Package 'waveTiling'

October 9, 2013

Version 1.2.0

Date 2012-05-14

License GPL (>=2)

Title Wavelet-Based Models for Tiling Array Transcriptome Analysis

Author Kristof De Beuf kristof.debeuf@UGent.be, Peter Pipelers <a href="mailto:kristof.debeuf@UGent.be, Peter Pipelers <a href="mailto:kristof.debeuf@UGent.be, Peter Pipelers <a href="mailto:kristof.debeuf@

Maintainer Kristof De Beuf <kristof.debeuf@UGent.be>

Depends oligo, oligoClasses, Biobase, Biostrings, GenomeGraphs

Imports methods, affy, preprocessCore, GenomicRanges, waveslim, IRanges

Suggests BSgenome, BSgenome.Athaliana.TAIR.TAIR9, waveTiling-Data,pd.atdschip.tiling, TxDb.Athaliana.BioMart.plantsmart12

Description

This package is designed to conduct transcriptome analysis for tiling arrays based on fast wavelet-based functional models.

Collate allClasses.R allGenerics.R helperFunctions.R initialize-methods.R methods-MapFilterProbe.R methods-WaveTilingFeatureSet.R methods-WfmFit.R methods-WfmInf.R show-methods.R

URL https://r-forge.r-project.org/projects/wavetiling/

LazyLoad yes

LazyData

biocViews MicroArray, DifferentialExpression, TimeCourse, GeneExpression

2 addPheno

R topics documented:

ndex		3
	WfmInfOverallMean-class	3.
	WfmInfMeans-class	
	WfmInfEffects-class	
	WfmInfCustom-class	
	WfmInfCompare-class	
	WfmInf-class	
	WfmFitTime-class	
	WfmFitFactor-class	
	WfmFitCustom-class	
	WfmFitCircadian-class	
	WfmFit-class	19
	wfm.inference	1'
	wfm.fit	10
	WaveTilingFeatureSet-class	14
	selectProbesFromTilingFeatureSet	1.
	selectProbesFromFilterOverlap	12
	plotWfm	1
	MapFilterProbe-class	9
	makeDesign	9
	makeContrasts	8
	getSigGenes	1
	getNonAnnotatedRegions	(
	GenomeInfo-class	(
	filterOverlap	
	cel2TilingFeatureSet	2
	bgCorrQn	3
	addPheno	-

Description

 $Function\ to\ add\ phenotypic\ information\ such\ as\ sample\ names\ or\ group\ names\ to\ a\ Wave \verb|TilingFeatureSetclass|\ object$

Usage

```
addPheno(object, noGroups, groupNames, replics, ...)
```

bgCorrQn 3

Arguments

object of class WaveTilingFeatureSet

noGroups Number of groups in the tiling array experiment groupNames Vector containing the group or sample names in the ti

Vector containing the group or sample names in the tiling array experiment. The

vector length should be equal to the indicated number of groups.

replics Numeric vector containing the number of replicates for each group. The vector

length should be equal to the indicated number of groups.

... other arguments

Value

object of class WaveTilingFeatureSet annotated with the phenotypic data

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
## Not run:
    data(leafdev)
    leafdev <- as(leafdev,"WaveTilingFeatureSet")
    leafdev <- addPheno(leafdev,noGroups=6,groupNames=c("day8","day9","day10","day11","day12","day13"),replics=re|
    leafdev
## End(Not run)</pre>
```

bgCorrQn Background correction and quantile normalization

Description

Function to perform background correction and quantile normalization of tiling arrays

Usage

```
bgCorrQn(object, useMapFilter=NULL)
```

Arguments

object of class WaveTilingFeatureSet

useMapFilter NULL or object of class mapFilterProbe indicating the probes to use for back-

ground correction and quantile normalization

cel2TilingFeatureSet

Value

4

 $object\ of\ class\ Wave Tiling Feature Set\ containing\ the\ background-corrected\ and\ quantile-normalized\ intensity\ signals$

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
## Not run:
    data(leafdev)
    leafdev <- as(leafdev,"WaveTilingFeatureSet")
    data(leafdevMapAndFilterTAIR9)
    leafdevBQ <- bgCorrQn(leafdev,useMapFilter=leafdevMapAndFilterTAIR9)
## End(Not run)</pre>
```

cel2TilingFeatureSet Read CEL-files to TilingFeatureSet

Description

Wrapper function to read in CEL-files and output their content as a TilingFeatureSet-class object.

Usage

```
cel2TilingFeatureSet(dataPath, annotationPackage)
```

Arguments

dataPath character indicating the data path where the CEL-files to read in are stored annotationPackage

name of the package containing the array probe and annotation information, as produces by the pdInfoBuilder-package

Details

Uses the list.celfiles and read.celfiles functions of the oligo-package.

Value

```
object of class TilingFeatureSet
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

filterOverlap 5

Examples

```
## No example
```

filterOverlap function to filter probe seq	uence overlaps and remap probes
filterOverlap function to filter probe seq	uence overlaps and remap probes

Description

This function remaps the probe sequence to a reference sequence and filters probe sequence overlaps between PM and MM probes and/or between probes on the forward and reverse strand

Usage

```
filterOverlap(object, remap = TRUE, BSgenomeObject, chrId, strand = c("forward", "reverse", "both"), MM=
```

Arguments

object	object of class WaveTilingFeatureSet
remap	logical to determine whether the tiling array probe sequences have to be remapped to a more recent reference DNA sequence
BSgenomeObject	object of class BSgenome containing the genome sequence of the species for which the probes need to be filtered and remapped
chrId	vector of numerics identifying the chromosomes for which the probes have to be filtered and/or remapped
strand	character indicating the strands for which the probes have to be filtered and/or remapped (forward, reverse or both)
MM	logical to indicate whether the tiling array contains MM probes or not

Value

An object of class mapFilterProbe is returned containing the indices of the filtered probes.

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

other arguments

Examples

```
## Not run:
    data(leafdev)
    as(leafdev, "WaveTilingFeatureSet")
    library(BSgenome.Athaliana.TAIR.TAIR9)
    leafdevMapAndFilterTAIR9 <- filterOverlap(leafdev,remap=TRUE,BSgenomeObject=Athaliana,chrId=1:7,strand="both",
## End(Not run)</pre>
```

GenomeInfo-class

Class "genomeInfo"

Description

class to store the genomic info from a WfmFit-class object

Objects from the Class

Objects can be created by calls of the form new("GenomeInfo", chromosome, strand, minPos, maxPos).

Slots

```
chromosome: Object of class "vector" ~~
strand: Object of class "character" ~~
minPos: Object of class "numeric" ~~
maxPos: Object of class "numeric" ~~
```

Methods

