Package 'synlet'

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Type Package

Title Hits Selection for Synthetic Lethal RNAi Screen Data

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Description Select hits from synthetic lethal RNAi screen data. For example, there are two identical celllines except one gene is knocked-down in one cellline. The interest is to find genes that lead to stronger lethal effect when they are knocked-down further by siRNA. Quality control and various visualisation tools are implemented. Four different algorithms could be used to pick up the interesting hits. This package is designed based on 384 wells plates, but may apply to other platforms with proper configuration.

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LazyData TRUE

biocViews ImmunoOncology, CellBasedAssays, QualityControl, Preprocessing, Visualization, FeatureExtraction

Depends R (>= 3.2.0), ggplot2

Imports doBy, dplyr, grid, magrittr, RColorBrewer, RankProd, reshape2

Suggests knitr, testthat, rmarkdown

VignetteBuilder knitr

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bScore Calculate B-score

Description

Calculate the B-score for plates belonging to the same master plate. Positive / negative controls are removed from the calculation.

Usage

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bScore(masterPlate, dat, treatment, control, outFile = FALSE)

Arguments

masterPlate a maste plate to be normalized
dat synthetic lethal RNAi screen data
treatment the treatment experiment condition in EXPERIMENT_MODIFICATION
control the control experiment condition in EXPERIMENT_MODIFICATION
outFile should calculated B-score files be written to the current folder? File names is
(masterPlate).bscore.csv.

Value

A list contains B-score for each master plate, treatment plates are the first columns, followed by control plates

References

Brideau, C., Gunter, B., Pikounis, B. & Liaw, A. Improved statistical methods for hit selection in high-throughput screening. J. Biomol. Screen. 8, 634-647 (2003).

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Examples

```
bscore.res <- sapply(as.character(unique(exampleDat$MASTER_PLATE)), bScore,
  exampleDat, control = "control", treatment = "treatment", simplify = FALSE)</pre>
```

exampleDat

Synthetic lethal RNAi screen example data.

Description

A dataset containing synthetic lethal RNAi screen data to show how functions work. The variables are as follows:

Usage

```
data(exampleDat)
```

Format

A data frame with 4320 rows and 8 variables

Details

- PLATE. plate names.
- MASTER_PLATE. master plate names.
- WELL_CONTENT_NAME. siRNA targets of wells.
- EXPERIMENT_TYPE. sample, negative/positive controls.
- EXPERIMENT_MODIFICATION. experiment conditions, "treatment" or "control".
- ROW_NAME. row names of plates.
- COL_NAME. column names of plates.
- READOUT. screen results.

Value

A data frame containing RANi screen data, the READOUT value has no real biological meaning.

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Select hits basing on median +- k*MAD

Description

Select hits basing on median +- k*MAD, by default k is three.

Usage

```
madSelect(masterPlate, dat, k = 3, treatment, control, outFile = FALSE,
    normMethod = "PLATE")
```

Arguments

masterPlate the master plate to analysis
dat synthetic lethal RNAi screen data
k cutoff for selecting hits, default is three

treatment the treatment condition in EXPERIMENT_MODIFICATION control the control condition in EXPERIMENT_MODIFICATION

outFile whether or not write the median normalized results

normMethod normalization methods to be used. If "PLATE", the raw readouts are normalized

by plate median, otherwise use median provided control siRNA.

Value

A data frame contains the hits selection results.

- MASTER PLATE: location of siRNA
- treat_cont_ratio: ratio of treatment / control
- treat_median: median value of treatment plates
- control_median: median value of control plates
- Hits: Is this siRNA a hit?

References

Chung, N. et al. Medianabsoluted eviation to improve hitselection forgenomes cale RNA is screens. J. Biomol. Screen. 13, 149-158 (2008).

Examples

```
madSelection <- sapply(as.character(unique(exampleDat$MASTER_PLATE)),
  madSelect, exampleDat, control = "control",
  treatment = "treatment", simplify = FALSE)
madSelection.c <- do.call(rbind,
  lapply(names(madSelection), function(x) madSelection[[x]]))</pre>
```

plateHeatmap 5

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Description

Put all individual plates in one graph, values are the readout in experiments.

