

Package ‘EasyCellType’

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Title Annotate cell types for scRNA-seq data

Version 1.11.0

Description We developed EasyCellType which can automatically examine the input marker lists obtained from existing software such as Seurat over the cell markerdatabases. Two quantification approaches to annotate cell types are provided: Gene set enrichment analysis (GSEA) and a modified versio of Fisher's exact test. The function presents annotation recommendations in graphical outcomes: bar plots for each cluster showing candidate cell types, as well as a dot plot summarizing the top 5 significant annotations for each cluster.

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Depends R (>= 4.2.0)

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Contents

cellmarker_tissue	2
clustermole_tissue	2
coremarkers	3
easycyt	3
gene_pbmc	4
mapsymbol	5
panglao_tissue	5
pbmc_data	6
plot_bar	6
plot_dot	7
process_results	7
summarycelltype	8
test_fisher	9
Index	10

cellmarker_tissue	<i>Tissues in CellMarker database.</i>
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Description

A list containing 2 elements: Human tissues and Mouse tissues.

Usage

data(cellmarker_tissue)

Format

A list with 2 elements:

Human Human tissue

Mouse Mouse tissue

clustermole_tissue	<i>Tissues in Clustermole database.</i>
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Description

A list containing 2 elements: Human tissues and Mouse tissues.

Usage

data(clustermole_tissue)

Format

A list with 2 elements:

Human Human tissue

Mouse Mouse tissue

coremarkers	<i>Title Summarize markers contirbuting to the cell type annotation</i>
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Description

Title Summarize markers contirbuting to the cell type annotation

Usage

```
coremarkers(test, data, species)
```

Arguments

test	Test used to annotation cell types: "GSEA" or "fisher"
data	Annotation results.
species	"Human" or "Mouse"

Value

A data frame containing genes contributed to cell annotation

Examples

```
## core_markers <- coremarkers("GSEA", data)
```

easyct	<i>Annotate cell types for scRNA-seq data</i>
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Description

This function is used to run the annotation analysis using either GSEA or a modified Fisher's exact test. We expect users to input a data frame containing expressed markers, cluster information and the differential score (log fold change). The gene lists in that data frame should be sorted by their differential score.

Usage

```
easyct(
  data,
  db = "cellmarker",
  genotype = "Entrezid",
  species = "Human",
  tissue = NULL,
  p_cut = 0.5,
  test = "GSEA",
  scoretype = "std"
)
```

Arguments

<code>data</code>	A data frame containing the markers, cluster, and expression scores; Marker genes should be sorted in each cluster. Order of the columns should be gene, cluster and expression level score. An example data can be loaded using <code>'data(gene_pbmc)'</code> .
<code>db</code>	Name of the reference database: cellmarker, clustermole or panglaodb;
<code>genetype</code>	Indicate the gene type in the input data frame: "Entrezid" or "symbol".
<code>species</code>	Human or Mouse. Human in default.
<code>tissue</code>	Tissue types can be specified when running the analysis. Length of tissue can be larger than 1. The possible tissues can be seen using <code>'data(cellmarker_tissue)'</code> , <code>'data(clustermole_tissue)'</code> and <code>'data(panglao_tissue)'</code> .
<code>p_cut</code>	Cutoff of the P value for GSEA.
<code>test</code>	"GSEA" or "fisher"; "GSEA" is used in default.
<code>scoretype</code>	Argument used for GSEA. Default value is "std". If all scores are positive, then scoretype should be "pos".

Value

A list containing the test results for each cluster.

Examples

```
data(gene_pbmc)
result <- easyct(gene_pbmc, db="cellmarker", species="Human",
  tissue=c("Blood", "Peripheral blood", "Blood vessel",
    "Umbilical cord blood", "Venous blood"), p_cut=0.3, test="GSEA", scoretype="pos")
```

gene_pbmc

Differential expressed marker genes in 9 clusters.

Description

A data frame containing marker genes, clusters as well as the average of log 2 fold changes. The original data set is from 10X genomics, and we followed the standard workflow provided by Seurat package to process data, and then format to get the data frame.

Usage

```
data(gene_pbmc)
```

Format

A data frame with 727 rows and 3 variables:

gene Entrez IDs of the marker genes

cluster Cluster

score Average of log 2 fold changes getting from the process procedure

Source

https://cf.10xgenomics.com/samples/cell/pbmc3k/pbmc3k_filtered_gene_bc_matrices.tar.gz

mapsymbol	<i>Title Convert gene symbol to Entrez ID</i>
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Description

This function is used to convert the gene symbol to Entrez Id. Used in easyct function.

Usage

```
mapsymbol(d, species)
```

Arguments

d	A data frame where first column contains gene symbols.
species	"Human" or "Mouse".

