

# Package ‘rawDiag’

July 11, 2025

**Type** Package

**Title** Brings Orbitrap Mass Spectrometry Data to Life; Fast and Colorful

**Version** 1.5.1

**Depends** R (>= 4.4)

**Imports** dplyr, ggplot2 (>= 3.4), grDevices, hexbin, htmltools, BiocManager, BiocParallel, rawrr (>= 1.15.5), rlang, reshape2, scales, shiny (>= 1.5), stats, utils

**Suggests** BiocStyle (>= 2.28), ExperimentHub, tartare, knitr, testthat

**Description** Optimizing methods for liquid chromatography coupled to mass spectrometry (LC-MS) poses a nontrivial challenge. The rawDiag package facilitates rational method optimization by generating MS operator-tailored diagnostic plots of scan-level metadata. The package is designed for use on the R shell or as a Shiny application on the Orbitrap instrument PC.

**License** GPL-3

**URL** <https://github.com/fgcz/rawDiag/>

**BugReports** <https://github.com/fgcz/rawDiag/issues>

**Encoding** UTF-8

**NeedsCompilation** no

**RoxygenNote** 7.3.2

**VignetteBuilder** knitr

**biocViews** MassSpectrometry, Proteomics, Metabolomics, Infrastructure, Software, ShinyApps

**git\_url** <https://git.bioconductor.org/packages/rawDiag>

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rawDiag-package

*rawDiag: Brings Orbitrap Mass Spectrometry Data to Life; Fast and Colorful*

## Description

Optimizing methods for liquid chromatography coupled to mass spectrometry (LC-MS) poses a nontrivial challenge. The rawDiag package facilitates rational method optimization by generating MS operator-tailored diagnostic plots of scan-level metadata. The package is designed for use on the R shell or as a Shiny application on the Orbitrap instrument PC.

## Author(s)

**Maintainer:** Christian Panse <cp@fgcz.ethz.ch> ([ORCID](#))

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- Tobias Kockmann <[Tobias.Kockmann@fgcz.ethz.ch](mailto:Tobias.Kockmann@fgcz.ethz.ch)>

## See Also

Useful links:

- <https://github.com/fgcz/rawDiag/>
- Report bugs at <https://github.com/fgcz/rawDiag/issues>

---

.calculatioMasterScan *Calculate Master Scan Number*

---

### Description

calculates the MS1 master scan number of an MS2 scan and populates the MasterScanNumber with it

### Usage

.calculatioMasterScan(x)

### Arguments

x a data.frame object adhering to the specified criteria for the `is.rawDiag` function.

### Value

a data.frame containing a MasterScanNumber column.

### Author(s)

Christian Trachsel

---

.cycleTime *Calculate MS Cycle Time*

---

### Description

calculates the lock mass deviations along RT.

### Usage

.cycleTime(x)

### Arguments

x a data.frame object adhering to the specified criteria for the `is.rawDiag` function.

### Value

calculates the time of all ms cycles and the 95 the cycle time is defined as the time between two consecutive MS1 scans

### Note

TODO: quantile part needed? If no MS1 scan is present? E.g., DIA take lowest window as cycle indicator?

**Author(s)**

Christian Trachsel (2017), Christian Panse (20231201) refactored

`.fillNAgaps`

*Fill NA values with last previous value*

**Description**

Fill NA values with last previous value

**Usage**

`.fillNAgaps(x)`

**Arguments**

`x` a vector of values

**Value**

a vector with any NA values replaced with the last previous actual value

**Author(s)**

Christian Trachsel

**Examples**

```
c(NA, 1, 2, 3, NA, 4, 5, NA, NA, NA, 6) |>
  rawDiag:::fillNAgaps()
```

`buildRawDiagShinyApp` *Build the rawDiag shiny application*

**Description**

Build the rawDiag shiny application

**Usage**

`buildRawDiagShinyApp(rawDir = (dirname(rawrr::sampleFilePath())))`

**Arguments**

`rawDir` A directory containing the input raw files, default is set to the \$HOME/Downloads directory.