```
initialize signature(.Object = "GenomeInfo"): ...
show signature(.Object = "GenomeInfo"): ...
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("GenomeInfo")
```

getNonAnnotatedRegions

Get non-annotated regions

Description

Extract the significant regions found in the wavelet-based transcriptome analysis that don't show any overlap with the existing annotation.

Usage

```
getNonAnnotatedRegions(fit,inf,biomartObj)
```

getSigGenes 7

Arguments

fit object of class WfmFit inf object of class WfmInf

biomart0bj object of class TranscriptDb representing an annotation database generated from

BioMart.

Value

GRangesList object with the non-annotated regions. The first element gives the regions with no annotation overlap on the strand used in the analysis, the second element gives the regions with no annotation overlap on both strands.

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
library(waveTilingData)
library(TxDb.Athaliana.BioMart.plantsmart12)
data(leafdevFit)
data(leafdevInfCompare)
nonAnnoCompare <- getNonAnnotatedRegions(fit=leafdevFit,inf=leafdevInfCompare,biomartObj=TxDb.Athaliana.BioMannonAnnoCompare</pre>
```

getSigGenes Get significant genes

Description

Extract the annotated regions (often genes) that overlap with the significant regions found in the wavelet-based transcriptome analysis.

Usage

```
getSigGenes(fit,inf,biomartObj)
```

Arguments

fit object of class WfmFit inf object of class WfmInf

biomart0bj object of class TranscriptDb representing an annotation database generated from

BioMart.

8 makeContrasts

Value

GRangesList object. In the elementMetadata of the GRanges elements percoverGene gives the percentage of basepair overlap of the annotated regions by the detected significant region in the analysis; percoverReg gives the percentage of basepair overlap of the detected singificant region in the analysis with the annotated region; totPercoverGene gives per annotated region the total percentage of basepair overlap by all detected significant regions in the analysis that map to that annotated region.

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
library(waveTilingData)
library(TxDb.Athaliana.BioMart.plantsmart12)
data(leafdevFit)
data(leafdevInfCompare)
sigGenesCompare <- getSigGenes(fit=leafdevFit,inf=leafdevInfCompare,biomartObj=TxDb.Athaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.BioMart.plantsmartobaliana.B
```

makeContrasts

Construct contrast matrix

Description

Helper function to construct a contrast matrix to be used in the inference procedure of the waveletbased transcriptome analysis when conducting a pairwise comparison analysis.

Usage

```
makeContrasts(contrasts, nlevels)
```

Arguments

contrasts compare: contrasts for pairwise comparison analysis.

Number of groups for pairwise comparison analysis.

Value

numeric matrix

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
makeContrasts(contrasts="compare",nlevels=6)
```

makeDesign 9

makeDesign	Construct design matrix	

Description

Helper function to construct a design matrix to be used in the wavelet-based transcriptome analysis.

Usage

```
\verb|makeDesign(design=c("time","circadian","group","factorial")|, \verb|replics|, \verb|noGroups|, factor.levels=NULL|| \\
```

Arguments

design	character indicating the design of the tiling array experiment. Currently, the following designs are implemented: time for a time-course design based on polynomial contrasts; circadian for circadian rhythm analysis; group for unordered one-factor designs; factorial for two-factor designs
replics	Numeric vector containing the number of replicates for each group. The vector length should be equal to the indicated number of groups.
noGroups	Number of groups in the tiling array experiment
factor.levels	Factor levels to use if applying two-factor design

Value

numeric matrix

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
makeDesign(design="time",replics=rep(3,6),noGroups=6)
```

MapFilterProbe-class Class "MapFilterProbe"

Description

class to store probe information after remapping and/or filtering of probes.

Usage

```
## Accessors
getChromosome(object)
getFilteredIndices(object)
getPosition(object)
getStrand(object)
```

Arguments

object An instance of MapFilterProbe-class.

Objects from the Class

Objects can be created by calls of the form new("mapFilterProbe", filteredIndices, chromosome, position, strand

Slots

```
filteredIndices: Object of class "vector" ~~ chromosome: Object of class "vector" ~~ position: Object of class "vector" ~~ strand: Object of class "vector" ~~
```

Methods

```
getChromosome signature(object = "MapFilterProbe"): ...
getFilteredIndices signature(object = "MapFilterProbe"): ...
getPosition signature(object = "MapFilterProbe"): ...
getStrand signature(object = "MapFilterProbe"): ...
initialize signature(.Object = "MapFilterProbe"): ...
selectProbesFromFilterOverlap signature(object = "MapFilterProbe"): ...
show signature(object = "MapFilterProbe"): ...
```

Accessors

In the following code snippets, x is a MapFilterProbe object.

```
getChromosome(x): Extract the chromosome identifiers.
getFilteredIndices(x): Extract the filtered probe indices.
getPosition(x): Extract the genomic position of the filtered probes.
getStrand(x): Extract the strand orientation info for the filtered probes.
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

plotWfm 11

Examples

```
showClass("MapFilterProbe")
library(waveTilingData)
data(leafdevMapAndFilterTAIR9)
tt1 <- getChromosome(leafdevMapAndFilterTAIR9)
tt2 <- getFilteredIndices(leafdevMapAndFilterTAIR9)
tt3 <- getPosition(leafdevMapAndFilterTAIR9)
tt4 <- getStrand(leafdevMapAndFilterTAIR9)</pre>
```

plotWfm

plot model fit and genomic regions

Description

Plot function to visualize the results of the wavelet-based transcriptome analysis. Both the model fit and the significant genomic regions can be plotted and compared with the annotation.

Usage

plotWfm(fit, inf, biomartObj, minPos, maxPos, trackFeature="exon", two.strand=TRUE, plotData=TRUE, plotData=TRU

Arguments

fit object of class WfmFit inf object of class WfmInf

biomart0bj object of class TranscriptDb representing an annotation database generated from

BioMart.

minPos minimum genomic position to plot maxPos maximum genomic position to plot

trackFeature track feature. See GenomeGraphs-package. Default is exon.

two.strand logical indicating whether to plot two strands or not plotData logical indicating whether to plot the raw data or not

plotMean logical indicating whether to plot the fitted overall mean function or not

tracks vector of integers containing the track numbers to plot. Track numbers corre-

spond with the order of the elements in the list output from the getGenmicRegions-

function.

Details

The plot utilities of the GenomeGraphs-package constitute the backbone of the plotWfm function.

Value

nothing returned

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

References

[1] Durinck S, Bullard J, Spellman PT, Dudoit S: GenomeGraphs: integrated genomic data visualization with R. BMC Bioinformatics 2009, 10:Article 2.