Value

A data frame containing gene symbols and the corresponding Entrez ID

panglao_tissue	<i>Tissues in Panglao database.</i>
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Description

A list containing 2 elements: Human tissues and Mouse tissues.

Usage

```
data(panglao_tissue)
```

Format

A list with 2 elements:

Human Human tissue

Mouse Mouse tissue

pbmc_data	<i>Peripheral Blood Mononuclear Cells (PBMC) data.</i>
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Description

Count matrix of Peripheral Blood Mononuclear Cells (PBMC). The original data set is from 10X genomics.

Usage

```
data(pbmc_data)
```

Format

A large dgCMatrix: 32378 * 2700

i Row index of the non-zero values

p A vector to refer the column index of the non-zero values

Dim Dimension of the matrix

Dimnames A list of length 2 containing the row names and column names of the matrix

x Vector containing all the non-zero values

Source

https://cf.10xgenomics.com/samples/cell/pbmc3k/pbmc3k_filtered_gene_bc_matrices.tar.gz

plot_bar	<i>Create bar plots for each cluster</i>
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Description

This function is used to generate set of bar plots presenting up to 10 candidate cell types for each cluster.

Usage

```
plot_bar(test = "GSEA", data, cluster = NULL)
```

Arguments

test	"GSEA" or "fisher"
data	Annotation results
cluster	Cluster can be specified to print plots.

Value

Bar plots showing show up to 10 candidate cell types for each cluster.

Examples

```
data(gene_pbmc)
result <- easyct(gene_pbmc, db="cellmarker", species="Human",
  tissue=c("Blood", "Peripheral blood", "Blood vessel",
    "Umbilical cord blood", "Venous blood"), p_cut=0.3, test="GSEA", scoretype="pos")
plot_bar("GSEA", result)
```

plot_dot

*Create dot plot for annotation results***Description**

This function is used to generate a dot plot presenting the top 5 candidate cell types for each cluster.

Usage

```
plot_dot(test = "GSEA", data)
```

Arguments

test	Test used to annotate cell types: "GSEA" or "fisher"
data	Annotation results

Value

A dot plot showing the top 5 significant cell types for each cluster.

Examples

```
data(gene_pbmc)
result <- easyct(gene_pbmc, db="cellmarker", species="Human",
  tissue=c("Blood", "Peripheral blood", "Blood vessel",
    "Umbilical cord blood", "Venous blood"), p_cut=0.3, test="GSEA", scoretype="pos")
plot_dot("GSEA", result)
```

process_results

*Title Annotate cell types for single cell RNA data***Description**

This function is used to process the annotation test results. Processed data will be used to generate plots.

Usage

```
process_results(test, data)
```

Arguments

test	Test used to annotation cell types: "GSEA" or "fisher"
data	Annotation results.

Value

A data frame used to generate plots.

summarycelltype	<i>Print test results</i>
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Description

This function is used to print summary table of annotation results for a specific cluster.

Usage

```
summarycelltype(test, results, cluster)
```

Arguments

test	"GSEA" or "fisher".
results	Annotation results.
cluster	Cluster of interest.

Value

A summary table of a annotation results. "core_enrichment" contains markers contributing on the annotation.

Examples

```
data(gene_pbmc)
result <- easyct(gene_pbmc, db="cellmarker", species="Human",
tissue=c("Blood", "Peripheral blood", "Blood vessel",
"Umbilical cord blood", "Venous blood"), p_cut=0.3, test="GSEA", scoretype="pos")
summarycelltype(test="GSEA", results=result, cluster=0)
```

test_fisher	<i>Fisher exact test used in function 'easyct'</i>
-------------	--

Description

This function is used to conduct the modified Fisher's exact test.

Usage

```
test_fisher(testgenes, ref, cols)
```

Arguments

testgenes	A data frame containing query genes and the expression scores.
ref	The reference data base.
cols	Column names of the input data frame

Value

A data frame containg the results of fisher's exact test.

Index

* datasets

- cellmarker_tissue, [2](#)
- clustermole_tissue, [2](#)
- gene_pbmc, [4](#)
- panglao_tissue, [5](#)
- pbmc_data, [6](#)

- cellmarker_tissue, [2](#)
- clustermole_tissue, [2](#)
- coremarkers, [3](#)

- easyct, [3](#)

- gene_pbmc, [4](#)

- mapsymbol, [5](#)

- panglao_tissue, [5](#)
- pbmc_data, [6](#)
- plot_bar, [6](#)
- plot_dot, [7](#)
- process_results, [7](#)

- summarycelltype, [8](#)

- test_fisher, [9](#)