Usage

```
plateHeatmap(dat, baseSize = 12)
```

Arguments

dat synthetic lethal RNAi screen data

baseSize basic font size used for x/y axis and title for heatmaps

Value

```
a ggplot object
```

Examples

```
tem.1 <- plateHeatmap(exampleDat)
ggsave("platesHeatmap.pdf", plot = tem.1, width = 500, height = 500, limitsize = FALSE)</pre>
```

rankProdHits

Select hits by the rank product method

Description

Select hits by rank product methods by comparing treatment and control.

Usage

```
rankProdHits(masterPlate, dat, treatment, control, normMethod = "PLATE")
```

Arguments

masterPlate the master plate to be analyzed dat synthetic lethal RNAi screen data

treatment the treatment condition in EXPERIMENT_MODIFICATION control the control condition in EXPERIMENT_MODIFICATION

normMethod normalization methods to be used. If "PLATE", the raw readouts are normalized

by plate median, otherwise use provided control siRNA

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Value

A list contains results by the rank product method for each master plate.

- MASTER_PLATE: location of siRNA
- pvalue_treat_lowerthan_cont: p-value for the hypothesis that treatment has lower normalized readout compared to control
- FDR_treat_lowerthan_cont: FDR value
- treat_cont_log2FC: log2 fold change of treatment / control

References

Breitling, R., Armengaud, P., Amtmann, A. & Herzyk, P. Rank products: a simple, yet powerful, new method to detect differentially regulated genes in replicated microarray experiments. FEBS Lett 573, 83-92 (2004).

Hong, F. et al. RankProd: a bioconductor package for detecting differentially expressed genes in meta-analysis. Bioinformatics 22, 2825-2827 (2006).

Examples

```
rankp.res <- sapply(as.character(unique(exampleDat$MASTER_PLATE)),
   rankProdHits, exampleDat, control = "control", treatment = "treatment",
   simplify = FALSE)
rankp.c <- data.frame(do.call(rbind,
   lapply(names(rankp.res), function(x) rankp.res[[x]])))</pre>
```

rsaHits

Select hits by RSA

Description

Selected hits by redundant siRNA activity method. Here is a wrapper function of RSA 1.8 by Yingyao Zhou.

Usage

```
rsaHits(dat, treatment, control, normMethod = "PLATE", LB, UB,
revHits = FALSE, Bonferroni = FALSE, outputFile = "RSAhits.csv",
scoreFile = "RSA_score.csv")
```

Arguments

dat synthetic lethal RNAi screen data

treatment the treatment condition in EXPERIMENT_MODIFICATION control the control condition in EXPERIMENT_MODIFICATION

normMethod normalization methods. If "PLATE", then values are normalized by plate me-

dian, otherwise use the provided control siRNA

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LB	Low bound
UB	up bound
revHits	reverse hit picking, default the lower the score the better
Bonferroni	conceptually useful when there are different number of siRNAs per gene, default \ensuremath{FALSE}
outputFile	output file name
scoreFile	name of the score file to be written under the current folder

Value

A result file written to the current folder.

- Gene_ID,Well_ID,Score: columns from input spreadsheet
- LogP: OPI p-value in log10, i.e., -2 means 0.01
- OPI_Hit: whether the well is a hit, 1 means yes, 0 means no
- #hitWell: number of hit wells for the gene
- #totalWell: total number of wells for the gene. If gene A has three wells w1, w2 and w3, and w1 and w2 are hits, #totalWell should be 3, #hitWell should be 2, w1 and w2 should have OPI_Hit set as 1 and w3 should have OPI_Hit set as 0.
- OPI_Rank: ranking column to sort all wells for hit picking
- Cutoff_Rank: ranking column to sort all wells based on Score in the simple activity-based method

Note: a rank value of 999999 means the well is not a hit

References

Koenig, R. et al. A probability-based approach for the analysis of large-scale RNAi screens. Nat Methods 4, 847-849 (2007).

Examples

```
rsaHits(exampleDat, treatment = "treatment", control = "control",
normMethod = "PLATE", LB = 0.2, UB = 0.8, revHits = FALSE,
Bonferroni = FALSE, outputFile = "RSAhits.csv")
```

scatterPlot

Scatter plot of RNAi screen results

Description

Produce a single plot for readous of each plate, with the option of highlighting specific signals, like positive/negative controls.