Examples

```
library(waveTilingData)
library(TxDb.Athaliana.BioMart.plantsmart12)
data(leafdevFit)
data(leafdevInfCompare)
trs <- transcripts(TxDb.Athaliana.BioMart.plantsmart12)
sel <- trs[elementMetadata(trs)$tx_name %in% "AT1G62500.1",]
start <- start(ranges(sel))-2000
end <- end(ranges(sel))+2000
plotWfm(fit=leafdevFit,inf=leafdevInfCompare,biomartObj=TxDb.Athaliana.BioMart.plantsmart12,minPos=start,maxF</pre>
```

```
selectProbesFromFilterOverlap
```

select probes from MapFilterProbe object

Description

Extract the probe indices from a MapFilterProbe object that map to a region between two specified genomic positions

Usage

selectProbesFromFilterOverlap(object, chromosome, strand=c("forward","reverse"), minPos=min(getPosit

Arguments

object of class MapFilterProbe

chromosome chromosome

strand strand

minPos minimum genomic position
maxPos maximum genomic position

Value

A list of 2 elements is returned. The first element "selection" gives the probe indices in the filtered MapFilterProbe object; the second element "selectionInit" gives the probe indices in the original WaveTilingFeatureSet object.

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
library(waveTilingData)
data(leafdevMapAndFilterTAIR9)
tt <- selectProbesFromFilterOverlap(leafdevMapAndFilterTAIR9,chromosome=1,strand="forward",minPos=10000,maxPossel <- tt$selection
length(sel)
head(sel)
selfil <- tt$selectionFiltered
length(selfil)
head(selfil)</pre>
```

```
select Probes From Tiling Feature Set
```

select probes from WaveTilingFeatureSet object

Description

Extract the probe indices from a WaveTilingFeatureSet object that map to a region between two specified genomic positions

Usage

```
selectProbesFromTilingFeatureSet(object, chromosome, strand=c("forward", "reverse"), minPos, maxPos)
```

Arguments

object of class WaveTilingFeatureSet

chromosome chromosome

strand strand

minPos minimum genomic position
maxPos maximum genomic position

Value

vector of integers indicating the probe indices

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

Description

Class to store expression and phenotypic data from a tiling array experiment, used as input for the wavelet-based transcriptome analysis.

Usage

```
## Accessors
getGroupNames(object)
getNoGroups(object)
getReplics(object)
```

Arguments

object

An instance of WaveTilingFeatureSet-class.

Objects from the Class

Objects can be created by calls of the form new("WaveTilingFeatureSet").

Slots

```
manufacturer: Object of class "character" ~~
intensityFile: Object of class "character" ~~
assayData: Object of class "AssayData" ~~
phenoData: Object of class "AnnotatedDataFrame" ~~
featureData: Object of class "AnnotatedDataFrame" ~~
experimentData: Object of class "MIAME" ~~
annotation: Object of class "character" ~~
protocolData: Object of class "AnnotatedDataFrame" ~~
.__classVersion__: Object of class "Versions" ~~
```

Extends

Class "TilingFeatureSet", directly. Class "FeatureSet", by class "TilingFeatureSet", distance 2. Class "NChannelSet", by class "TilingFeatureSet", distance 3. Class "eSet", by class "TilingFeatureSet", distance 4. Class "VersionedBiobase", by class "TilingFeatureSet", distance 5. Class "Versioned", by class "TilingFeatureSet", distance 6.

Methods

```
addPheno signature(object = "WaveTilingFeatureSet"): ...
bgCorrQn signature(object = "WaveTilingFeatureSet"): ...
filterOverlap signature(object = "WaveTilingFeatureSet"): ...
getGroupNames signature(object = "WaveTilingFeatureSet"): ...
getNoGroups signature(object = "WaveTilingFeatureSet"): ...
getReplics signature(object = "WaveTilingFeatureSet"): ...
selectProbesFromTilingFeatureSet signature(object = "WaveTilingFeatureSet"): ...
wfm.fit signature(object = "WaveTilingFeatureSet"): ...
```

Accessors

In the following code snippets, x is a WaveTilingFeatureSet object. The described accessors are specific for WaveTilingFeatureSet-class objects. Other inherited accessors work as expected on this class.

```
getGroupNames(x): Extract the group or sample names in the tiling array experiment. getNoGroups(x): Extract the number of groups or samples in the tiling array experiment. getReplics(x): Extract the number of replicates in the tiling array experiment.
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WaveTilingFeatureSet")
library(waveTilingData)
data(leafdev)
leafdev <- as(leafdev,"WaveTilingFeatureSet")
leafdev <- addPheno(leafdev,noGroups=6,groupNames=c("day8","day9","day10","day11","day12","day13"),replics=rel
tt1 <- getGroupNames(leafdev)
tt2 <- getNoGroups(leafdev)
tt3 <- getReplics(leafdev)</pre>
```

16 wfm.fit

wfm.fit	Fit Wfm model to trancriptome data	
---------	------------------------------------	--

Description

Main function to fit a wavelet-based functional model to the tiling array expression data.

Usage

```
wfm.fit(object, filter.overlap=NULL, design=c("time","circadian","group","factorial","custom"), n.le
```

Arguments

8	
object	object of class WaveTilingFeatureSet
filter.overlap	object of class mapFilterProbe
design	character indicating the design of tiling array experiment. Currently, the following designs are implemented: time for a time-course design based on polynomial contrasts; circadian for circadian rhythm analysis; group for unordered one-factor designs; factorial for two-factor designs; custom for other designs. When using design="custom" a specific design.matrix needs to be given.
n.levels	number of levels in wavelet decomposition (integer)
factor.levels	factor levels in case of two-factor analysis. Vector of integers with length the number of factors in the experiment, and with elements the number of levels for the respective factors.
chromosome	numeric to indicate the chromosome associated with transcriptome data to fit
strand	character to indicate the strand orientation associated with transcriptome data to fit. Either "forward" or "reverse".
minPos	integer to indicate minimum genomic position
maxPos	integer to indicate maximum genomic position
design.matrix	custom design matrix to use
var.eps	character indicating how to estimate residual variance. Either "margLik" for marginal maximum likelihood based estimation or "mad" for estimation based on the MAD (more info see references).
prior	character indicating which prior distribution to put on effect functions. Either "normal" for a normally distributed prior, or "improper" for an improper prior (more info see references).
eqsmooth	logical indicating whether to force equal amount of smooth for different effect functions or not
max.it	integer giving the maximum number of iteration for estimation
wave.filt	character indicating which wavelet filter to use. Default is "haar".
skiplevels	integer indicating how many wavelet levels to put equal to 0
trace	logical indicating whether to trace estimation
save.obs	character to indicate which output to store in return object. Either "plot": all info needed to make the plots or "all": store all possible info.

wfm.inference 17

Value

object of class WfmFit

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

References

- [1] Clement L, De Beuf K, Thas O, Vuylsteke M, Irizarry RA and Crainiceanu CM. (2012) Fast wavelet based functional models for transcriptome analysis with tiling arrays. Statistical Applications in Genetics and Molecular Biology 11: Iss. 1, Article 4.
- [2] De Beuf K, Andriankaja, M, Thas O, Inze D, Crainiceanu CM and Clement L (2012) Model-based analysis of tiling array expression studies with flexible designs. Technical document.

Examples

