# Package 'DegNorm'

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Description This package performs degradation normalization in bulk RNA-seq data to improve dif-
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License LGPL (>= 3)
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## **Description**

DegNorm is an R package for degradation normalication for bulk RNA-seq data. DegNorm, short for degradation normalization, is a bioinformatics pipeline designed to correct for bias due to the heterogeneous patterns of transcript degradation in RNA-seq data.

## **Details**

DegNorm is a data-driven approach for RNA-Seq normalization resulting in the adjusted read count matrix. This adjustment applies to each gene within each sample, accounting for sample- and gene-specific degradation bias while simultaneously controlling for the sequencing depth. The algorithm at the center of DegNorm is the rank-one over-approximation of a gene's coverage score matrix, which is comprised of the different samples' coverage score curves along the transcript for each gene. For each gene, DegNorm estimates (1) an envelope function representing the ideal shape of the gene's coverage curve when no degradation is present, and (2) scale factors for each sample (for said gene) that indicates the relative abundance of the gene within the sample.

functions: read\_coverage\_batch,degnorm,plot\_coverage,plot\_heatmap,plot\_corr,plot\_boxplot

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#### References

DegNorm reference:

Xiong, B., Yang, Y., Fineis, F. Wang, J.-P., DegNorm: normalization of generalized transcript degradation improves accuracy in RNA-seq analysis, Genome Biology, 2019,20:75

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verage_res_chr21 Example CoverageClass data
---------------------------------------------

## **Description**

Example of CoverageClass data from DegNorm package. It is the output from read\_coverage\_batch function for human chromosome 21.

## Usage

```
data(coverage_res_chr21)
```

#### **Format**

A coverageClass list of the following

coverage a list of converage matrices for all genes within each sample counts a data.frame of read counts for all genes within each sample.

## **Examples**

```
data(coverage_res_chr21)
summary_CoverageClass(coverage_res_chr21)
```

degnorm

Main function to perform degradation normalization.

## **Description**

degnorm calcualtes the degradation index score for each gene within each sample and return the degradation-normalized read counts.

#### Usage

```
degnorm(read_coverage,counts,iteration,loop,down_sampling=1,grid_size=10,
cores=1)
```

# Arguments

read\_coverage a list of converage matrices, one per gene

counts dataframe of read counts, each row for one gene, and column for sample. The

order and number of genes must match the order in read\_coverage matrices.

iteration iteration number for degnorm algorithm. 5 is sufficient.

loop iteration number inside of nonnegative matrix factorization-over approximation.

Default is 100.

down\_sampling 1 for yes (default) and 0 for no. If yes, average coverage score is calcualted on a

grid of size specified by grid\_size argument. The new coverage matrix formed by the grid average score will be used for baseline selection. This increases the

efficiency of algorithm while maintaining comparable accuracy.

grid\_size default size is 10 bp.

cores number of cores. Default number if 1. Users should input the maximum possible

number of cores for efficiency.

#### Value

degnorm outputs a list of following objects:

counts a data.drame of read counts for each gene within each sample.

counts\_normed a data.drame of degradation-normalized read counts for each gene within each

sample.

DI a matrix of degradation index scores for each gene within each sample.

K normalizing scale factor for each gene within each sample after accounting for

degradation normalization.

convergence convergence tag; 0 = degnorm was not done on this gene because smaller counts

or too short length.1 = degnorm was done with baseline selection. 2 = degnorm done without baseline selection because gene length (after filtering out low count regions)<200 bp. 3= baseline was found, but DI score is too large. 4 = baseline

selection didn't coverge.

envelop list of the envelop curves for all genes.

## **Examples**

DegNorm-plot-functions

Degradation index (DI) score plot functions

## **Description**

DegNorm provides three functions for visualization gene-/sample-wise degradation.

## Usage

```
plot_corr(DI)
plot_heatmap(DI)
plot_boxplot(DI)
```

# Arguments

DI a matrix or data.frame of degradation index (DI) scores with each row corre-

sponding to one gene and each column for a sample.

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#### **Details**

plot\_corr plots the correlation matrix of DI scores between samples. plot\_heatmap plots the heatmap of DI scores. Left is ploted in descending order of average DI scores of genes where each row corresponds to one gene. In the right plot, DI scores were sorted within each sample and plotted in descending order. plot\_boxplotplots the boxplot of DI scores by samples.

#### Value

These functions return a boxplot of DI scores by sample, a heatmap of DIS scores of all genes in all samples and a correlation plot of DI scores between samples respectively.

## **Examples**

```
## res_DegNorm_chr21 is degnorm otuput stored in sysdata.Rda
data(res_DegNorm_chr21)
plot_boxplot(res_DegNorm_chr21$DI)
plot_heatmap(res_DegNorm_chr21$DI)
plot_corr(res_DegNorm_chr21$DI)
```

plot\_coverage

Coverage plot functions for DegNorm

#### **Description**

plot\_coverage plots the before- and after-degradation coverage curves

## Usage

```
plot_coverage(gene_name, coverage_output, degnorm_output, group=NULL, samples=NULL)
```

## **Arguments**

gene\_name the name of the gene whose coverage to be plotted.

coverage\_output

CoverageClass object, the output from function coverage\_cal\_batch.

degnorm\_output DegNormClass object, the output from function DegNorm.

group a vector of integers or character strings indicating the biological conditions of

the samples. Coverage curves will be plotted in the same color for the same group. Default is NULL. By default all curves will plotted in different colors.

samples a string vector for the subset of samples to be plotted. NULL means all samples

to be plotted. The length of samples must be of the same length of group if both

specified.

