

Package ‘Spaniel’

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Type Package

Title Spatial Transcriptomics Analysis

Version 1.2.0

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Description Spaniel includes a series of tools to aid the quality control and analysis of Spatial Transcriptomics data. The package contains functions to create either a Seurat object or SingleCellExperiment from a count matrix and spatial barcode file and provides a method of loading a histological image into R. The spanielPlot function allows visualisation of metrics contained within the S4 object overlaid onto the image of the tissue.

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Encoding UTF-8

LazyData true

Depends R (>= 3.6), Seurat, SingleCellExperiment,
SummarizedExperiment, dplyr

Imports methods, ggplot2, scater (>= 1.13.27), shiny, jpeg, magrittr,
utils, S4Vectors

Suggests knitr, rmarkdown, testthat, devtools

VignetteBuilder knitr

RoxxygenNote 6.1.1.9000

Collate 'utilities.R' 'addClusterCols.R' 'parseImage.R' 'readData.R'
'removeSpots.R' 'spaniel_plot_internals.R' 'spatialPlot.R'
'shinySpaniel.R'

biocViews SingleCell, RNASeq, QualityControl, Preprocessing,
Normalization, Visualization, Transcriptomics, GeneExpression,
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R topics documented:

createSCE	2
createSeurat	3
markClusterCol	4
parseImage	4
removeSpots	5
runShinySpaniel	5
selectSpots	6
spanielPlot	7

Index

9

createSCE	<i>Create a SingleCellExperiment Object From Spatial Transcriptomics Data</i>
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Description

This function converts a count matrix into a SingleCellExperiment object. The barcodes for each spot are added to the coldata of the SingleCellExperiment object and are used in plotting the data.

Usage

```
createSCE(counts, barcodeFile, projectName=projectName,
          sectionNumber=sectionNo)
```

Arguments

counts	Raw count matrix or data frame where each row represents a gene and each column represents barcoded location on a spatial transcriptomics slide. The columns should be named using the spot barcode (eg "GTCCGATATGATTGC-CGC")
barcodeFile	a tab seperated barcode file supplied by Spatial Trancscriptomics. The file should contains three column: The first column contains the Spatial Transcriptomics barcode, the second and third column equate to the x and y location
projectName	The name of the project which is stored in the Seurat Object.
sectionNumber	The location of the sample on the slide

Value

A SingleCellExeriment Object

Examples

```
## Data is taken from DOI: 10.1126/science.aaf2403
examplecounts <- readRDS(file.path(system.file(package = "Spaniel"),
                                    "extdata/counts.rds"))
exampleBarcodes <- file.path(system.file(package = "Spaniel"),
                               "1000L2_barcodes.txt")
seurat0b <- createSCE(examplecounts,
                       exampleBarcodes,
                       projectName = "TestProj",
                       sectionNumber = 1)
```

createSeurat*Create a Seurat Object From Spatial Transcriptomics Data*

Description

This function converts a count matrix into a Seurat object. The barcodes for each spot are added to the metadata of the Seurat object and are used in plotting the data.

Usage

```
createSeurat(counts, barcodeFile, projectName = projectName,
             sectionNumber = sectionNo)
```

Arguments

counts	Raw count matrix or data frame where each row represents a gene and each column represents barcoded location on a spatial transcriptomics slide. The columns should be named using the spot barcode (eg "GTCCGATATGATTGC-CGC")
barcodeFile	a tab seperated barcode file supplied by Spatial Trancscriptomics. The file should contains three column: The first column contains the Spatial Transcriptomics barcode, the second and third column equate to the x and y location
projectName	The name of the project which is stored in the Seurat Object.
sectionNumber	The location of the sample on the slide

Value

A Seurat Object

Examples

```
## Data is taken from DOI: 10.1126/science.aaf2403
examplecounts <- readRDS(file.path(system.file(package = "Spaniel"),
                                    "extdata/counts.rds"))
exampleBarcodes <- file.path(system.file(package = "Spaniel"),
                               "1000L2_barcodes.txt")
SeuratObj <- createSeurat(examplecounts,
                           exampleBarcodes,
                           projectName = "TestProj",
                           sectionNumber = 1
                           )
```

`markClusterCol` *markClusterCol*

Description

A function to mark the columns containing cluster information in the metadata or colData of a Seurat or SCE object. Columns are marked with "cluster_" prefix.

Usage

```
markClusterCol(object, pattern)
```

Arguments

<code>object</code>	Either a Seurat or SCE object containing clustering information
<code>pattern</code>	pattern indicating which columns contain cluster information

Value

A Seurat or SCE object

Examples

```
SeuratObj <- readRDS(file.path(system.file(package = "Spaniel"),
                                "extdata/SeuratData.rds"))
SeuratObj <- markClusterCol(SeuratObj, "res")
```

`parseImage` *This function parses a HE image to use as the background for plots*

Description

This function parses a HE image to use as the background for plots

Usage

```
parseImage(imgFile)
```

Arguments

<code>imgFile</code>	Path to the image file
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Value

A rasterized grob

Examples

```
imgFile <- file.path(system.file(package = "Spaniel"),
                      "HE_Rep1_resized.jpg")
img <- parseImage(imgFile)
```

`removeSpots`*removeSpots*

Description

A function to filter spots from analysis. It requires selectSpots to be run first.

Usage

```
removeSpots(sObj, pointsToRemove = "points_to_remove.txt")
```

Arguments

`sObj` Either a Seurat object (version 3) or a SingleCellExperiment object containing barcode coordinates in the metadata (Seurat) or colData (SingleCellExperiment).