```
library(waveTilingData)
data(leafdevBQ)
data(leafdevMapAndFilterTAIR9)
leafdevFit <- wfm.fit(leafdevBQ,filter.overlap=leafdevMapAndFilterTAIR9,design="time",n.levels=10,chromosome=</pre>
```

wfm.inference

Perform transcriptome analysis on fitted wavelet-based functional model

Description

Main function to perform trancriptome analysis on a fitted wavelet-based functional model of class WfmFit.

Usage

```
wfm.inference(object, contrast.matrix=NULL, contrasts=c("compare", "means", "effects", "overallMean"),
```

Arguments

object of class WfmFit

contrast.matrix

custom contrasts matrix

contrasts

character indicating the type of transcriptome analysis is to be applied. Currently the following types are implemented: compare for doing a pairwise differential expression analysis between any combination of two groups; effects, which corresponds to a circadian rhythm analysis if a circadian design is used for the fit, and to a time effects analysis (linear, quadratic,...) if a time-course design is used for the fit; means for doing a group-wise transcript discovery analysis.

18 wfm.inference

delta threshold value to be used in the inference procedure. This should be a nu-

meric vector with as first element the threshold for the overall mean transcript discovery and the other elements the threshold for the differential expression, the effects analysis or group-wise mean analysis. If the threshold should be equal for all comparisons, effects or group-wise means only a vector of length 2 is needed. Otherwise, the vector must be of length r+1 with r the number of

pairwise comparisons, effects or group-wise means.

two.sided logical indicating if one-sided or two-sided tests are desired

minRunPos minrun by position. An integer to indicate the minimum number of basepairs a

significant genomic region should contain.

minRunProbe minrun by probes. An integer to indicate the minimum number of probes the

significant genomic region should map to.

alpha significance level

nsim number of simulations used when doing circadian rhythm inference

rescale rescale matrix

Value

object of class WfmFit

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

References

- [1] Clement L, De Beuf K, Thas O, Vuylsteke M, Irizarry RA and Crainiceanu CM. (2012) Fast wavelet based functional models for transcriptome analysis with tiling arrays. Statistical Applications in Genetics and Molecular Biology 11: Iss. 1, Article 4.
- [2] De Beuf K, Andriankaja, M, Thas O, Inze D, Crainiceanu CM and Clement L (2012) Model-based analysis of tiling array expression studies with flexible designs. Technical document.

Examples

```
library(waveTilingData)
data(leafdevFit)
delta <- log(1.2,2)
leafdevInfCompare <- wfm.inference(leafdevFit,contrasts="compare",delta=c("median",delta))</pre>
```

WfmFit-class 19

WfmFit-class Class "WfmFit"

Description

class to store model fits in the wavelet-based transcriptome analysis.

Usage

```
## Accessors
getProbePosition(object)
getNoProbes(object)
getBetaWav(object)
getVarBetaWav(object)
getSmoothPar(object)
getVarEps(object)
getGenomeInfo(object)
getMinPos(object)
getMaxPos(object)
getNoLevels(object)
getDesignMatrix(object)
getPhenoInfo(object)
getDataOrigSpace(object)
getDataWaveletSpace(object)
getWaveletFilter(object)
getKj(object)
getPrior(object)
getF(object)
getVarF(object)
```

Arguments

object An instance of WfmFit-class.

Objects from the Class

Objects can be created by calls of the form new("WfmFit", betaWav, varbetaWav, smoothPar, varEps, dataOrigSpace

Slots

```
betaWav: Object of class "matrix" ~~ varbetaWav: Object of class "matrix" ~~ smoothPar: Object of class "matrix" ~~ varEps: Object of class "numeric" ~~
```

20 WfmFit-class

```
dataOrigSpace: Object of class "matrix" ~~
dataWaveletSpace: Object of class "matrix" ~~
design.matrix: Object of class "matrix" ~~
phenoData: Object of class "data.frame" ~~
genome.info: Object of class "genomeInfo" ~~
n.levels: Object of class "numeric" ~~
probePosition: Object of class "numeric" ~~
wave.filt: Object of class "character" ~~
Kj: Object of class "numeric" ~~
prior: Object of class "character" ~~
F: Object of class "matrix" ~~
varF: Object of class "matrix" ~~
P: Object of class "numeric" ~~
Z: Object of class "matrix" ~~
noGroups: Object of class "numeric" ~~
replics: Object of class "numeric" ~~
```

Methods

```
getBetaWav signature(object = "WfmFit"): ...
getChromosome signature(object = "WfmFit"): ...
getDataOrigSpace signature(object = "WfmFit"): ...
getDataWaveletSpace signature(object = "WfmFit"): ...
getDesignMatrix signature(object = "WfmFit"): ...
getF signature(object = "WfmFit"): ...
getGenomeInfo signature(object = "WfmFit"): ...
getKj signature(object = "WfmFit"): ...
getMaxPos signature(object = "WfmFit"): ...
getMinPos signature(object = "WfmFit"): ...
getNoLevels signature(object = "WfmFit"): ...
getNoProbes signature(object = "WfmFit"): ...
getPhenoInfo signature(object = "WfmFit"): ...
getPrior signature(object = "WfmFit"): ...
getProbePosition signature(object = "WfmFit"): ...
getSmoothPar signature(object = "WfmFit"): ...
getStrand signature(object = "WfmFit"): ...
getVarBetaWav signature(object = "WfmFit"): ...
getVarEps signature(object = "WfmFit"): ...
```

WfmFit-class 21

```
getVarF signature(object = "WfmFit"): ...
getWaveletFilter signature(object = "WfmFit"): ...
initialize signature(.Object = "WfmFit"): ...
show signature(object = "WfmFit"): ...
wfm.inference signature(object = "WfmFit"): ...
```

Accessors

In the following code snippets, x is a WfmFit object.