**Value**

returns the rawDiag shiny apps

**Note**

launch the shiny application by embracing your command line while expecting the raw file in \$HOME/Downloads

- MacOSX and Linux: R -q -e "library(rawDiag); buildRawDiagShinyApp() |> shiny::runApp(launch.browser=TRUE)"
- Microsoft Windows: R.exe -q -e "library(rawDiag); buildRawDiagShinyApp() |> shiny::runApp(launch.browser=TRUE)"

**Author(s)**

Christian Trachsel (2017), Christian Panse (2023)

**References**

- rawDiag: [doi:10.1021/acs.jproteome.8b00173](https://doi.org/10.1021/acs.jproteome.8b00173),
- rawrr: [doi:10.1021/acs.jproteome.0c00866](https://doi.org/10.1021/acs.jproteome.0c00866)

**Examples**

```
rawrr::sampleFilePath() |>
  dirname() |>
  rawDiag::buildRawDiagShinyApp() |>
  shiny::runApp()

# or use your 'Download' folder
Sys.getenv('HOME') |>
  file.path("Downloads") |>
  rawDiag::buildRawDiagShinyApp() |>
  shiny::runApp()
```

---

checkRawrr

*Checks Bioconductor installation instructions*

---

**Description**

Checks Bioconductor installation instructions

**Usage**

```
checkRawrr()
```

**Value**

TRUE if everything is installed correctly

`is.rawDiag`*Is an Object an rawDiag Object?***Description**

Is an Object an rawDiag Object?

**Usage**

```
is.rawDiag(object)
```

**Arguments**

`object` any R object.

**Value**

a boolean

**Author(s)**

Christian Panse 2018

**Examples**

```
rawrr::sampleFilePath() |> rawDiag::readRaw() |> rawDiag::is.rawDiag()
```

`plotChargeState`*Charge State Overview Plot***Description**

graphs the number of occurrences of all selected precursor charge states.

**Usage**

```
plotChargeState(x, method = "trellis")
```

**Arguments**

`x` a `data.frame` object adhering to the specified criteria for the `is.rawDiag` function.  
`method` specifying the plot method 'trellis' | 'violin' | 'overlay'. The default is 'trellis'.

**Value**

a `ggplot` object.

**Author(s)**

Christian Trachsel (2017), Christian Panse (2023)

**References**

- rawDiag: doi:10.1021/acs.jproteome.8b00173,
- rawrr: doi:10.1021/acs.jproteome.0c00866

**Examples**

```
rawrr::sampleFilePath() |> rawDiag::readRaw() -> S  
S |> rawDiag::plotLockMassCorrection()
```

---

**plotCycleLoad**

*Cycle Load Plot*

---

**Description**

plotting the number of MS2 per MS1 (the duty cycle) scan versus retention time. The deepskyblue colored loess curve shows the trend.

**Usage**

```
plotCycleLoad(x, method = "trellis")
```

**Arguments**

- x a data.frame object adhering to the specified criteria for the `is.rawDiag` function.
- method specifying the plot method 'trellis' | 'violin' | 'overlay'. The default is 'trellis'.

**Value**

a `ggplot` object.

**Author(s)**

Christian Trachsel (2017), Christian Panse (2023)

**References**

- rawDiag: doi:10.1021/acs.jproteome.8b00173,
- rawrr: doi:10.1021/acs.jproteome.0c00866

**Examples**

```
rawrr::sampleFilePath() |> rawDiag::readRaw() -> S  
S |> rawDiag::plotCycleLoad()
```

`plotCycleTime`      *Plot Cycle Time*

## Description

graphs the time difference between two consecutive MS1 scans (cycle time) with respect to RT (scatter plots) or its density (violin). A smooth curve graphs the trend. The 95th percentile is indicated by a red dashed line.

## Usage

```
plotCycleTime(x, method = "trellis")
```

## Arguments

- |                     |   |
|---------------------|---|
| <code>x</code>      | a <code>data.frame</code> object adhering to the specified criteria for the <code>is.rawDiag</code> function. |
| <code>method</code> | specifying the plot method 'trellis'   'violin'   'overlay'. The default is 'trellis'.                        |

## Value

a `ggplot` object.

## Examples

```
rawrr::sampleFilePath() |> rawDiag::readRaw() |> rawDiag::plotCycleTime()
```

`plotInjectionTime`      *Plot Injection Time*

## Description

shows the injection time density of each mass spectrometry file as a violin plot. The higher the maximum number of MS2 scans is in the method, the more the density is shifted towards the maximum injection time value.