`pointsToRemove` path to points to remove file. Default is "points_to_remove.txt"

Value

A filtered Seurat or SingleCellExperiment Object

Examples

```
seuratObj <- readRDS(file.path(system.file(package = "Spaniel"),
                                "extdata/SeuratData.rds"))
toRemove <- file.path(system.file(package = "Spaniel"),
                      "points_to_remove.txt")
sObjFiltered <- removeSpots(sObj = seuratObj, pointsToRemove = toRemove)
```

`runShinySpaniel`*RunShinySpaniel*

Description

A function to visualise Spatial Transcriptomics. It requires a preprocessed Seurat Object or a SingleCellExperiment object as well as a rasterised image saved as an .rds object. There are 4 plots available in the app showing: a) the number of genes detected per spot, b) the number of reads detected per spot, c) clustering results, d) the gene expression of a selected gene." To view the clustering results the columns of the meta.data or colData containing clustering results must be prefixed with cluster_. This can be done by using the markClusterCol() function included in Spaniel.

Usage

```
runShinySpaniel()
```

Value

Runs a Shiny App

Examples

```

## mark the columns of metadata/colData that contain clustering
## information see ?markClusterCol for more details'
sObj <- readRDS(file.path(system.file(package = "Spaniel"),
                           "extdata/SeuratData.rds"))
sObj <- markClusterCol(sObj, "res")

### parse background image
imgFile <- file.path(system.file(package = "Spaniel"),
                      "HE_Rep1_resized.jpg")
img <- parseImage(imgFile)

## run shinySpaniel (upload data.rds and image.rds in the shiny app)
## Not Run:
# runShinySpaniel()

```

selectSpots

selectSpots

Description

A function to select spots to remove from analysis

Usage

```
selectSpots(sObj, imgObj)
```

Arguments

sObj	Either a Seurat object (version 3) or a SingleCellExperiment object containing barcode coordinates in the metadata (Seurat) or colData (SingleCellExperiment).
imgObj	a ggplot grob (see parseImage function)

Value

Runs a shiny application

Examples

```

## Run the shiny app (Not run):
# selectSpots(sObj, imgObj)

# Click on the spots to remove from downstream analysis. Once all the spots
# have been selected close the shiny app window. A list of spots is
# stored in a text file called points_to_remove.txt in the working directory.

# Once this step has been run a filtered Seurat or SCE object can be
# created using removeSpots (see removeSpots for more details)

```

spanielPlot
Spatial Transcriptomics Plot

Description

This function overlays information from a Seurat object or SingleCellExperiment object containing barcodes onto a H & E image. There are 4 plots available showing a) the number of genes detected per spot, b) the number of reads detected per spot, c) clustering results, d) the gene expression of a selected gene.

Usage

```
spanielPlot(object, grob, plotType = c("NoGenes",
                                         "CountsPerSpot",
                                         "Cluster",
                                         "Gene"),
            gene= NULL, clusterRes = NULL, customTitle = NULL,
            scaleData = TRUE, showFilter = NULL, ptSize = 2,
            ptSizeMin = 0, ptSizeMax = 5)
```

Arguments

<code>object</code>	Either a Seurat object (version 3) or a SingleCellExperiment object containing barcode coordinates in the metadata (Seurat) or colData (SingleCellExperiment).
<code>grob</code>	an grob to be used as the background image see(parseImage)
<code>plotType</code>	There are 5 types of plots available: 1) NoGenes - This shows the number of genes per spot and uses information from "nFeature_RNA" column of Seurat object or "detected" from a SingleCellExperiment object. 2) CountsPerSpot - This shows the number of counts per spot. It uses information from "nCount_RNA" column of Seurat object or "sum" from a singleCellExperiment object. 3) Cluster - This plot is designed to show clustering results stored in the meta.data or colData of an object 4) Gene- This plot shows the expression of a single gene. This plot uses scaled/normalised expressin data from the scale.data slot of Seurat object or logcounts of a SingleCellExperiment object. 5) Other - A generic plot to plot any column from the meta.data or colData of an object.
<code>gene</code>	Gene to plot
<code>clusterRes</code>	which cluster resolution to plot
<code>customTitle</code>	Specify plot title (optional)
<code>scaleData</code>	Show scaled data on plot (default is TRUE)
<code>showFilter</code>	Logical filter showing pass/fail for spots
<code>ptSize</code>	Point size used for cluster plot default is 2
<code>ptSizeMin</code>	Minimum point size used for QC and Gene Expression plots default is 0
<code>ptSizeMax</code>	Maximum point size used for QC and Gene Expression plots default is 5

Value

A ggplot spatial transcriptomics plot

Examples

```
## Data is taken from DOI: 10.1126/science.aaf2403
SeuratObj <- readRDS(file.path(system.file(package = "Spaniel"),
                                "extdata/SeuratData.rds"))
imgFile <- readRDS(file.path(system.file(package = "Spaniel"),
                                "extdata/image.rds"))

## Counts per spot with a QC filter
minGenes <- 2000
minUMI <- 300000
filter <- SeuratObj$nFeature_RNA > minGenes &
           SeuratObj$nCount_RNA > minUMI
spanielPlot(object = SeuratObj, grob = imgFile,
            plotType = "CountsPerSpot",
            showFilter = filter)

## Cluster plot
spanielPlot(object = SeuratObj, grob = imgFile,
            plotType = "Cluster",
            clusterRes = "cluster_RNA_snn_res.0.6")

## Gene plot
spanielPlot(object = SeuratObj, grob = imgFile,
            plotType = "Gene",
            gene= "Nrgn")
```

Index

createSCE, 2
createSeurat, 3
markClusterCol, 4
parseImage, 4
removeSpots, 5
runShinySpaniel, 5
selectSpots, 6
spanielPlot, 7