```
getBetaWav(x): Extract the fitted effect functions in the wavelet space.
getChromsome(x): Extract the chromosome identifiers.
getDataOrigSpace(x): Extract the raw expression data in the original data space.
getDataWaveletSpace(x): Extract the raw data in the wavelet space, i.e. the wavelet coefficients.
getDesignMatrix(x): Extract the design matrix used in the wavelet-based analysis.
getF(x): Extract the fitted functional effects in the original data space.
getGenomeInfo(x): Extract the genomic information.
getKj(x): Extract the number of wavelet coefficients estimated per wavelet level.
getMaxPos(x): Extract the maximum genomic probe position.
getMinPos(x): Extract the minimum genomic probe position.
getNoLevels(x): Extract the number of levels in in the wavelet decomposition when fitting the wavelet-b
getNoProbes(x): Extract the number of probes.
getPhenoInfo(x): Extract the phenotypic information for the tiling array experiment.
getPrior(x): Extract the type or distribution of the prior imposed on the functional effects in the
getProbePosition(x): Extract probe position.
getSmoothPar(x): Extract the estimated smoothing parameters that control the regularization of the eff
getStrand(x): Extract the strand orientation info.
getVarBetaWav(x): Extract the variance of the fitted effect functions in the wavelet space.
```

getVarEps(x): Extract the estimated residual variance in the wavelet space. One variance parameter is e

getWaveletFilter(x): Extract the wavelet filter used to transform the data from the original space to t

getVarF(x): Extract the variance of the fitted functional effects in the original data space.

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

22 WfmFitCircadian-class

Examples

```
showClass("WfmFit")
library(waveTilingData)
data(leafdevFit)
tt1 <- getBetaWav(leafdevFit)</pre>
tt2 <- getChromosome(leafdevFit)</pre>
tt3 <- getDataOrigSpace(leafdevFit)</pre>
tt4 <- getDataWaveletSpace(leafdevFit)</pre>
tt5 <- getDesignMatrix(leafdevFit)</pre>
tt6 <- getF(leafdevFit)</pre>
tt7 <- getGenomeInfo(leafdevFit)</pre>
tt8 <- getKj(leafdevFit)</pre>
tt9 <- getMaxPos(leafdevFit)</pre>
tt10 <- getMinPos(leafdevFit)</pre>
tt11 <- getNoLevels(leafdevFit)</pre>
tt12 <- getNoProbes(leafdevFit)
tt13 <- getPhenoInfo(leafdevFit)
tt14 <- getPrior(leafdevFit)</pre>
tt15 <- getProbePosition(leafdevFit)</pre>
tt16 <- getSmoothPar(leafdevFit)</pre>
tt17 <- getStrand(leafdevFit)
tt18 <- getVarBetaWav(leafdevFit)
tt19 <- getVarEps(leafdevFit)</pre>
tt20 <- getVarF(leafdevFit)
tt21 <- getWaveletFilter(leafdevFit)
```

WfmFitCircadian-class Class "WfmFitCircadian"

Description

class to store model fits with a circadian rhythm design in the wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmFitCircadian").

Slots

```
betaWav: Object of class "matrix" ~~
varbetaWav: Object of class "matrix" ~~
smoothPar: Object of class "matrix" ~~
varEps: Object of class "numeric" ~~
dataOrigSpace: Object of class "matrix" ~~
dataWaveletSpace: Object of class "matrix" ~~
design.matrix: Object of class "matrix" ~~
```

WfmFitCustom-class 23

```
phenoData: Object of class "data.frame" ~~
    genome.info: Object of class "genomeInfo" ~~
    n.levels: Object of class "numeric" ~~
    probePosition: Object of class "numeric" ~~
    wave.filt: Object of class "character" ~~
    Kj: Object of class "numeric" ~~
    prior: Object of class "character" ~~
    F: Object of class "matrix" ~~
    varF: Object of class "matrix" ~~
    P: Object of class "numeric" ~~
    Z: Object of class "matrix" ~~
    noGroups: Object of class "numeric" ~~
    replics: Object of class "numeric" ~~
Extends
    Class "WfmFit", directly.
Methods
    initialize signature(.Object = "WfmFitCircadian"): ...
    show signature(object = "WfmFitCircadian"): ...
Author(s)
    Kristof De Beuf <kristof.debeuf@ugent.be>
Examples
    showClass("WfmFitCircadian")
```

Description

class to store model fits with custom design in the wavelet-based transcriptome analysis.

Class "WfmFitCustom"

Objects from the Class

WfmFitCustom-class

Objects can be created by calls of the form new("WfmFitCustom").

24 WfmFitCustom-class

Slots

```
betaWav: Object of class "matrix" ~~
varbetaWav: Object of class "matrix" ~~
smoothPar: Object of class "matrix" ~~
varEps: Object of class "numeric" ~~
dataOrigSpace: Object of class "matrix" ~~
dataWaveletSpace: Object of class "matrix" ~~
design.matrix: Object of class "matrix" ~~
phenoData: Object of class "data.frame" ~~
genome.info: Object of class "genomeInfo" ~~
n.levels: Object of class "numeric" ~~
probePosition: Object of class "numeric" ~~
wave.filt: Object of class "character" ~~
Kj: Object of class "numeric" ~~
prior: Object of class "character" ~~
F: Object of class "matrix" ~~
varF: Object of class "matrix" ~~
P: Object of class "numeric" ~~
Z: Object of class "matrix" ~~
noGroups: Object of class "numeric" ~~
replics: Object of class "numeric" ~~
```

Extends

```
Class "WfmFit", directly.
```

Methods

```
initialize signature(.Object = "WfmFitCustom"): ...
show signature(object = "WfmFitCustom"): ...
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmFitCustom")
```

WfmFitFactor-class 25

WfmFitFactor-class

Class "WfmFitFactor"

Description

class to store model fits with factorial design in the wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmFitFactor").

Slots

```
betaWav: Object of class "matrix" ~~
varbetaWav: Object of class "matrix" ~~
smoothPar: Object of class "matrix" ~~
varEps: Object of class "numeric" ~~
dataOrigSpace: Object of class "matrix" ~~
dataWaveletSpace: Object of class "matrix" ~~
design.matrix: Object of class "matrix" ~~
phenoData: Object of class "data.frame" ~~
genome.info: Object of class "genomeInfo" ~~
n.levels: Object of class "numeric" ~~
probePosition: Object of class "numeric" ~~
wave.filt: Object of class "character" ~~
Kj: Object of class "numeric" ~~
prior: Object of class "character" ~~
F: Object of class "matrix" ~~
varF: Object of class "matrix" ~~
P: Object of class "numeric" ~~
Z: Object of class "matrix" ~~
noGroups: Object of class "numeric" ~~
replics: Object of class "numeric" ~~
```

Extends

```
Class "WfmFit", directly.
```

Methods

```
initialize signature(.Object = "WfmFitFactor"): ...
show signature(object = "WfmFitFactor"): ...
```

26 WfmFitTime-class

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmFitFactor")
```

WfmFitTime-class

Class "WfmFitTime"

Description

class to store model fits with a time-course design in the wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmFitTime").

