDDboost-package, 2  
sim\_dat, 16