#### **Details**

plot\_coverageoutputs the coverage curves before- and after-degradation normalization.

#### Value

The coverage curve before and after degradation normalization.

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#### **Examples**

```
## gene named "SOD1",plot coverage curves
data(coverage_res_chr21)
data(res_DegNorm_chr21)
plot_coverage(gene_name="SOD1", coverage_output=coverage_res_chr21,
degnorm_output=res_DegNorm_chr21, group=c(0,1,1))
```

read\_coverage

Function to calculate read coverage score for one bam file

#### **Description**

This function judges whether bam file is single-end and paired-end, and generate bam file index if needed. It calls function paired\_end\_cov\_by\_ch or single\_end\_by\_ch. It takes multiple-core structure for parallel computing for efficiency.

# Usage

```
read_coverage(bam_file,all_genes,cores)
```

#### **Arguments**

bam\_file The name of the bam file.

all\_genes An GRangesList object. It's the parsed genes annotation file from GTF file.

cores number of cores to use.

#### **Details**

This function judges whether bam file is single-end and paired-end, and generate bam file index if needed. It takes multiple-core structure for parallel computing for efficiency.

#### Value

This function returns a coverageClass object. It contains a list of: (1) a list of coverage score for each gene in RLE format and (2) a dataframe for read counts

## See Also

```
read_coverage_batch
```

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#### **Description**

This function calls read\_coverage to compute read coverage score and read counts for all genes and samples.

#### Notes:

- 1. Coverage score is calculated per gene, i.e., concatenation of all exons from the same gene.
- 2. We follow the HTseq protocol for counting valid reads or read pairs for each gene.
- 3. When reading the alignment file, isSecondaryAlignment is set to FALSE to avoid redundant counting.
- 4. For paired-end data, isPaired is set to TRUE. We do not recommend setting isProperPair to TRUE as some fragment lengths may exceed 200 bp.
- 5. Users can modify scanBamParam in the R code as needed.

## Usage

```
read_coverage_batch(bam_file_list, gtf_file, cores = 1)
```

## **Arguments**

```
bam_file_list A character vector of BAM file paths.

gtf_file The GTF file that RNA-seq reads were aligned to.

cores Number of processor cores to use. Default = 1.
```

# Value

A list with the following components:

coverage A list of coverage matrices for all genes within each sample.

A data. frame of read counts for all genes within each sample.

#### See Also

```
read_coverage
```

## **Examples**

```
## Read BAM file and GTF file from the package
bam_file_list <- list.files(
    path = system.file("extdata", package = "DegNorm"),
    pattern = ".bam$", full.names = TRUE
)
gtf_file <- list.files(
    path = system.file("extdata", package = "DegNorm"),
    pattern = ".gtf$", full.names = TRUE
)

## Run read_coverage_batch to calculate read coverage curves and read counts
coverage_res <- read_coverage_batch(bam_file_list, gtf_file, cores = 2)</pre>
```

res\_DegNorm\_chr21

Example DegNormClass data

#### **Description**

Example of DegNormClass data from DegNorm package. It is the output from degnorm function for human chromosome 21.

## Usage

```
data("res_DegNorm_chr21")
```

#### **Format**

A DegNormClass list of the following items:

counts a data.drame of read counts for each gene within each sample.

counts\_normed a data.drame of degradation-normalized read counts for each gene within each sample.

DI a matrix of degradation index scores for each gene within each sample.

K normalizing scale factor for each gene within each sample after accounting for degradation normalization.

convergence convergence tag; 0 = degnorm was not done on this gene because smaller counts or too short length.1 = degnorm was done with baseline selection. 2 = degnorm done without baseline selection because gene length (after filtering out low count regions)<200 bp. 3= baseline was found, but DI score is too large. 4 = baseline selection didn't coverge.

envelop a list of the envelop curves for all genes.

#### **Examples**

```
data(res_DegNorm_chr21)
summary_DegNormClass(res_DegNorm_chr21)
```

 ${\tt summary\_CoverageClass} \ \ \textit{Summary method for CoverageClass}.$ 

## Description

It prints a summary of the data objects contained in the list from  ${\tt read\_coverage\_batch}$ .

#### Usage

```
summary_CoverageClass(object)
```

#### **Arguments**

object

CoverageClass from coderead\_coverage\_batch.

## Value

On-screen plot of summary of CoverageClass object.

## **Examples**

```
## Summary of coverage_cal_batch output (CoverageClass)
data(coverage_res_chr21)
summary_CoverageClass(coverage_res_chr21)
```

 $summary\_DegNormClass \quad \textit{Summary method for DegNormClass}.$ 

# **Description**

It prints a summary of the data objects contained in the list from degnorm function.

# Usage

```
summary_DegNormClass(object)
```

## **Arguments**

object

DegNormClass from degnorm function.

## Value

On-screen summary of DegNormClass object.

## **Examples**

```
## Summary of degnorm output (DegNormlass)
data(res_DegNorm_chr21)
summary_DegNormClass(res_DegNorm_chr21)
```

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