Slots

```
betaWav: Object of class "matrix" ~~
varbetaWav: Object of class "matrix" ~~
smoothPar: Object of class "matrix" ~~
varEps: Object of class "numeric" ~~
dataOrigSpace: Object of class "matrix" ~~
dataWaveletSpace: Object of class "matrix" ~~
design.matrix: Object of class "matrix" ~~
phenoData: Object of class "data.frame" ~~
genome.info: Object of class "genomeInfo" ~~
n.levels: Object of class "numeric" ~~
probePosition: Object of class "numeric" ~~
wave.filt: Object of class "character" ~~
Kj: Object of class "numeric" ~~
prior: Object of class "character" ~~
F: Object of class "matrix" ~~
varF: Object of class "matrix" ~~
P: Object of class "numeric" ~~
Z: Object of class "matrix" ~~
noGroups: Object of class "numeric" ~~
replics: Object of class "numeric" ~~
```

WfmInf-class 27

Extends

```
Class "WfmFit", directly.
```

Methods

```
initialize signature(.Object = "WfmFitTime"): ...
show signature(object = "WfmFitTime"): ...
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmFitTime")
```

WfmInf-class

Class "WfmInf"

Description

class to store outputs from the inference in the wavelet-based transcriptome analysis.

Usage

```
## Accessors

getAlpha(object)
getDelta(object)
getTwoSided(object)
getSigProbes(object)
getRegions(object)
getGenomicRegions(object)
getFDR(object)
getEff(object)
getVarEff(object)
```

Arguments

object

An instance of WfmInf-class.

Objects from the Class

Objects can be created by calls of the form new ("WfmInf", alpha, delta, two.sided, sigProbes, regions, GlocRegi

28 WfmInf-class

Slots

```
alpha: Object of class "numeric" ~~

delta: Object of class "numeric" ~~

two.sided: Object of class "numeric" ~~

sigProbes: Object of class "list" ~~

regions: Object of class "list" ~~

GlocRegions: Object of class "list" ~~

FDR: Object of class "matrix" ~~

CI: Object of class "array" ~~

eff: Object of class "matrix" ~~

varEff: Object of class "matrix" ~~

genome.info: Object of class "genomeInfo" ~~
```

Methods

```
getAlpha signature(object = "WfmInf"): ...
getTwoSided signature(object = "WfmInf"): ...
getSigProbes signature(object = "WfmInf"): ...
getRegions signature(object = "WfmInf"): ...
getGenomicRegions signature(object = "WfmInf"): ...
getFDR signature(object = "WfmInf"): ...
getEff signature(object = "WfmInf"): ...
getVarEff signature(object = "WfmInf"): ...
getGenomeInfo signature(object = "WfmInf"): ...
initialize signature(.0bject = "WfmInf"): ...
show signature(object = "WfmInf"): ...
plotWfm signature(fit = "WfmInf"): ...
getSigGenes signature(fit = "WfmFit", inf = "WfmInf"): ...
getNonAnnotatedRegions signature(fit = "WfmFit", inf = "WfmInf"): ...
```

Accessors

In the following code snippets, x is a WfmInf object.

```
getAlpha(x): Extract the alpha level of significance used in the wavelet-based analysis.

getDelta(x): Extract the threshold values used in the wavelet-based transcriptome analysis.

getTwoSided(x): Extract the direction of inference conducted in the wavelet-based transcriptome analysis getSigProbes(x): Extract the significant probe ids for the wavelet-based transcriptome analysis.

getRegions(x): Extract the significant regions from the wavelet-based transcriptome analysis. Regions
```

WfmInfCompare-class 29

```
getGenomicRegions(x): Extract the significant genomic regions from the wavelet-based transcriptome and getFDR(x): Extract the FDR for each test in the wavelet-based transcriptome analysis. getEff(x): Extract the estimated effects or contrasts of the wavelet-based transcriptome analysis. getVarEff(x): Extract the estimated variances of the effects or contrasts in the wavelet-based transcriptome getGenomeInfo(x): Extract the genomic information.
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmInf")

library(waveTilingData)
data(leafdevInfCompare)
tt1 <- getAlpha(leafdevInfCompare)
tt2 <- getDelta(leafdevInfCompare)
tt3 <- getTwoSided(leafdevInfCompare)
tt4 <- getSigProbes(leafdevInfCompare)
tt5 <- getRegions(leafdevInfCompare)
tt6 <- getGenomicRegions(leafdevInfCompare)
tt7 <- getFDR(leafdevInfCompare)
tt8 <- getEff(leafdevInfCompare)
tt9 <- getVarEff(leafdevInfCompare)
tt10 <- getGenomeInfo(leafdevInfCompare)</pre>
```

WfmInfCompare-class Class "WfmInfCompare"

Description

class to store outputs from the inference for a pairwise comparison wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmInfCompare").

Slots

```
alpha: Object of class "numeric" ~~

delta: Object of class "numeric" ~~

two.sided: Object of class "numeric" ~~

sigProbes: Object of class "list" ~~

regions: Object of class "list" ~~
```

30 WfmInfCustom-class

```
GlocRegions: Object of class "list" ~~
FDR: Object of class "matrix" ~~
CI: Object of class "array" ~~
eff: Object of class "matrix" ~~
varEff: Object of class "matrix" ~~
```

Extends

```
Class "WfmInf", directly.
```

Methods

```
initialize signature(.Object = "WfmInfCompare"): ...
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmInfCompare")
```

WfmInfCustom-class

Class "WfmInfCustom"

Description

class to store outputs from the inference for a custom design fit in the wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmInfCustom").

Slots

```
alpha: Object of class "numeric" ~~

delta: Object of class "numeric" ~~

two.sided: Object of class "numeric" ~~

sigProbes: Object of class "list" ~~

regions: Object of class "list" ~~

GlocRegions: Object of class "list" ~~

FDR: Object of class "matrix" ~~

CI: Object of class "array" ~~

eff: Object of class "matrix" ~~

varEff: Object of class "matrix" ~~
```

WfmInfEffects-class 31

Extends

```
Class "WfmInf", directly.
```

Methods

```
initialize signature(.Object = "WfmInfCustom"): ...
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmInfCustom")
```

WfmInfEffects-class

Class "WfmInfEffects"

Description

class to store outputs from the inference on the effects in the wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmInfEffects").

Slots

```
alpha: Object of class "numeric" ~~

delta: Object of class "numeric" ~~

two.sided: Object of class "numeric" ~~

sigProbes: Object of class "list" ~~

regions: Object of class "list" ~~

GlocRegions: Object of class "list" ~~

FDR: Object of class "matrix" ~~

CI: Object of class "array" ~~

eff: Object of class "matrix" ~~

varEff: Object of class "matrix" ~~
```

Extends

```
Class "WfmInf", directly.
```

32 WfmInfMeans-class

Methods

```
initialize signature(.Object = "WfmInfEffects"): ...
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmInfEffects")
```

WfmInfMeans-class

Class "WfmInfMeans"

Description

class to store outputs from the inference on the group-wise means in the wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmInfMeans").