## Usage

```
plotInjectionTime(x, method = "trellis")
```

## Arguments

- |                     |   |
|---------------------|---|
| <code>x</code>      | a <code>data.frame</code> object adhering to the specified criteria for the <code>is.rawDiag</code> function. |
| <code>method</code> | specifying the plot method 'trellis'   'violin'   'overlay'. The default is 'trellis'.                        |

## Value

a `ggplot` object.

**Author(s)**

Christian Trachsel (2017), Christian Panse (2023)

**References**

- rawDiag: doi:10.1021/acs.jproteome.8b00173,
- rawrr: doi:10.1021/acs.jproteome.0c00866

**Examples**

```
rawrr::sampleFilePath() |> rawDiag::readRaw() |> rawDiag::plotInjectionTime()
```

---

**plotLockMassCorrection***Lock Mass Correction Plot*

---

**Description**

Lock Mass Correction Plot

**Usage**

```
plotLockMassCorrection(x, method = "trellis")
```

**Arguments**

- x a data.frame object adhering to the specified criteria for the `is.rawDiag` function.
- method specifying the plot method 'trellis' | 'violin' | 'overlay'. The default is 'trellis'.

**Value**

a `ggplot` object.

**Author(s)**

Christian Trachsel (2017), Christian Panse (2023)

**References**

- rawDiag: doi:10.1021/acs.jproteome.8b00173,
- rawrr: doi:10.1021/acs.jproteome.0c00866

**Examples**

```
rawrr::sampleFilePath() |>  
  rawDiag::readRaw() |>  
  rawDiag::plotLockMassCorrection()
```

---

**plotMassDistribution** *Mass Distribution Plot*

---

**Description**

plots the mass frequency in dependency to the charge state

**Usage**

```
plotMassDistribution(x, method = "trellis")
```

**Arguments**

- |        |   |
|--------|---|
| x      | a <code>data.frame</code> object adhering to the specified criteria for the <code>is.rawDiag</code> function. |
| method | specifying the plot method 'trellis'   'violin'   'overlay'. The default is 'trellis'.                        |

**Details**

displays charge state resolved frequency of precursor masses.

**Value**

a `ggplot` object.

**Author(s)**

Christian Trachsel (2017), Christian Panse (2023)

**References**

- rawDiag: [doi:10.1021/acs.jproteome.8b00173](https://doi.org/10.1021/acs.jproteome.8b00173),
- rawrr: [doi:10.1021/acs.jproteome.0c00866](https://doi.org/10.1021/acs.jproteome.0c00866)

**Examples**

```
rawrr::sampleFilePath() |> rawDiag::readRaw() |> rawDiag::plotMassDistribution('overlay')
```

---

**plotMzDistribution** *mZ Distribution Plot of Ms2 Scans*

---

**Description**

draws precursor mass vs retention time for each MS2 scan in the raw file.

**Usage**

```
plotMzDistribution(x, method = "trellis")
```

**Arguments**

- x a `data.frame` object adhering to the specified criteria for the `is.rawDiag` function.
- method specifying the plot method 'trellis' | 'violin' | 'overlay'. The default is 'trellis'.

**Value**

a `ggplot` object.

**Author(s)**

Christian Trachsel (2017), Christian Panse (2023)

**References**

- rawDiag: [doi:10.1021/acs.jproteome.8b00173](https://doi.org/10.1021/acs.jproteome.8b00173),
- rawrr: [doi:10.1021/acs.jproteome.0c00866](https://doi.org/10.1021/acs.jproteome.0c00866)

**Examples**

```
rawrr::sampleFilePath() |> rawDiag::readRaw() -> S
rawDiag::plotMzDistribution(S)
```

`plotPrecursorHeatmap` *Precursor Mass versus StartTime MS2 based hexagons*

**Description**

Precursor Mass versus StartTime MS2 based hexagons

**Usage**

```
plotPrecursorHeatmap(x, method = "overlay", bins = 80)
```

**Arguments**

- x a `data.frame` object adhering to the specified criteria for the `is.rawDiag` function.
- method specifying the plot method 'trellis' | 'violin' | 'overlay'. The default is 'trellis'.
- bins number of bins in both vertical and horizontal directions. default is 80.

**Value**

a `ggplot` object.