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Usage

```
scatterPlot(dat, controlOnly = FALSE, colour, ...)
```

Arguments

dat synthetic lethal RNAi screen data controlOnly whether or not to plot control wells only

colour colour for different signals

... positive/negative signals, must be specified

Value

a ggplot object

Examples

```
scatterPlot(exampleDat, controlOnly = FALSE, colour = rainbow(10),
   "PLK1 si1", "scrambled control si1", "lipid only")
```

siRNAPlot

Plot siRNA data and quality metrics.

Description

Plot the normalized RNAi screen data, row data, control signals and Z' factor.

Usage

```
siRNAPlot(gene, dat, controlsiRNA, FILEPATH = ".", colour = rainbow(10),
   zPrimeMed, zPrimeMean, treatment, control, normMethod = c("PLATE"),
   width = 15, height = 14)
```

Arguments

gene gene symbol, case sensitive
dat synthetic lethal RNAi screen data

controlsiRNA could be a vector of several siRNA, including postive/negative

control

FILEPATH path to store the figure colour colour used in graphs

zPrimeMed zPrime factor basing on median zPrimeMean, zPrime factor basing on mean

treatment the treatment condition in EXPERIMENT_MODIFICATION control the control condition in EXPERIMENT_MODIFICATION

normMethod could be a PLATE and negative controls

width width of the plot height height of the plot tTest 9

Value

Return the ggplot2 objects in a list, which could be plotted individually.

Examples

```
zF_mean <- zFactor(exampleDat, negativeCon = "scrambled control si1",
   positiveCon = "PLK1 si1")
zF_med <- zFactor(exampleDat, negativeCon = "scrambled control si1",
   positiveCon = "PLK1 si1", useMean = FALSE)
tem.1 <- siRNAPlot("AAK1", exampleDat,
   controlsiRNA = c("lipid only", "scrambled control si1"),
   FILEPATH = ".", zPrimeMed = zF_med, zPrimeMean = zF_mean,
   treatment = "treatment", control = "control",
   normMethod = c("PLATE", "lipid only", "scrambled control si1"))</pre>
```

tTest

student's t-test basing on B-score

Description

Select hits by student's t-test using B-score from treatment and control plates.

Usage

```
tTest(masterPlate, bScore, numTreat, numCont)
```

Arguments

masterPlate the master plate to be analyzed

bScore normalized bScore

numTreat number of treatment plates numCont number of control plates

Value

A list containing student's t-test for each master plate

- pvalue: p-value of the t-test
- Treat_Cont: difference in bscore: treatment control
- p_adj: BH adjusted p-value

References

Birmingham, A. et al. Statistical methods for analysis of high-throughput RNA interference screens. Nat Methods 6, 569-575 (2009).

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Examples

```
bscore.res <- sapply(as.character(unique(exampleDat$MASTER_PLATE)), bScore,
   exampleDat, control = "control", treatment = "treatment", simplify = FALSE)
bscore.ttest <- sapply(names(bscore.res), tTest, bscore.res, numTreat = 3,
   numCont = 3, simplify = FALSE, USE.NAMES = TRUE)
bscore.combined <- data.frame(do.call(rbind, lapply(names(bscore.ttest),
   function(x) if (!is.null(bscore.ttest[[x]])) {data.frame(MASTER_PLATE = x,
   siRNAs = rownames(bscore.ttest[[x]]), bscore.ttest[[x]])})))</pre>
```

zFactor

Calcualte the Z and Z' factor

Description

calcualte the Z and Z' factor for each plate.

Usage

```
zFactor(dat, negativeCon, positiveCon, useMean = TRUE)
```

Arguments

dat synthetic lethal RNAi screen data.

negativeCon the negative control used in the WELL_CONTENT_NAME.

positiveCon the positive control used in the WELL_CONTENT_NAME.

use Mean use mean to calcualate z factor and z' factor by default; otherwise use median.

Value

A data.frame contains z factor and z' factor

References

Zhang J.H., Chung T.D. & Oldenburg K.R. A simple statistical parameter for use in evaluation and validation of high throughput screening assays. J. Biomol. Screen. B, 4 67-73 (1999). Birmingham, A. et al. (2009) Statistical methods for analysis of high-throughput RNA interference screens. Nat Methods, 6, 569-575.

Examples

```
zFactor(exampleDat, negativeCon = "scrambled control si1", positiveCon = "PLK1 si1")
```

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