Slots

```
alpha: Object of class "numeric" ~~

delta: Object of class "numeric" ~~

two.sided: Object of class "numeric" ~~

sigProbes: Object of class "list" ~~

regions: Object of class "list" ~~

GlocRegions: Object of class "list" ~~

FDR: Object of class "matrix" ~~

CI: Object of class "array" ~~

eff: Object of class "matrix" ~~

varEff: Object of class "matrix" ~~
```

Extends

```
Class "WfmInf", directly.
```

Methods

```
initialize signature(.Object = "WfmInfMeans"): ...
```

WfmInfOverallMean-class 33

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

```
showClass("WfmInfMeans")
```

WfmInfOverallMean-class

Class "WfmInfOverallMean"

Description

class to store outputs from the inference on the overall mean expression in the wavelet-based transcriptome analysis.

Objects from the Class

Objects can be created by calls of the form new("WfmInfOverallMean").

Slots

```
alpha: Object of class "numeric" ~~

delta: Object of class "numeric" ~~

two.sided: Object of class "numeric" ~~

sigProbes: Object of class "list" ~~

regions: Object of class "list" ~~

GlocRegions: Object of class "list" ~~

FDR: Object of class "matrix" ~~

CI: Object of class "array" ~~

eff: Object of class "matrix" ~~

varEff: Object of class "matrix" ~~
```

Extends

```
Class "WfmInf", directly.
```

Methods

```
initialize signature(.Object = "WfmInfOverallMean"): ...
```

Author(s)

Kristof De Beuf <kristof.debeuf@ugent.be>

Examples

34

showClass("WfmInfOverallMean")

Index

*Topic classes	14
<pre>GenomeInfo-class, 6</pre>	
MapFilterProbe-class,9	${\sf cel2TilingFeatureSet,4}$
WaveTilingFeatureSet-class, 14	
WfmFit-class, 19	eSet, <i>15</i>
WfmFitCircadian-class, 22	
WfmFitCustom-class, 23	FeatureSet, 15
WfmFitFactor-class, 25	filterOverlap, 5
WfmFitTime-class, 26	filter Overlap, Wave Tiling Feature Set-method
WfmInf-class, 27	(WaveTilingFeatureSet-class),
WfmInfCompare-class, 29	14
WfmInfCustom-class, 30	
WfmInfEffects-class, 31	<pre>GenomeInfo (GenomeInfo-class), 6</pre>
WfmInfMeans-class, 32	GenomeInfo-class, 6
WfmInfOverallMean-class, 33	<pre>getAlpha(WfmInf-class), 27</pre>
*Topic hplot	<pre>getAlpha,WfmInf-method(WfmInf-class),</pre>
plotWfm, 11	27
*Topic manip	getBetaWav (WfmFit-class), 19
addPheno, 2	getBetaWav,WfmFit-method
bgCorrQn, 3	(WfmFit-class), 19
<pre>cel2TilingFeatureSet, 4</pre>	<pre>getChromosome (MapFilterProbe-class), 9</pre>
filterOverlap, 5	<pre>getChromosome,MapFilterProbe-method</pre>
getNonAnnotatedRegions, 6	(MapFilterProbe-class), 9
<pre>getSigGenes, 7</pre>	<pre>getChromosome,WfmFit-method</pre>
makeContrasts, 8	(WfmFit-class), 19
makeDesign, 9	<pre>getDataOrigSpace (WfmFit-class), 19</pre>
selectProbesFromFilterOverlap, 12	<pre>getDataOrigSpace,WfmFit-method</pre>
<pre>selectProbesFromTilingFeatureSet,</pre>	(WfmFit-class), 19
13	<pre>getDataWaveletSpace (WfmFit-class), 19</pre>
wfm.fit, 16	<pre>getDataWaveletSpace,WfmFit-method</pre>
wfm.inference, 17	(WfmFit-class), 19
	<pre>getDelta(WfmInf-class), 27</pre>
addPheno, 2	<pre>getDelta,WfmInf-method(WfmInf-class),</pre>
addPheno,WaveTilingFeatureSet-method	27
<pre>(WaveTilingFeatureSet-class),</pre>	<pre>getDesignMatrix(WfmFit-class), 19</pre>
14	getDesignMatrix,WfmFit-method
	(WfmFit-class), 19
ogCorrQn, 3	<pre>getEff (WfmInf-class), 27</pre>
ogCorrQn,WaveTilingFeatureSet-method	<pre>getEff,WfmInf-method(WfmInf-class), 27</pre>
<pre>(WaveTilingFeatureSet-class),</pre>	getF (WfmFit-class), 19