**Note**

TODO: define bin with dynamically as  $h = 2 \times \text{IQR} \times n^{1/3}$  or number of bins  $(\text{max-min})/h$

**Author(s)**

Christian Trachsel (2017)

## References

- rawDiag: doi:10.1021/acs.jproteome.8b00173,
- rawrr: doi:10.1021/acs.jproteome.0c00866

## Examples

```
rawrr::sampleFilePath() |> rawDiag::readRaw() |> rawDiag::plotPrecursorHeatmap()
```

`plotScanTime`

*Scan Event Plot*

## Description

Plotting the elapsed scan time for each individual scan event.

## Usage

```
plotScanTime(x, method = "trellis")
```

## Arguments

- |        |   |
|--------|---|
| x      | a <code>data.frame</code> object adhering to the specified criteria for the <code>is.rawDiag</code> function. |
| method | specifying the plot method 'trellis'   'violin'   'overlay'. The default is 'trellis'.                        |

## Value

a `ggplot` object.

## Author(s)

Christian Trachsel (2017), Christian Panse (2023)

## References

- rawDiag: doi:10.1021/acs.jproteome.8b00173,
- rawrr: doi:10.1021/acs.jproteome.0c00866

## Examples

```
## for debugging bioconductor check
if (Sys.info()['sysname'] %in% c("Darwin", "Linux")) {Sys.which('mono')}
rawrr::sampleFilePath() |> rawDiag::readRaw() -> S

rawDiag::checkRawrr()

S |> rawDiag::plotScanTime()
```

---

plotTicBasepeak      *Total Ion Count and Base Peak Plot*

---

**Description**

displays the Total Ion Count (TIC) and the Base Peak Chromatogram of a mass spectrometry measurement. Multiple files are handled by faceting based on rawfile name.

**Usage**

```
plotTicBasepeak(x, method = "trellis")
```

**Arguments**

- |        |   |
|--------|---|
| x      | a <code>data.frame</code> object adhering to the specified criteria for the <code>is.rawDiag</code> function. |
| method | specifying the plot method 'trellis'   'violin'   'overlay'. The default is 'trellis'.                        |

**Value**

a `ggplot2` object for graphing the TIC and the Base Peak chromatogram.

**Author(s)**

Christian Trachsel (2017), Christian Panse (20231130) refactored

**Examples**

```
rawrr::sampleFilePath() |> rawDiag::readRaw() |> rawDiag::plotTicBasepeak()
```

---

rawDiagServer      *rawDiag shiny module*

---

**Description**

rawDiag shiny module

**Usage**

```
rawDiagServer(id, vals)
```

**Arguments**

- |      |  |
|------|--|
| id   | An ID string that corresponds with the ID used to call the module's UI function. |
| vals | containing rawfile   |

**Value**

rawDiag shiny module server

**Examples**

```
shiny::shiny(rawDir = (rawrr::sampleFilePath() |> dirname()))
```

---

**rawDiagUI**

*rawDiag shiny module UI*

---

**Description**

rawDiag shiny module UI

**Usage**

```
rawDiagUI(id)
```

**Arguments**

|    |  |
|----|--|
| id | An ID string that corresponds with the ID used to call the module's UI function. |
|----|--|

**Value**

a shiny UI module

rawDiag shiny module UI

**Examples**

```
rawDiag::shiny(rawDir = (rawrr::sampleFilePath() |> dirname()))
```

---

**readRaw**

*Reads selected raw file trailer information for rawDiag plot functions*

---

**Description**

implements a wrapper function using the rawrr methods [readIndex](#), [readTrailer](#), and [readChromatogram](#) to read proprietary mass spectrometer generated data using third-party libraries.

**Usage**

```
readRaw(
  rawfile,
  msgFUN = function(x) {
    message(x)
  }
)
```

**Arguments**

|         |   |
|---------|---|
| rawfile | the name of the raw file containing the mass spectrometry data from the Thermo Fisher Scientific instrument.  |
| msgFUN  | this function is used for logging information while composing the resulting data.frame. It can also be used for shiny progress bar. The default is using the message. |

**Value**

a data.frame containing the selected trailer information.

**Note**

The set up procedure for the rawrr package needs to be run in order to use this package.

**Author(s)**

Christian Panse (2016-2023)

**References**

[doi:10.1021/acs.jproteome.8b00173](https://doi.org/10.1021/acs.jproteome.8b00173)

**Examples**

```
rawDiag::checkRawrr()  
rawrr::sampleFilePath() |>  
  rawDiag::readRaw()
```

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