36 INDEX

<pre>getF,WfmFit-method(WfmFit-class), 19</pre>	getPosition,MapFilterProbe-method
<pre>getFDR (WfmInf-class), 27</pre>	(MapFilterProbe-class), 9
<pre>getFDR,WfmInf-method(WfmInf-class), 27</pre>	getPrior(WfmFit-class), 19
getFilteredIndices	<pre>getPrior,WfmFit-method(WfmFit-class),</pre>
(MapFilterProbe-class), 9	19
<pre>getFilteredIndices,MapFilterProbe-method</pre>	<pre>getProbePosition(WfmFit-class), 19</pre>
(MapFilterProbe-class), 9	<pre>getProbePosition,WfmFit-method</pre>
<pre>getGenomeInfo (WfmFit-class), 19</pre>	(WfmFit-class), 19
<pre>getGenomeInfo,WfmFit-method</pre>	<pre>getRegions (WfmInf-class), 27</pre>
(WfmFit-class), 19	<pre>getRegions,WfmInf-method</pre>
<pre>getGenomeInfo,WfmInf-method</pre>	(WfmInf-class), 27
(WfmInf-class), 27	getReplics
<pre>getGenomicRegions (WfmInf-class), 27</pre>	<pre>(WaveTilingFeatureSet-class),</pre>
getGenomicRegions, WfmInf-method	14
(WfmInf-class), 27	<pre>getReplics,WaveTilingFeatureSet-method</pre>
getGroupNames	<pre>(WaveTilingFeatureSet-class),</pre>
(WaveTilingFeatureSet-class),	14
14	getSigGenes, 7
<pre>getGroupNames,WaveTilingFeatureSet-method</pre>	<pre>getSigGenes,WfmFit,WfmInf-method</pre>
(WaveTilingFeatureSet-class),	(WfmInf-class), 27
14	<pre>getSigProbes (WfmInf-class), 27</pre>
<pre>getKj (WfmFit-class), 19</pre>	getSigProbes,WfmInf-method
<pre>getKj,WfmFit-method(WfmFit-class), 19</pre>	(WfmInf-class), 27
getMaxPos (WfmFit-class), 19	<pre>getSmoothPar (WfmFit-class), 19</pre>
<pre>getMaxPos,WfmFit-method(WfmFit-class),</pre>	getSmoothPar,WfmFit-method
19	(WfmFit-class), 19
<pre>getMinPos (WfmFit-class), 19</pre>	<pre>getStrand (MapFilterProbe-class), 9</pre>
<pre>getMinPos,WfmFit-method(WfmFit-class),</pre>	getStrand,MapFilterProbe-method
19	(MapFilterProbe-class), 9
getNoGroups	<pre>getStrand,WfmFit-method(WfmFit-class),</pre>
(WaveTilingFeatureSet-class),	19
14	<pre>getTwoSided (WfmInf-class), 27</pre>
getNoGroups,WaveTilingFeatureSet-method	getTwoSided,WfmInf-method
(WaveTilingFeatureSet-class),	(WfmInf-class), 27
14	getVarBetaWav (WfmFit-class), 19
getNoLevels (WfmFit-class), 19	getVarBetaWav,WfmFit-method
getNoLevels, WfmFit-method	(WfmFit-class), 19
(WfmFit-class), 19	getVarEff (WfmInf-class), 27
getNonAnnotatedRegions, 6	<pre>getVarEff, WfmInf-method (WfmInf-class),</pre>
getNonAnnotatedRegions, WfmFit, WfmInf-method	27
(WfmInf-class), 27	getVarEps (WfmFit-class), 19
getNoProbes (WfmFit-class), 19	getVarEps, WfmFit-method (WfmFit-class),
getNoProbes, WfmFit-method	19
(WfmFit-class), 19	getVarF (WfmFit-class), 19
getPhenoInfo (WfmFit-class), 19	getVarF, WfmFit-method (WfmFit-class), 19
getPhenoInfo,WfmFit-method	getWaveletFilter (WfmFit-class), 19
(WfmFit-class), 19	getWaveletFilter(WillFit-Class), 19 getWaveletFilter, WfmFit-method
getPosition (MapFilterProbe-class), 9	(WfmFit-class). 19
ECT OSTITUITION TITELLIONE CIOSSI'S	[HIIII IL CIGSS], 17

INDEX 37

<pre>initialize,GenomeInfo-method</pre>	show,MapFilterProbe-method
(GenomeInfo-class), 6	(MapFilterProbe-class), 9
initialize, MapFilterProbe-method	show, WaveTilingFeatureSet-method
(MapFilterProbe-class), 9	(WaveTilingFeatureSet-class),
<pre>initialize,WaveTilingFeatureSet-method</pre>	14
<pre>(WaveTilingFeatureSet-class),</pre>	<pre>show,WfmFit-method(WfmFit-class), 19</pre>
14	show, WfmFitCircadian-method
initialize,WfmFit-method	(WfmFitCircadian-class), 22
(WfmFit-class), 19	show, WfmFitCustom-method
initialize, WfmFitCircadian-method	(WfmFitCustom-class), 23
(WfmFitCircadian-class), 22	show, WfmFitFactor-method
initialize,WfmFitCustom-method	(WfmFitFactor-class), 25
(WfmFitCustom-class), 23	show, WfmFitTime-method
initialize, WfmFitFactor-method	(WfmFitTime-class), 26
(WfmFitFactor-class), 25	show, WfmInf-method (WfmInf-class), 27
initialize, WfmFitTime-method	show, WfmInfCompare-method
(WfmFitTime-class), 26	(WfmInfCompare-class), 29
initialize, WfmInf-method	show, WfmInfCustom-method
(WfmInf-class), 27	(WfmInfCustom-class), 30
initialize, WfmInfCompare-method	show, WfmInfEffects-method
(WfmInfCompare-class), 29	(WfmInfEffects-class), 31
initialize, WfmInfCustom-method	show, WfmInfMeans-method
(WfmInfCustom-class), 30	(WfmInfMeans-class), 32
initialize, WfmInfEffects-method	show, WfmInfOverallMean-method
(WfmInfEffects-class), 31	(WfmInfOverallMean-class), 33
initialize, WfmInfMeans-method	(
(WfmInfMeans-class), 32	TilingFeatureSet, 15
initialize, WfmInfOverallMean-method	-
(WfmInfOverallMean-class), 33	Versioned, 15
	VersionedBiobase, 15
makeContrasts, 8	
makeDesign, 9	WaveTilingFeatureSet
MapFilterProbe (MapFilterProbe-class), 9	(WaveTilingFeatureSet-class),
MapFilterProbe-class, 9	14
NChannal Cat 15	WaveTilingFeatureSet-class, 14
NChannelSet, 15	wfm.fit, 16
plotWfm, 11	wfm.fit,WaveTilingFeatureSet-method
plotWfm,WfmFit,WfmInf-method	(WaveTilingFeatureSet-class),
(WfmInf-class), 27	14
(wfm.inference, 17
selectProbesFromFilterOverlap, 12	wfm.inference,WfmFit-method
select Probes From Filter Overlap, Map Filter Probe	-method (WfmFit-class), 19
(MapFilterProbe-class), 9	WfmFit, $23-25$, 27
selectProbesFromTilingFeatureSet, 13	WfmFit (WfmFit-class), 19
${\tt selectProbesFromTilingFeatureSet,WaveTilingF}$	elatorieseclass, bol
(WaveTilingFeatureSet-class),	WfmFitCircadian
14	(WfmFitCircadian-class), 22
<pre>show,GenomeInfo-method</pre>	WfmFitCircadian-class, 22
(GenomeInfo-class), 6	<pre>WfmFitCustom (WfmFitCustom-class), 23</pre>

38 INDEX

```
WfmFitCustom-class, 23
WfmFitFactor (WfmFitFactor-class), 25
WfmFitFactor-class, 25
WfmFitTime (WfmFitTime-class), 26
WfmFitTime-class, 26
\texttt{WfmInf}, 30 \!\!-\!\! 33
WfmInf (WfmInf-class), 27
WfmInf-class, 27
WfmInfCompare (WfmInfCompare-class), 29
WfmInfCompare-class, 29
{\tt WfmInfCustom}\,({\tt WfmInfCustom-class}),\,30
WfmInfCustom-class, 30
WfmInfEffects (WfmInfEffects-class), 31
WfmInfEffects-class, 31
WfmInfMeans (WfmInfMeans-class), 32
WfmInfMeans-class, 32
WfmInfOverallMean
        (WfmInfOverallMean-class), 33
WfmInfOverallMean-class, 33
```